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(54) Title: COMPOSITIONS AND METHODS FOR THE THERAPY AND DIAGNOSIS OF OVARIAN CANCER

(57) Abstract: Compositions and methods for the therapy and diagnosis of cancer, particularly ovarian cancer, are disclosed. Illustrative compositions comprise one or more ovarian tumor polypeptides, immunogenic portions thereof, polynucleotides that encode such polypeptides, antigen presenting cell that expresses such polypeptides, and T cells that are specific for cells expressing such polypeptides. The disclosed compositions are useful, for example, in the diagnosis, prevention and/or treatment of diseases, particularly ovarian cancer.

COMPOSITIONS AND METHODS FOR THE THERAPY  
AND DIAGNOSIS OF OVARIAN CANCER

BACKGROUND OF THE INVENTION

5    Field of the Invention

The present invention relates generally to ovarian cancer therapy. The invention is more specifically related to polypeptides comprising at least a portion of an ovarian carcinoma protein, and to polynucleotides encoding such polypeptides, as well as antibodies and immune system cells that specifically recognize such polypeptides.

- 10   Such polypeptides, polynucleotides, antibodies and cells may be used in vaccines and pharmaceutical compositions for treatment of ovarian cancer.

Description of Related Art

- Ovarian cancer is a significant health problem for women in the United States and throughout the world. Although advances have been made in detection and therapy of this cancer, no vaccine or other universally successful method for prevention or treatment is currently available. Management of the disease currently relies on a combination of early diagnosis and aggressive treatment, which may include one or more of a variety of treatments such as surgery, radiotherapy, chemotherapy and hormone therapy. The course of treatment for a particular cancer is often selected based on a variety of prognostic parameters, including an analysis of specific tumor markers. However, the use of established markers often leads to a result that is difficult to interpret, and high mortality continues to be observed in many cancer patients.

- Immunotherapies have the potential to substantially improve cancer treatment and survival. Such therapies may involve the generation or enhancement of an immune response to an ovarian carcinoma antigen. However, to date, relatively few ovarian carcinoma antigens are known and the generation of an immune response against such antigens has not been shown to be therapeutically beneficial.

Accordingly, there is a need in the art for improved methods for identifying ovarian tumor antigens and for using such antigens in the therapy of ovarian cancer. The present invention fulfills these needs and further provides other related advantages.

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#### BRIEF SUMMARY OF THE INVENTION

Briefly stated, this invention provides compositions and methods for the therapy of cancer, such as ovarian cancer.

In one aspect, the present invention provides polynucleotide compositions comprising a sequence selected from the group consisting of:

(a) sequences provided in SEQ ID NO:1, 2, 5, 9, 10, 13, 16, 19, 23, 27, 28, 32, 33, 35, 38, 41-50, 52, 53, 56, 57, 63, 65, 69-72, 75, 78, 80-82, 84, 86, 89-93, 95, 97-100, 103, 107, 111, 114, 117, 120, 121, 125, 128, 132-134, 136, 137, 140, 143-146, 148-151, 156, 158, 160-162, 166-168, 171, 174-183, 185, 193-199, 203-206, and 208;

(b) complements of the sequences provided in SEQ ID NO:1, 2, 5, 9, 10, 13, 16, 19, 23, 27, 28, 32, 33, 35, 38, 41-50, 52, 53, 56, 57, 63, 65, 69-72, 75, 78, 80-82, 84, 86, 89-93, 95, 97-100, 103, 107, 111, 114, 117, 120, 121, 125, 128, 132-134, 136, 137, 140, 143-146, 148-151, 156, 158, 160-162, 166-168, 171, 174-183, 185, 193-199, 203-206, and 208;

(c) sequences consisting of at least 20 contiguous residues of a sequence provided in SEQ ID NO:1, 2, 5, 9, 10, 13, 16, 19, 23, 27, 28, 32, 33, 35, 38, 41-50, 52, 53, 56, 57, 63, 65, 69-72, 75, 78, 80-82, 84, 86, 89-93, 95, 97-100, 103, 107, 111, 114, 117, 120, 121, 125, 128, 132-134, 136, 137, 140, 143-146, 148-151, 156, 158, 160-162, 166-168, 171, 174-183, 185, 193-199, 203-206, and 208;

(d) sequences that hybridize to a sequence provided in SEQ ID NO:1, 2, 5, 9, 10, 13, 16, 19, 23, 27, 28, 32, 33, 35, 38, 41-50, 52, 53, 56, 57, 63, 65, 69-72, 75, 78, 80-82, 84, 86, 89-93, 95, 97-100, 103, 107, 111, 114, 117, 120, 121, 125, 128, 132-134, 136, 137, 140, 143-146, 148-151, 156, 158, 160-162, 166-168, 171, 174-183, 185, 193-199, 203-206, and 208 under moderately stringent conditions;

(e) sequences having at least 75% identity to a sequence provided in SEQ ID NO:1, 2, 5, 9, 10, 13, 16, 19, 23, 27, 28, 32, 33, 35, 38, 41-50, 52, 53, 56, 57, 63, 65, 69-72, 75, 78, 80-82, 84, 86, 89-93, 95, 97-100, 103, 107, 111, 114, 117, 120, 121, 125, 128, 132-134, 136, 137, 140, 143-146, 148-151, 156, 158, 160-162, 166-168, 5 171, 174-183, 185, 193-199, 203-206, 208, and 210-214;

(f) sequences having at least 90% identity to a sequence provided in SEQ ID NO:1, 2, 5, 9, 10, 13, 16, 19, 23, 27, 28, 32, 33, 35, 38, 41-50, 52, 53, 56, 57, 63, 65, 69-72, 75, 78, 80-82, 84, 86, 89-93, 95, 97-100, 103, 107, 111, 114, 117, 120, 121, 125, 128, 132-134, 136, 137, 140, 143-146, 148-151, 156, 158, 160-162, 166-168, 10 171, 174-183, 185, 193-199, 203-206, 208 and 210-214; and

(g) degenerate variants of a sequence provided in SEQ ID NO:1, 2, 5, 9, 10, 13, 16, 19, 23, 27, 28, 32, 33, 35, 38, 41-50, 52, 53, 56, 57, 63, 65, 69-72, 75, 78, 80-82, 84, 86, 89-93, 95, 97-100, 103, 107, 111, 114, 117, 120, 121, 125, 128, 132-134, 136, 137, 140, 143-146, 148-151, 156, 158, 160-162, 166-168, 171, 174-183, 185, 15 193-199, 203-206, 208 and 210-214.

In one preferred embodiment, the polynucleotide compositions of the invention are expressed in at least about 20%, more preferably in at least about 30%, and most preferably in at least about 50% of ovarian tumors samples tested, at a level that is at least about 2-fold, preferably at least about 5-fold, and most preferably at least 20 about 10-fold higher than that for normal tissues.

In one aspect, the present invention provides polypeptides comprising an immunogenic portion of an ovarian carcinoma protein, or a variant thereof that differs in one or more substitutions, deletions, additions and/or insertions such that the ability of the variant to react with ovarian carcinoma protein-specific antisera is not 25 substantially diminished. Within certain embodiments, the ovarian carcinoma protein comprises a sequence that is encoded by a polynucleotide sequence selected from the group consisting of SEQ ID NO:1, 2, 5, 9, 10, 13, 16, 19, 23, 27, 28, 32, 33, 35, 38, 41-50, 52, 53, 56, 57, 63, 65, 69-72, 75, 78, 80-82, 84, 86, 89-93, 95, 97-100, 103, 107, 111, 114, 117, 120, 121, 125, 128, 132-134, 136, 137, 140, 143-146, 148-151, 156, 158, 30 160-162, 166-168, 171, 174-183, 185, 193-199, 203-205, 208 and 210-214, and complements of such polynucleotides.

The present invention further provides polynucleotides that encode a polypeptide as described above or a portion thereof, expression vectors comprising such polynucleotides and host cells transformed or transfected with such expression vectors.

The present invention further provides polypeptide compositions  
5 comprising an amino acid sequence selected from the group consisting of sequences recited in SEQ ID NO:200-202, 207, 209 and 215.

In certain preferred embodiments, the polypeptides of the present invention are immunogenic, *i.e.*, they are capable of eliciting an immune response, particularly a humoral and/or cellular immune response, as further described herein.

10 The present invention further provides fragments, variants and/or derivatives of the disclosed polypeptide sequences, wherein the fragments, variants and/or derivatives preferably have a level of immunogenic activity of at least about 50%, preferably at least about 70% and more preferably at least about 90% of the level of immunogenic activity of a the ovarian carcinoma protein comprises an amino acid  
15 sequence encoded by a polynucleotide that comprises a sequence recited in any one of SEQ ID NO:1-185, 187-199, 203-206, 208 and 210-214.

Within other aspects, the present invention provides pharmaceutical compositions comprising a polypeptide and/or polynucleotide as described above and a physiologically acceptable carrier.

20 Within a related aspect of the present invention, the pharmaceutical compositions, *e.g.*, vaccine compositions, are provided for prophylactic or therapeutic applications. Such compositions generally comprise an immunogenic polypeptide or polynucleotide of the invention and an immunostimulant, such as an adjuvant.

The present invention further provides pharmaceutical compositions that  
25 comprise: (a) an antibody or antigen-binding fragment thereof that specifically binds to a polypeptide of the present invention, or a fragment thereof; and (b) a physiologically acceptable carrier.

Within further aspects, the present invention provides pharmaceutical compositions comprising: (a) an antigen presenting cell that expresses a polypeptide as  
30 described above and (b) a pharmaceutically acceptable carrier or excipient. Illustrative

antigen presenting cells include dendritic cells, macrophages, monocytes, fibroblasts and B cells.

Within related aspects, pharmaceutical compositions are provided that comprise: (a) an antigen presenting cell that expresses a polypeptide as described above  
5 and (b) an immunostimulant.

The present invention further provides, in other aspects, fusion proteins that comprise at least one polypeptide as described above, as well as polynucleotides encoding such fusion proteins, typically in the form of pharmaceutical compositions, *e.g.*, vaccine compositions, comprising a physiologically acceptable carrier and/or an  
10 immunostimulant. The fusions proteins may comprise multiple immunogenic polypeptides or portions/variants thereof, as described herein, and may further comprise one or more polypeptide segments for facilitating the expression, purification and/or immunogenicity of the polypeptide(s).

Within further aspects, the present invention provides methods for  
15 stimulating an immune response in a patient, preferably a T cell response in a human patient, comprising administering a pharmaceutical composition described herein. The patient may be afflicted with ovarian cancer, in which case the methods provide treatment for the disease, or patient considered at risk for such a disease may be treated prophylactically.

20 Within further aspects, the present invention provides methods for inhibiting the development of a cancer in a patient, comprising administering to a patient a pharmaceutical composition as recited above. The patient may be afflicted with ovarian cancer, in which case the methods provide treatment for the disease, or patient considered at risk for such a disease may be treated prophylactically.

25 The present invention further provides, within other aspects, methods for removing tumor cells from a biological sample, comprising contacting a biological sample with T cells that specifically react with a polypeptide of the present invention, wherein the step of contacting is performed under conditions and for a time sufficient to permit the removal of cells expressing the protein from the sample.

Within related aspects, methods are provided for inhibiting the development of a cancer in a patient, comprising administering to a patient a biological sample treated as described above.

Methods are further provided, within other aspects, for stimulating  
5 and/or expanding T cells specific for a polypeptide of the present invention, comprising contacting T cells with one or more of: (i) an ovarian carcinoma polypeptide as described above; (ii) a polynucleotide encoding such a polypeptide; and/or (iii) an antigen presenting cell that expresses such a polypeptide; under conditions and for a time sufficient to permit the stimulation and/or expansion of T cells. Isolated T cell  
10 populations comprising T cells prepared as described above are also provided.

Within further aspects, the present invention provides methods for inhibiting the development of a cancer in a patient, comprising administering to a patient an effective amount of a T cell population as described above.

The present invention further provides methods for inhibiting the  
15 development of a cancer in a patient, comprising the steps of: (a) incubating CD4<sup>+</sup> and/or CD8<sup>+</sup> T cells isolated from a patient with one or more of: (i) a polypeptide comprising at least an immunogenic portion of polypeptide disclosed herein; (ii) a polynucleotide encoding such a polypeptide; and (iii) an antigen-presenting cell that expressed such a polypeptide; and (b) administering to the patient an effective amount  
20 of the proliferated T cells, and thereby inhibiting the development of a cancer in the patient. Proliferated cells may, but need not, be cloned prior to administration to the patient.

Within further aspects, the present invention provides methods for determining the presence or absence of a cancer, preferably an ovarian cancer, in a  
25 patient comprising: (a) contacting a biological sample obtained from a patient with a binding agent that binds to a polypeptide as recited above; (b) detecting in the sample an amount of polypeptide that binds to the binding agent; and (c) comparing the amount of polypeptide with a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient. Within preferred embodiments, the binding agent  
30 is an antibody, more preferably a monoclonal antibody.

The present invention also provides, within other aspects, methods for monitoring the progression of a cancer in a patient. Such methods comprise the steps of: (a) contacting a biological sample obtained from a patient at a first point in time with a binding agent that binds to a polypeptide as recited above; (b) detecting in the sample an amount of polypeptide that binds to the binding agent; (c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and (d) comparing the amount of polypeptide detected in step (c) with the amount detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

The present invention further provides, within other aspects, methods for determining the presence or absence of a cancer in a patient, comprising the steps of: (a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a polypeptide of the present invention; (b) detecting in the sample a level of a polynucleotide, preferably mRNA, that hybridizes to the oligonucleotide; and (c) comparing the level of polynucleotide that hybridizes to the oligonucleotide with a predetermined cut-off value, and therefrom determining the presence or absence of a cancer in the patient. Within certain embodiments, the amount of mRNA is detected via polymerase chain reaction using, for example, at least one oligonucleotide primer that hybridizes to a polynucleotide encoding a polypeptide as recited above, or a complement of such a polynucleotide. Within other embodiments, the amount of mRNA is detected using a hybridization technique, employing an oligonucleotide probe that hybridizes to a polynucleotide that encodes a polypeptide as recited above, or a complement of such a polynucleotide.

In related aspects, methods are provided for monitoring the progression of a cancer in a patient, comprising the steps of: (a) contacting a biological sample obtained from a patient with an oligonucleotide that hybridizes to a polynucleotide that encodes a polypeptide of the present invention; (b) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide; (c) repeating steps (a) and (b) using a biological sample obtained from the patient at a subsequent point in time; and (d) comparing the amount of polynucleotide detected in step (c) with the amount

detected in step (b) and therefrom monitoring the progression of the cancer in the patient.

Within further aspects, the present invention provides antibodies, such as monoclonal antibodies, that bind to a polypeptide as described above, as well as  
5 diagnostic kits comprising such antibodies. Diagnostic kits comprising one or more oligonucleotide probes or primers as described above are also provided.

These and other aspects of the present invention will become apparent upon reference to the following detailed description. All references disclosed herein are hereby incorporated by reference in their entirety as if each was incorporated  
10 individually.

#### BRIEF DESCRIPTION OF THE SEQUENCE IDENTIFIERS

SEQ ID NO:1, 2, 5, 9, 10, 13, 16, 19, 23, 27, 28, 32, 33, 35, 38, 41-50, 52, 53, 56, 57, 63, 65, 69-72, 75, 78, 80-82, 84, 86, 89-93, 95, 97-100, 103, 107, 111, 114, 117, 120, 121, 125, 128, 132-134, 136, 137, 140, 143-146, 148-151, 156, 158, 160-162, 166-  
15 168, 171, 174-183, 185, and 193-199 are described in Tables III-VII below.

SEQ ID NO:200 is the amino acid sequence of a polypeptide encoded by the polynucleotide recited in SEQ ID NO:182;

SEQ ID NO:201 is the amino acid sequence of a polypeptide encoded by the polynucleotide recited in SEQ ID NO:182;

20 SEQ ID NO:202 is the amino acid sequence of a polypeptide encoded by the polynucleotide recited in SEQ ID NO:182.

SEQ ID NO:203 is the determined extended cDNA sequence for SEQ ID NO:197.

25 SEQ ID NO:204 is the determined extended cDNA sequence for SEQ ID NO:198.

SEQ ID NO:205 is the determined extended cDNA sequence for SEQ ID NO:199.

SEQ ID NO:206 is the determined cDNA sequence for the coding region of O568S fused to an N-terminal His tag.

SEQ ID NO:207 is the amino acid sequence of the polypeptide encoded by the polynucleotide recited in SEQ ID NO:206.

SEQ ID NO:208 is the determined cDNA sequence for the coding region of GPR39 as downloaded from the High Throughput Genomics Database.

5 SEQ ID NO:209 is the amino acid sequence encoded by the cDNA sequence recited in SEQ ID NO:208.

SEQ ID NO:210 is the nucleotide sequence of O1034C an ovary specific EST clone discovered using electronic subtraction.

SEQ ID NO:211 is the full length nucleotide sequence of O591S.

10 SEQ ID NO:212 is the sequence BF345141 which shows sequence homology with O1034C/O591S allowing for the extension of O591S.

SEQ ID NO:213 is the sequence BE336607 which shows sequence homology with O1034C/O591S allowing for the extension of O591S.

15 SEQ ID NO:214 is the consensus nucleotide sequence of O1034C/O591S containing 1897 base pairs.

SEQ ID NO:215 is the predicted translation of the open reading frame identified within SEQ ID NO:214 (nucleotides 260-682).

#### DETAILED DESCRIPTION OF THE INVENTION

20 The present invention is directed generally to compositions and their use in the therapy and diagnosis of cancer, particularly ovarian cancer. As described further below, illustrative compositions of the present invention include, but are not restricted to, polypeptides, particularly immunogenic polypeptides, polynucleotides encoding such polypeptides, antibodies and other binding agents, antigen presenting cells (APCs) and immune system cells (*e.g.*, T cells).

25 The practice of the present invention will employ, unless indicated specifically to the contrary, conventional methods of virology, immunology, microbiology, molecular biology and recombinant DNA techniques within the skill of the art, many of which are described below for the purpose of illustration. Such techniques are explained fully in the literature. See, *e.g.*, Sambrook, et al. Molecular Cloning: A Laboratory Manual (2nd Edition, 1989); Maniatis et al. Molecular Cloning:  
30

- A Laboratory Manual (1982); DNA Cloning: A Practical Approach, vol. I & II (D. Glover, ed.); Oligonucleotide Synthesis (N. Gait, ed., 1984); Nucleic Acid Hybridization (B. Hames & S. Higgins, eds., 1985); Transcription and Translation (B. Hames & S. Higgins, eds., 1984); Animal Cell Culture (R. Freshney, ed., 1986); Perbal,
- 5 A Practical Guide to Molecular Cloning (1984).

All publications, patents and patent applications cited herein, whether supra or infra, are hereby incorporated by reference in their entirety.

As used in this specification and the appended claims, the singular forms "a," "an" and "the" include plural references unless the content clearly dictates

10 otherwise.

#### Polypeptide Compositions

As used herein, the term "polypeptide" is used in its conventional meaning, *i.e.*, as a sequence of amino acids. The polypeptides are not limited to a specific length of the product; thus, peptides, oligopeptides, and proteins are included

15 within the definition of polypeptide, and such terms may be used interchangeably herein unless specifically indicated otherwise. This term also does not refer to or exclude post-expression modifications of the polypeptide, for example, glycosylations, acetylations, phosphorylations and the like, as well as other modifications known in the art, both naturally occurring and non-naturally occurring. A polypeptide may be an entire

20 protein, or a subsequence thereof. Particular polypeptides of interest in the context of this invention are amino acid subsequences comprising epitopes, *i.e.*, antigenic determinants substantially responsible for the immunogenic properties of a polypeptide and being capable of evoking an immune response.

Particularly illustrative polypeptides of the present invention comprise

25 those encoded by a polynucleotide sequence set forth in any one of SEQ ID NO:1, 2, 5, 9, 10, 13, 16, 19, 23, 27, 28, 32, 33, 35, 38, 41-50, 52, 53, 56, 57, 63, 65, 69-72, 75, 78, 80-82, 84, 86, 89-93, 95, 97-100, 103, 107, 111, 114, 117, 120, 121, 125, 128, 132-134, 136, 137, 140, 143-146, 148-151, 156, 158, 160-162, 166-168, 171, 174-183, 185, 193-199, 203-206, 208, and 210-214 or a sequence that hybridizes under moderately

30 stringent conditions, or, alternatively, under highly stringent conditions, to a

polynucleotide sequence identified above. Certain other illustrative polypeptides of the invention comprise amino acid sequences as set forth in any one of SEQ ID NO:200-202, 207, 209 and 215.

The polypeptides of the present invention are sometimes herein referred  
5 to as ovarian tumor proteins or ovarian tumor polypeptides, as an indication that their identification has been based at least in part upon their increased levels of expression in ovarian tumor samples. Thus, a "ovarian tumor polypeptide" or "ovarian tumor protein," refers generally to a polypeptide sequence of the present invention, or a polynucleotide sequence encoding such a polypeptide, that is expressed in a substantial  
10 proportion of ovarian tumor samples, for example preferably greater than about 20%, more preferably greater than about 30%, and most preferably greater than about 50% or more of ovarian tumor samples tested, at a level that is at least two fold, and preferably at least five fold, greater than the level of expression in normal tissues, as determined using a representative assay provided herein. An ovarian tumor polypeptide sequence  
15 of the invention, based upon its increased level of expression in tumor cells, has particular utility both as a diagnostic marker as well as a therapeutic target, as further described below.

In certain preferred embodiments, the polypeptides of the invention are immunogenic, *i.e.*, they react detectably within an immunoassay (such as an ELISA or  
20 T-cell stimulation assay) with antisera and/or T-cells from a patient with ovarian cancer. Screening for immunogenic activity can be performed using techniques well known to the skilled artisan. For example, such screens can be performed using methods such as those described in Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In one illustrative example, a polypeptide may be  
25 immobilized on a solid support and contacted with patient sera to allow binding of antibodies within the sera to the immobilized polypeptide. Unbound sera may then be removed and bound antibodies detected using, for example, <sup>125</sup>I-labeled Protein A.

As would be recognized by the skilled artisan, immunogenic portions of the polypeptides disclosed herein are also encompassed by the present invention. An  
30 "immunogenic portion," as used herein, is a fragment of an immunogenic polypeptide of the invention that itself is immunologically reactive (*i.e.*, specifically binds) with the

B-cells and/or T-cell surface antigen receptors that recognize the polypeptide. Immunogenic portions may generally be identified using well known techniques, such as those summarized in Paul, *Fundamental Immunology*, 3rd ed., 243-247 (Raven Press, 1993) and references cited therein. Such techniques include screening polypeptides for the ability to react with antigen-specific antibodies, antisera and/or T-cell lines or clones. As used herein, antisera and antibodies are "antigen-specific" if they specifically bind to an antigen (*i.e.*, they react with the protein in an ELISA or other immunoassay, and do not react detectably with unrelated proteins). Such antisera and antibodies may be prepared as described herein, and using well-known techniques.

10 In one preferred embodiment, an immunogenic portion of a polypeptide of the present invention is a portion that reacts with antisera and/or T-cells at a level that is not substantially less than the reactivity of the full-length polypeptide (*e.g.*, in an ELISA and/or T-cell reactivity assay). Preferably, the level of immunogenic activity of the immunogenic portion is at least about 50%, preferably at least about 70% and most preferably greater than about 90% of the immunogenicity for the full-length polypeptide. In some instances, preferred immunogenic portions will be identified that have a level of immunogenic activity greater than that of the corresponding full-length polypeptide, *e.g.*, having greater than about 100% or 150% or more immunogenic activity.

20 In certain other embodiments, illustrative immunogenic portions may include peptides in which an N-terminal leader sequence and/or transmembrane domain have been deleted. Other illustrative immunogenic portions will contain a small N- and/or C-terminal deletion (*e.g.*, 1-30 amino acids, preferably 5-15 amino acids), relative to the mature protein.

25 In another embodiment, a polypeptide composition of the invention may also comprise one or more polypeptides that are immunologically reactive with T cells and/or antibodies generated against a polypeptide of the invention, particularly a polypeptide having an amino acid sequence disclosed herein, or to an immunogenic fragment or variant thereof.

30 In another embodiment of the invention, polypeptides are provided that comprise one or more polypeptides that are capable of eliciting T cells and/or antibodies

that are immunologically reactive with one or more polypeptides described herein, or one or more polypeptides encoded by contiguous nucleic acid sequences contained in the polynucleotide sequences disclosed herein, or immunogenic fragments or variants thereof, or to one or more nucleic acid sequences which hybridize to one or more of these sequences under conditions of moderate to high stringency.

The present invention, in another aspect, provides polypeptide fragments comprising at least about 5, 10, 15, 20, 25, 50, or 100 contiguous amino acids, or more, including all intermediate lengths, of a polypeptide compositions set forth herein, such as those set forth in SEQ ID NO:200-202, 207, 209, and 215 or those encoded by a polynucleotide sequence set forth in any one of SEQ ID NO:1, 2, 5, 9, 10, 13, 16, 19, 23, 27, 28, 32, 33, 35, 38, 41-50, 52, 53, 56, 57, 63, 65, 69-72, 75, 78, 80-82, 84, 86, 89-93, 95, 97-100, 103, 107, 111, 114, 117, 120, 121, 125, 128, 132-134, 136, 137, 140, 143-146, 148-151, 156, 158, 160-162, 166-168, 171, 174-183, 185, 193-199, 203-206, 208, and 210-214.

In another aspect, the present invention provides variants of the polypeptide compositions described herein. Polypeptide variants generally encompassed by the present invention will typically exhibit at least about 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, or 99% or more identity (determined as described below), along its length, to a polypeptide sequences set forth herein.

In one preferred embodiment, the polypeptide fragments and variants provide by the present invention are immunologically reactive with an antibody and/or T-cell that reacts with a full-length polypeptide specifically set for the herein.

In another preferred embodiment, the polypeptide fragments and variants provided by the present invention exhibit a level of immunogenic activity of at least about 50%, preferably at least about 70%, and most preferably at least about 90% or more of that exhibited by a full-length polypeptide sequence specifically set forth herein.

A polypeptide "variant," as the term is used herein, is a polypeptide that typically differs from a polypeptide specifically disclosed herein in one or more substitutions, deletions, additions and/or insertions. Such variants may be naturally

occurring or may be synthetically generated, for example, by modifying one or more of the above polypeptide sequences of the invention and evaluating their immunogenic activity as described herein and/or using any of a number of techniques well known in the art.

5 For example, certain illustrative variants of the polypeptides of the invention include those in which one or more portions, such as an N-terminal leader sequence or transmembrane domain, have been removed. Other illustrative variants include variants in which a small portion (*e.g.*, 1-30 amino acids, preferably 5-15 amino acids) has been removed from the N- and/or C-terminal of the mature protein.

10 In many instances, a variant will contain conservative substitutions. A "conservative substitution" is one in which an amino acid is substituted for another amino acid that has similar properties, such that one skilled in the art of peptide chemistry would expect the secondary structure and hydrophobic nature of the polypeptide to be substantially unchanged. As described above, modifications may be  
15 made in the structure of the polynucleotides and polypeptides of the present invention and still obtain a functional molecule that encodes a variant or derivative polypeptide with desirable characteristics, *e.g.*, with immunogenic characteristics. When it is desired to alter the amino acid sequence of a polypeptide to create an equivalent, or even an improved, immunogenic variant or portion of a polypeptide of the invention,  
20 one skilled in the art will typically change one or more of the codons of the encoding DNA sequence according to Table 1.

For example, certain amino acids may be substituted for other amino acids in a protein structure without appreciable loss of interactive binding capacity with structures such as, for example, antigen-binding regions of antibodies or binding sites  
25 on substrate molecules. Since it is the interactive capacity and nature of a protein that defines that protein's biological functional activity, certain amino acid sequence substitutions can be made in a protein sequence, and, of course, its underlying DNA coding sequence, and nevertheless obtain a protein with like properties. It is thus contemplated that various changes may be made in the peptide sequences of the  
30 disclosed compositions, or corresponding DNA sequences which encode said peptides without appreciable loss of their biological utility or activity.

TABLE 1

Amino Acids			Codons						
Alanine	Ala	A	GCA	GCC	GCG	GCU			
Cysteine	Cys	C	UGC	UGU					
Aspartic acid	Asp	D	GAC	GAU					
Glutamic acid	Glu	E	GAA	GAG					
Phenylalanine	Phe	F	UUC	UUU					
Glycine	Gly	G	GGA	GGC	GGG	GGU			
Histidine	His	H	CAC	CAU					
Isoleucine	Ile	I	AUA	AUC	AUU				
Lysine	Lys	K	AAA	AAG					
Leucine	Leu	L	UUA	UUG	CUA	CUC	CUG	CUU	
Methionine	Met	M	AUG						
Asparagine	Asn	N	AAC	AAU					
Proline	Pro	P	CCA	CCC	CCG	CCU			
Glutamine	Gln	Q	CAA	CAG					
Arginine	Arg	R	AGA	AGG	CGA	CGC	CGG	CGU	
Serine	Ser	S	AGC	AGU	UCA	UCC	UCG	UCU	
Threonine	Thr	T	ACA	ACC	ACG	ACU			
Valine	Val	V	GUA	GUC	GUG	GUU			
Tryptophan	Trp	W	UGG						
Tyrosine	Tyr	Y	UAC	UAU					

In making such changes, the hydropathic index of amino acids may be considered. The importance of the hydropathic amino acid index in conferring interactive biologic function on a protein is generally understood in the art (Kyte and Doolittle, 1982, incorporated herein by reference). It is accepted that the relative hydropathic character of the amino acid contributes to the secondary structure of the resultant protein, which in turn defines the interaction of the protein with other molecules, for example, enzymes, substrates, receptors, DNA, antibodies, antigens, and the like. Each amino acid has been assigned a hydropathic index on the basis of its hydrophobicity and charge characteristics (Kyte and Doolittle, 1982). These values are:

isoleucine (+4.5); valine (+4.2); leucine (+3.8); phenylalanine (+2.8); cysteine/cystine (+2.5); methionine (+1.9); alanine (+1.8); glycine (−0.4); threonine (−0.7); serine (−0.8); tryptophan (−0.9); tyrosine (−1.3); proline (−1.6); histidine (−3.2); glutamate (−3.5); glutamine (−3.5); aspartate (−3.5); asparagine (−3.5); lysine (−3.9); and arginine (−4.5).

5           It is known in the art that certain amino acids may be substituted by other amino acids having a similar hydropathic index or score and still result in a protein with similar biological activity, *i.e.* still obtain a biological functionally equivalent protein. In making such changes, the substitution of amino acids whose hydropathic indices are within  $\pm 2$  is preferred, those within  $\pm 1$  are particularly preferred, and those within  $\pm 0.5$  are even more particularly preferred. It is also understood in the art that the substitution of like amino acids can be made effectively on the basis of hydrophilicity. U. S. Patent 10 4,554,101 (specifically incorporated herein by reference in its entirety), states that the greatest local average hydrophilicity of a protein, as governed by the hydrophilicity of its adjacent amino acids, correlates with a biological property of the protein.

15           As detailed in U. S. Patent 4,554,101, the following hydrophilicity values have been assigned to amino acid residues: arginine (+3.0); lysine (+3.0); aspartate (+3.0  $\pm$  1); glutamate (+3.0  $\pm$  1); serine (+0.3); asparagine (+0.2); glutamine (+0.2); glycine (0); threonine (−0.4); proline (−0.5  $\pm$  1); alanine (−0.5); histidine (−0.5); cysteine (−1.0); methionine (−1.3); valine (−1.5); leucine (−1.8); isoleucine (−1.8); tyrosine (−2.3); phenylalanine (−2.5); tryptophan (−3.4). It is understood that an amino acid can be 20 substituted for another having a similar hydrophilicity value and still obtain a biologically equivalent, and in particular, an immunologically equivalent protein. In such changes, the substitution of amino acids whose hydrophilicity values are within  $\pm 2$  is preferred, those within  $\pm 1$  are particularly preferred, and those within  $\pm 0.5$  are even 25 more particularly preferred.

          As outlined above, amino acid substitutions are generally therefore based on the relative similarity of the amino acid side-chain substituents, for example, their hydrophobicity, hydrophilicity, charge, size, and the like. Exemplary substitutions that take various of the foregoing characteristics into consideration are well known to those 30 of skill in the art and include: arginine and lysine; glutamate and aspartate; serine and threonine; glutamine and asparagine; and valine, leucine and isoleucine.

In addition, any polynucleotide may be further modified to increase stability *in vivo*. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends; the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages in the backbone; and/or the inclusion of  
5 nontraditional bases such as inosine, queosine and wybutosine, as well as acetyl-methyl-, thio- and other modified forms of adenine, cytidine, guanine, thymine and uridine.

Amino acid substitutions may further be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity and/or the amphipathic  
10 nature of the residues. For example, negatively charged amino acids include aspartic acid and glutamic acid; positively charged amino acids include lysine and arginine; and amino acids with uncharged polar head groups having similar hydrophilicity values include leucine, isoleucine and valine; glycine and alanine; asparagine and glutamine; and serine, threonine, phenylalanine and tyrosine. Other groups of amino acids that may  
15 represent conservative changes include: (1) ala, pro, gly, glu, asp, gln, asn, ser, thr; (2) cys, ser, tyr, thr; (3) val, ile, leu, met, ala, phe; (4) lys, arg, his; and (5) phe, tyr, trp, his. A variant may also, or alternatively, contain nonconservative changes. In a preferred embodiment, variant polypeptides differ from a native sequence by substitution, deletion or addition of five amino acids or fewer. Variants may also (or  
20 alternatively) be modified by, for example, the deletion or addition of amino acids that have minimal influence on the immunogenicity, secondary structure and hydrophobic nature of the polypeptide.

As noted above, polypeptides may comprise a signal (or leader) sequence at the N-terminal end of the protein, which co-translationally or post-translationally  
25 directs transfer of the protein. The polypeptide may also be conjugated to a linker or other sequence for ease of synthesis, purification or identification of the polypeptide (e.g., poly-His), or to enhance binding of the polypeptide to a solid support. For example, a polypeptide may be conjugated to an immunoglobulin Fc region.

When comparing polypeptide sequences, two sequences are said to be  
30 "identical" if the sequence of amino acids in the two sequences is the same when aligned for maximum correspondence, as described below. Comparisons between two

sequences are typically performed by comparing the sequences over a comparison window to identify and compare local regions of sequence similarity. A "comparison window" as used herein, refers to a segment of at least about 20 contiguous positions, usually 30 to about 75, 40 to about 50, in which a sequence may be compared to a  
5 reference sequence of the same number of contiguous positions after the two sequences are optimally aligned.

Optimal alignment of sequences for comparison may be conducted using the Megalign program in the Lasergene suite of bioinformatics software (DNASTAR, Inc., Madison, WI), using default parameters. This program embodies several  
10 alignment schemes described in the following references: Dayhoff, M.O. (1978) A model of evolutionary change in proteins – Matrices for detecting distant relationships. In Dayhoff, M.O. (ed.) Atlas of Protein Sequence and Structure, National Biomedical Research Foundation, Washington DC Vol. 5, Suppl. 3, pp. 345-358; Hein J. (1990) Unified Approach to Alignment and Phylogenies pp. 626-645 *Methods in Enzymology*  
15 vol. 183, Academic Press, Inc., San Diego, CA; Higgins, D.G. and Sharp, P.M. (1989) *CABIOS* 5:151-153; Myers, E.W. and Muller W. (1988) *CABIOS* 4:11-17; Robinson, E.D. (1971) *Comb. Theor* 11:105; Santou, N. Nes, M. (1987) *Mol. Biol. Evol.* 4:406-425; Sneath, P.H.A. and Sokal, R.R. (1973) *Numerical Taxonomy – the Principles and Practice of Numerical Taxonomy*, Freeman Press, San Francisco, CA; Wilbur, W.J. and  
20 Lipman, D.J. (1983) *Proc. Natl. Acad. Sci. USA* 80:726-730.

Alternatively, optimal alignment of sequences for comparison may be conducted by the local identity algorithm of Smith and Waterman (1981) *Add. APL. Math* 2:482, by the identity alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443, by the search for similarity methods of Pearson and Lipman (1988)  
25 *Proc. Natl. Acad. Sci. USA* 85: 2444, by computerized implementations of these algorithms (GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, WI), or by inspection.

One preferred example of algorithms that are suitable for determining  
30 percent sequence identity and sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul et al. (1977) *Nucl. Acids Res.* 25:3389-3402

and Altschul et al. (1990) *J. Mol. Biol.* 215:403-410, respectively. BLAST and BLAST 2.0 can be used, for example with the parameters described herein, to determine percent sequence identity for the polynucleotides and polypeptides of the invention. Software for performing BLAST analyses is publicly available through the National Center for  
5 Biotechnology Information. For amino acid sequences, a scoring matrix can be used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is  
10 reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment.

In one preferred approach, the "percentage of sequence identity" is determined by comparing two optimally aligned sequences over a window of comparison of at least 20 positions, wherein the portion of the polypeptide sequence in  
15 the comparison window may comprise additions or deletions (*i.e.*, gaps) of 20 percent or less, usually 5 to 15 percent, or 10 to 12 percent, as compared to the reference sequences (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical amino acid residue occurs in both sequences to yield the number of  
20 matched positions, dividing the number of matched positions by the total number of positions in the reference sequence (*i.e.*, the window size) and multiplying the results by 100 to yield the percentage of sequence identity.

Within other illustrative embodiments, a polypeptide may be a fusion polypeptide that comprises multiple polypeptides as described herein, or that comprises  
25 at least one polypeptide as described herein and an unrelated sequence, such as a known tumor protein. A fusion partner may, for example, assist in providing T helper epitopes (an immunological fusion partner), preferably T helper epitopes recognized by humans, or may assist in expressing the protein (an expression enhancer) at higher yields than the native recombinant protein. Certain preferred fusion partners are both immunological  
30 and expression enhancing fusion partners. Other fusion partners may be selected so as to increase the solubility of the polypeptide or to enable the polypeptide to be targeted to

desired intracellular compartments. Still further fusion partners include affinity tags, which facilitate purification of the polypeptide.

Fusion polypeptides may generally be prepared using standard techniques, including chemical conjugation. Preferably, a fusion polypeptide is expressed as a recombinant polypeptide, allowing the production of increased levels, relative to a non-fused polypeptide, in an expression system. Briefly, DNA sequences encoding the polypeptide components may be assembled separately, and ligated into an appropriate expression vector. The 3' end of the DNA sequence encoding one polypeptide component is ligated, with or without a peptide linker, to the 5' end of a DNA sequence encoding the second polypeptide component so that the reading frames of the sequences are in phase. This permits translation into a single fusion polypeptide that retains the biological activity of both component polypeptides.

A peptide linker sequence may be employed to separate the first and second polypeptide components by a distance sufficient to ensure that each polypeptide folds into its secondary and tertiary structures. Such a peptide linker sequence is incorporated into the fusion polypeptide using standard techniques well known in the art. Suitable peptide linker sequences may be chosen based on the following factors: (1) their ability to adopt a flexible extended conformation; (2) their inability to adopt a secondary structure that could interact with functional epitopes on the first and second polypeptides; and (3) the lack of hydrophobic or charged residues that might react with the polypeptide functional epitopes. Preferred peptide linker sequences contain Gly, Asn and Ser residues. Other near neutral amino acids, such as Thr and Ala may also be used in the linker sequence. Amino acid sequences which may be usefully employed as linkers include those disclosed in Maratea et al., *Gene* 40:39-46, 1985; Murphy et al., *Proc. Natl. Acad. Sci. USA* 83:8258-8262, 1986; U.S. Patent No. 4,935,233 and U.S. Patent No. 4,751,180. The linker sequence may generally be from 1 to about 50 amino acids in length. Linker sequences are not required when the first and second polypeptides have non-essential N-terminal amino acid regions that can be used to separate the functional domains and prevent steric interference.

The ligated DNA sequences are operably linked to suitable transcriptional or translational regulatory elements. The regulatory elements

responsible for expression of DNA are located only 5' to the DNA sequence encoding the first polypeptides. Similarly, stop codons required to end translation and transcription termination signals are only present 3' to the DNA sequence encoding the second polypeptide.

5           The fusion polypeptide can comprise a polypeptide as described herein together with an unrelated immunogenic protein, such as an immunogenic protein capable of eliciting a recall response. Examples of such proteins include tetanus, tuberculosis and hepatitis proteins (*see, for example, Stoute et al. New Engl. J. Med., 336:86-91, 1997*).

10           In one preferred embodiment, the immunological fusion partner is derived from a *Mycobacterium* sp., such as a *Mycobacterium tuberculosis*-derived Ra12 fragment. Ra12 compositions and methods for their use in enhancing the expression and/or immunogenicity of heterologous polynucleotide/polypeptide sequences is described in U.S. Patent Application 60/158,585, the disclosure of which is  
15 incorporated herein by reference in its entirety. Briefly, Ra12 refers to a polynucleotide region that is a subsequence of a *Mycobacterium tuberculosis* MTB32A nucleic acid. MTB32A is a serine protease of 32 KD molecular weight encoded by a gene in virulent and avirulent strains of *M. tuberculosis*. The nucleotide sequence and amino acid sequence of MTB32A have been described (for example, U.S. Patent Application  
20 60/158,585; *see also, Skeiky et al., Infection and Immun. (1999) 67:3998-4007*, incorporated herein by reference). C-terminal fragments of the MTB32A coding sequence express at high levels and remain as a soluble polypeptides throughout the purification process. Moreover, Ra12 may enhance the immunogenicity of heterologous immunogenic polypeptides with which it is fused. One preferred Ra12 fusion  
25 polypeptide comprises a 14 KD C-terminal fragment corresponding to amino acid residues 192 to 323 of MTB32A. Other preferred Ra12 polynucleotides generally comprise at least about 15 consecutive nucleotides, at least about 30 nucleotides, at least about 60 nucleotides, at least about 100 nucleotides, at least about 200 nucleotides, or at least about 300 nucleotides that encode a portion of a Ra12 polypeptide. Ra12  
30 polynucleotides may comprise a native sequence (*i.e.*, an endogenous sequence that encodes a Ra12 polypeptide or a portion thereof) or may comprise a variant of such a

sequence. Ra12 polynucleotide variants may contain one or more substitutions, additions, deletions and/or insertions such that the biological activity of the encoded fusion polypeptide is not substantially diminished, relative to a fusion polypeptide comprising a native Ra12 polypeptide. Variants preferably exhibit at least about 70% identity, more preferably at least about 80% identity and most preferably at least about 90% identity to a polynucleotide sequence that encodes a native Ra12 polypeptide or a portion thereof.

Within other preferred embodiments, an immunological fusion partner is derived from protein D, a surface protein of the gram-negative bacterium *Haemophilus influenza B* (WO 91/18926). Preferably, a protein D derivative comprises approximately the first third of the protein (e.g., the first N-terminal 100-110 amino acids), and a protein D derivative may be lipidated. Within certain preferred embodiments, the first 109 residues of a Lipoprotein D fusion partner is included on the N-terminus to provide the polypeptide with additional exogenous T-cell epitopes and to increase the expression level in *E. coli* (thus functioning as an expression enhancer). The lipid tail ensures optimal presentation of the antigen to antigen presenting cells. Other fusion partners include the non-structural protein from influenzae virus, NS1 (hemagglutinin). Typically, the N-terminal 81 amino acids are used, although different fragments that include T-helper epitopes may be used.

In another embodiment, the immunological fusion partner is the protein known as LYTA, or a portion thereof (preferably a C-terminal portion). LYTA is derived from *Streptococcus pneumoniae*, which synthesizes an N-acetyl-L-alanine amidase known as amidase LYTA (encoded by the *LytA* gene; *Gene* 43:265-292, 1986). LYTA is an autolysin that specifically degrades certain bonds in the peptidoglycan backbone. The C-terminal domain of the LYTA protein is responsible for the affinity to the choline or to some choline analogues such as DEAE. This property has been exploited for the development of *E. coli* C-LYTA expressing plasmids useful for expression of fusion proteins. Purification of hybrid proteins containing the C-LYTA fragment at the amino terminus has been described (see *Biotechnology* 10:795-798, 1992). Within a preferred embodiment, a repeat portion of LYTA may be incorporated

into a fusion polypeptide. A repeat portion is found in the C-terminal region starting at residue 178. A particularly preferred repeat portion incorporates residues 188-305.

Yet another illustrative embodiment involves fusion polypeptides, and the polynucleotides encoding them, wherein the fusion partner comprises a targeting  
5 signal capable of directing a polypeptide to the endosomal/lysosomal compartment, as described in U.S. Patent No. 5,633,234. An immunogenic polypeptide of the invention, when fused with this targeting signal, will associate more efficiently with MHC class II molecules and thereby provide enhanced in vivo stimulation of CD4<sup>+</sup> T-cells specific for the polypeptide.

10 Polypeptides of the invention are prepared using any of a variety of well known synthetic and/or recombinant techniques, the latter of which are further described below. Polypeptides, portions and other variants generally less than about 150 amino acids can be generated by synthetic means, using techniques well known to those of ordinary skill in the art. In one illustrative example, such polypeptides are  
15 synthesized using any of the commercially available solid-phase techniques, such as the Merrifield solid-phase synthesis method, where amino acids are sequentially added to a growing amino acid chain. See Merrifield, *J. Am. Chem. Soc.* 85:2149-2146, 1963. Equipment for automated synthesis of polypeptides is commercially available from suppliers such as Perkin Elmer/Applied BioSystems Division (Foster City, CA), and  
20 may be operated according to the manufacturer's instructions.

In general, polypeptide compositions (including fusion polypeptides) of the invention are isolated. An "isolated" polypeptide is one that is removed from its original environment. For example, a naturally-occurring protein or polypeptide is isolated if it is separated from some or all of the coexisting materials in the natural  
25 system. Preferably, such polypeptides are also purified, e.g., are at least about 90% pure, more preferably at least about 95% pure and most preferably at least about 99% pure.

#### Polynucleotide Compositions

The present invention, in other aspects, provides polynucleotide  
30 compositions. The terms "DNA" and "polynucleotide" are used essentially

interchangeably herein to refer to a DNA molecule that has been isolated free of total genomic DNA of a particular species. "Isolated," as used herein, means that a polynucleotide is substantially away from other coding sequences, and that the DNA molecule does not contain large portions of unrelated coding DNA, such as large  
5 chromosomal fragments or other functional genes or polypeptide coding regions. Of course, this refers to the DNA molecule as originally isolated, and does not exclude genes or coding regions later added to the segment by the hand of man.

As will be understood by those skilled in the art, the polynucleotide compositions of this invention can include genomic sequences, extra-genomic and  
10 plasmid-encoded sequences and smaller engineered gene segments that express, or may be adapted to express, proteins, polypeptides, peptides and the like. Such segments may be naturally isolated, or modified synthetically by the hand of man.

As will be also recognized by the skilled artisan, polynucleotides of the invention may be single-stranded (coding or antisense) or double-stranded, and may be  
15 DNA (genomic, cDNA or synthetic) or RNA molecules. RNA molecules may include HnRNA molecules, which contain introns and correspond to a DNA molecule in a one-to-one manner, and mRNA molecules, which do not contain introns. Additional coding or non-coding sequences may, but need not, be present within a polynucleotide of the present invention, and a polynucleotide may, but need not, be linked to other molecules  
20 and/or support materials.

Polynucleotides may comprise a native sequence (*i.e.*, an endogenous sequence that encodes a polypeptide/protein of the invention or a portion thereof) or may comprise a sequence that encodes a variant or derivative, preferably and immunogenic variant or derivative, of such a sequence.

25 Therefore, according to another aspect of the present invention, polynucleotide compositions are provided that comprise some or all of a polynucleotide sequence set forth in any one of SEQ ID NO:1, 2, 5, 9, 10, 13, 16, 19, 23, 27, 28, 32, 33, 35, 38, 41-50, 52, 53, 56, 57, 63, 65, 69-72, 75, 78, 80-82, 84, 86, 89-93, 95, 97-100, 103, 107, 111, 114, 117, 120, 121, 125, 128, 132-134, 136, 137, 140, 143-146, 148-151,  
30 156, 158, 160-162, 166-168, 171, 174-183, 185, 193-199, 203-206, 208 and 210-214, complements of a polynucleotide sequence set forth as described above, and degenerate

variants of a polynucleotide sequence set forth as described above. In certain preferred embodiments, the polynucleotide sequences set forth herein encode immunogenic polypeptides, as described above.

In other related embodiments, the present invention provides  
5 polynucleotide variants having substantial identity to the sequences disclosed herein in SEQ ID NO:1, 2, 5, 9, 10, 13, 16, 19, 23, 27, 28, 32, 33, 35, 38, 41-50, 52, 53, 56, 57, 63, 65, 69-72, 75, 78, 80-82, 84, 86, 89-93, 95, 97-100, 103, 107, 111, 114, 117, 120, 121, 125, 128, 132-134, 136, 137, 140, 143-146, 148-151, 156, 158, 160-162, 166-168, 171, 174-183, 185, 193-199, 203-206, 208 and 210-214, for example those comprising  
10 at least 70% sequence identity, preferably at least 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, or 99% or higher, sequence identity compared to a polynucleotide sequence of this invention using the methods described herein, (*e.g.*, BLAST analysis using standard parameters, as described below). One skilled in this art will recognize that these values can be appropriately adjusted to determine corresponding identity of  
15 proteins encoded by two nucleotide sequences by taking into account codon degeneracy, amino acid similarity, reading frame positioning and the like.

Typically, polynucleotide variants will contain one or more substitutions, additions, deletions and/or insertions, preferably such that the immunogenicity of the polypeptide encoded by the variant polynucleotide is not substantially diminished  
20 relative to a polypeptide encoded by a polynucleotide sequence specifically set forth herein). The term "variants" should also be understood to encompass homologous genes of xenogenic origin.

In additional embodiments, the present invention provides polynucleotide fragments comprising various lengths of contiguous stretches of  
25 sequence identical to or complementary to one or more of the sequences disclosed herein. For example, polynucleotides are provided by this invention that comprise at least about 10, 15, 20, 30, 40, 50, 75, 100, 150, 200, 300, 400, 500 or 1000 or more contiguous nucleotides of one or more of the sequences disclosed herein as well as all intermediate lengths there between. It will be readily understood that "intermediate  
30 lengths", in this context, means any length between the quoted values, such as 16, 17, 18, 19, *etc.*; 21, 22, 23, *etc.*; 30, 31, 32, *etc.*; 50, 51, 52, 53, *etc.*; 100, 101, 102, 103,

*etc.*; 150, 151, 152, 153, *etc.*; including all integers through 200-500; 500-1,000, and the like.

In another embodiment of the invention, polynucleotide compositions are provided that are capable of hybridizing under moderate to high stringency conditions to a polynucleotide sequence provided herein, or a fragment thereof, or a complementary sequence thereof. Hybridization techniques are well known in the art of molecular biology. For purposes of illustration, suitable moderately stringent conditions for testing the hybridization of a polynucleotide of this invention with other polynucleotides include prewashing in a solution of 5 X SSC, 0.5% SDS, 1.0 mM EDTA (pH 8.0); hybridizing at 50°C-60°C, 5 X SSC, overnight; followed by washing twice at 65°C for 20 minutes with each of 2X, 0.5X and 0.2X SSC containing 0.1% SDS. One skilled in the art will understand that the stringency of hybridization can be readily manipulated, such as by altering the salt content of the hybridization solution and/or the temperature at which the hybridization is performed. For example, in another embodiment, suitable highly stringent hybridization conditions include those described above, with the exception that the temperature of hybridization is increased, *e.g.*, to 60-65°C or 65-70°C.

In certain preferred embodiments, the polynucleotides described above, *e.g.*, polynucleotide variants, fragments and hybridizing sequences, encode polypeptides that are immunologically cross-reactive with a polypeptide sequence specifically set forth herein. In other preferred embodiments, such polynucleotides encode polypeptides that have a level of immunogenic activity of at least about 50%, preferably at least about 70%, and more preferably at least about 90% of that for a polypeptide sequence specifically set forth herein.

The polynucleotides of the present invention, or fragments thereof, regardless of the length of the coding sequence itself, may be combined with other DNA sequences, such as promoters, polyadenylation signals, additional restriction enzyme sites, multiple cloning sites, other coding segments, and the like, such that their overall length may vary considerably. It is therefore contemplated that a nucleic acid fragment of almost any length may be employed, with the total length preferably being limited by the ease of preparation and use in the intended recombinant DNA protocol. For

example, illustrative polynucleotide segments with total lengths of about 10,000, about 5000, about 3000, about 2,000, about 1,000, about 500, about 200, about 100, about 50 base pairs in length, and the like, (including all intermediate lengths) are contemplated to be useful in many implementations of this invention.

5                   When comparing polynucleotide sequences, two sequences are said to be “identical” if the sequence of nucleotides in the two sequences is the same when aligned for maximum correspondence, as described below. Comparisons between two sequences are typically performed by comparing the sequences over a comparison window to identify and compare local regions of sequence similarity. A “comparison  
10 window” as used herein, refers to a segment of at least about 20 contiguous positions, usually 30 to about 75, 40 to about 50, in which a sequence may be compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned.

Optimal alignment of sequences for comparison may be conducted using  
15 the Megalign program in the Lasergene suite of bioinformatics software (DNASTAR, Inc., Madison, WI), using default parameters. This program embodies several alignment schemes described in the following references: Dayhoff, M.O. (1978) A model of evolutionary change in proteins – Matrices for detecting distant relationships. In Dayhoff, M.O. (ed.) *Atlas of Protein Sequence and Structure*, National Biomedical  
20 Research Foundation, Washington DC Vol. 5, Suppl. 3, pp. 345-358; Hein J. (1990) Unified Approach to Alignment and Phylogenies pp. 626-645 *Methods in Enzymology* vol. 183, Academic Press, Inc., San Diego, CA; Higgins, D.G. and Sharp, P.M. (1989) *CABIOS* 5:151-153; Myers, E.W. and Muller W. (1988) *CABIOS* 4:11-17; Robinson, E.D. (1971) *Comb. Theor* 11:105; Santou, N. Nes, M. (1987) *Mol. Biol. Evol.* 4:406-  
25 425; Sneath, P.H.A. and Sokal, R.R. (1973) *Numerical Taxonomy – the Principles and Practice of Numerical Taxonomy*, Freeman Press, San Francisco, CA; Wilbur, W.J. and Lipman, D.J. (1983) *Proc. Natl. Acad., Sci. USA* 80:726-730.

Alternatively, optimal alignment of sequences for comparison may be conducted by the local identity algorithm of Smith and Waterman (1981) *Add. APL.*  
30 *Math* 2:482, by the identity alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443, by the search for similarity methods of Pearson and Lipman (1988)

*Proc. Natl. Acad. Sci. USA* 85: 2444, by computerized implementations of these algorithms (GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, WI), or by inspection.

5           One preferred example of algorithms that are suitable for determining percent sequence identity and sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul et al. (1977) *Nucl. Acids Res.* 25:3389-3402 and Altschul et al. (1990) *J. Mol. Biol.* 215:403-410, respectively. BLAST and BLAST 2.0 can be used, for example with the parameters described herein, to determine percent  
10 sequence identity for the polynucleotides of the invention. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information. In one illustrative example, cumulative scores can be calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always >0) and N (penalty score for mismatching residues; always <0). Extension of  
15 the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T and X determine the sensitivity and speed of the alignment. The BLASTN program (for  
20 nucleotide sequences) uses as defaults a wordlength (W) of 11, and expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff and Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915) alignments, (B) of 50, expectation (E) of 10, M=5, N=-4 and a comparison of both strands.

          Preferably, the "percentage of sequence identity" is determined by  
25 comparing two optimally aligned sequences over a window of comparison of at least 20 positions, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (*i.e.*, gaps) of 20 percent or less, usually 5 to 15 percent, or 10 to 12 percent, as compared to the reference sequences (which does not comprise additions or deletions) for optimal alignment of the two sequences. The  
30 percentage is calculated by determining the number of positions at which the identical nucleic acid bases occurs in both sequences to yield the number of matched positions,

dividing the number of matched positions by the total number of positions in the reference sequence (*i.e.*, the window size) and multiplying the results by 100 to yield the percentage of sequence identity.

It will be appreciated by those of ordinary skill in the art that, as a result of the degeneracy of the genetic code, there are many nucleotide sequences that encode a polypeptide as described herein. Some of these polynucleotides bear minimal homology to the nucleotide sequence of any native gene. Nonetheless, polynucleotides that vary due to differences in codon usage are specifically contemplated by the present invention. Further, alleles of the genes comprising the polynucleotide sequences provided herein are within the scope of the present invention. Alleles are endogenous genes that are altered as a result of one or more mutations, such as deletions, additions and/or substitutions of nucleotides. The resulting mRNA and protein may, but need not, have an altered structure or function. Alleles may be identified using standard techniques (such as hybridization, amplification and/or database sequence comparison).

Therefore, in another embodiment of the invention, a mutagenesis approach, such as site-specific mutagenesis, is employed for the preparation of immunogenic variants and/or derivatives of the polypeptides described herein. By this approach, specific modifications in a polypeptide sequence can be made through mutagenesis of the underlying polynucleotides that encode them. These techniques provides a straightforward approach to prepare and test sequence variants, for example, incorporating one or more of the foregoing considerations, by introducing one or more nucleotide sequence changes into the polynucleotide.

Site-specific mutagenesis allows the production of mutants through the use of specific oligonucleotide sequences which encode the DNA sequence of the desired mutation, as well as a sufficient number of adjacent nucleotides, to provide a primer sequence of sufficient size and sequence complexity to form a stable duplex on both sides of the deletion junction being traversed. Mutations may be employed in a selected polynucleotide sequence to improve, alter, decrease, modify, or otherwise change the properties of the polynucleotide itself, and/or alter the properties, activity, composition, stability, or primary sequence of the encoded polypeptide.

In certain embodiments of the present invention, the inventors contemplate the mutagenesis of the disclosed polynucleotide sequences to alter one or more properties of the encoded polypeptide, such as the immunogenicity of a polypeptide vaccine. The techniques of site-specific mutagenesis are well-known in the art, and are widely used to create variants of both polypeptides and polynucleotides. For example, site-specific mutagenesis is often used to alter a specific portion of a DNA molecule. In such embodiments, a primer comprising typically about 14 to about 25 nucleotides or so in length is employed, with about 5 to about 10 residues on both sides of the junction of the sequence being altered.

As will be appreciated by those of skill in the art, site-specific mutagenesis techniques have often employed a phage vector that exists in both a single stranded and double stranded form. Typical vectors useful in site-directed mutagenesis include vectors such as the M13 phage. These phage are readily commercially-available and their use is generally well-known to those skilled in the art. Double-stranded plasmids are also routinely employed in site directed mutagenesis that eliminates the step of transferring the gene of interest from a plasmid to a phage.

In general, site-directed mutagenesis in accordance herewith is performed by first obtaining a single-stranded vector or melting apart of two strands of a double-stranded vector that includes within its sequence a DNA sequence that encodes the desired peptide. An oligonucleotide primer bearing the desired mutated sequence is prepared, generally synthetically. This primer is then annealed with the single-stranded vector, and subjected to DNA polymerizing enzymes such as *E. coli* polymerase I Klenow fragment, in order to complete the synthesis of the mutation-bearing strand. Thus, a heteroduplex is formed wherein one strand encodes the original non-mutated sequence and the second strand bears the desired mutation. This heteroduplex vector is then used to transform appropriate cells, such as *E. coli* cells, and clones are selected which include recombinant vectors bearing the mutated sequence arrangement.

The preparation of sequence variants of the selected peptide-encoding DNA segments using site-directed mutagenesis provides a means of producing potentially useful species and is not meant to be limiting as there are other ways in which sequence variants of peptides and the DNA sequences encoding them may be

obtained. For example, recombinant vectors encoding the desired peptide sequence may be treated with mutagenic agents, such as hydroxylamine, to obtain sequence variants. Specific details regarding these methods and protocols are found in the teachings of Maloy *et al.*, 1994; Segal, 1976; Prokop and Bajpai, 1991; Kuby, 1994; and  
5 Maniatis *et al.*, 1982, each incorporated herein by reference, for that purpose.

As used herein, the term "oligonucleotide directed mutagenesis procedure" refers to template-dependent processes and vector-mediated propagation which result in an increase in the concentration of a specific nucleic acid molecule relative to its initial concentration, or in an increase in the concentration of a detectable  
10 signal, such as amplification. As used herein, the term "oligonucleotide directed mutagenesis procedure" is intended to refer to a process that involves the template-dependent extension of a primer molecule. The term template dependent process refers to nucleic acid synthesis of an RNA or a DNA molecule wherein the sequence of the newly synthesized strand of nucleic acid is dictated by the well-known  
15 rules of complementary base pairing (see, for example, Watson, 1987). Typically, vector mediated methodologies involve the introduction of the nucleic acid fragment into a DNA or RNA vector, the clonal amplification of the vector, and the recovery of the amplified nucleic acid fragment. Examples of such methodologies are provided by U. S. Patent No. 4,237,224, specifically incorporated herein by reference in its entirety.

20 In another approach for the production of polypeptide variants of the present invention, recursive sequence recombination, as described in U.S. Patent No. 5,837,458, may be employed. In this approach, iterative cycles of recombination and screening or selection are performed to "evolve" individual polynucleotide variants of the invention having, for example, enhanced immunogenic activity.

25 In other embodiments of the present invention, the polynucleotide sequences provided herein can be advantageously used as probes or primers for nucleic acid hybridization. As such, it is contemplated that nucleic acid segments that comprise a sequence region of at least about 15 nucleotide long contiguous sequence that has the same sequence as, or is complementary to, a 15 nucleotide long contiguous sequence  
30 disclosed herein will find particular utility. Longer contiguous identical or complementary sequences, *e.g.*, those of about 20, 30, 40, 50, 100, 200, 500, 1000

(including all intermediate lengths) and even up to full length sequences will also be of use in certain embodiments.

The ability of such nucleic acid probes to specifically hybridize to a sequence of interest will enable them to be of use in detecting the presence of complementary sequences in a given sample. However, other uses are also envisioned, such as the use of the sequence information for the preparation of mutant species primers, or primers for use in preparing other genetic constructions.

Polynucleotide molecules having sequence regions consisting of contiguous nucleotide stretches of 10-14, 15-20, 30, 50, or even of 100-200 nucleotides or so (including intermediate lengths as well), identical or complementary to a polynucleotide sequence disclosed herein, are particularly contemplated as hybridization probes for use in, *e.g.*, Southern and Northern blotting. This would allow a gene product, or fragment thereof, to be analyzed, both in diverse cell types and also in various bacterial cells. The total size of fragment, as well as the size of the complementary stretch(es), will ultimately depend on the intended use or application of the particular nucleic acid segment. Smaller fragments will generally find use in hybridization embodiments, wherein the length of the contiguous complementary region may be varied, such as between about 15 and about 100 nucleotides, but larger contiguous complementarity stretches may be used, according to the length complementary sequences one wishes to detect.

The use of a hybridization probe of about 15-25 nucleotides in length allows the formation of a duplex molecule that is both stable and selective. Molecules having contiguous complementary sequences over stretches greater than 15 bases in length are generally preferred, though, in order to increase stability and selectivity of the hybrid, and thereby improve the quality and degree of specific hybrid molecules obtained. One will generally prefer to design nucleic acid molecules having gene-complementary stretches of 15 to 25 contiguous nucleotides, or even longer where desired.

Hybridization probes may be selected from any portion of any of the sequences disclosed herein. All that is required is to review the sequences set forth herein, or to any continuous portion of the sequences, from about 15-25 nucleotides in

length up to and including the full length sequence, that one wishes to utilize as a probe or primer. The choice of probe and primer sequences may be governed by various factors. For example, one may wish to employ primers from towards the termini of the total sequence.

5                Small polynucleotide segments or fragments may be readily prepared by, for example, directly synthesizing the fragment by chemical means, as is commonly practiced using an automated oligonucleotide synthesizer. Also, fragments may be obtained by application of nucleic acid reproduction technology, such as the PCR™ technology of U. S. Patent 4,683,202 (incorporated herein by reference), by introducing  
10 selected sequences into recombinant vectors for recombinant production, and by other recombinant DNA techniques generally known to those of skill in the art of molecular biology.

              The nucleotide sequences of the invention may be used for their ability to selectively form duplex molecules with complementary stretches of the entire gene or  
15 gene fragments of interest. Depending on the application envisioned, one will typically desire to employ varying conditions of hybridization to achieve varying degrees of selectivity of probe towards target sequence. For applications requiring high selectivity, one will typically desire to employ relatively stringent conditions to form the hybrids, *e.g.*, one will select relatively low salt and/or high temperature conditions, such as  
20 provided by a salt concentration of from about 0.02 M to about 0.15 M salt at temperatures of from about 50°C to about 70°C. Such selective conditions tolerate little, if any, mismatch between the probe and the template or target strand, and would be particularly suitable for isolating related sequences.

              Of course, for some applications, for example, where one desires to  
25 prepare mutants employing a mutant primer strand hybridized to an underlying template, less stringent (reduced stringency) hybridization conditions will typically be needed in order to allow formation of the heteroduplex. In these circumstances, one may desire to employ salt conditions such as those of from about 0.15 M to about 0.9 M salt, at temperatures ranging from about 20°C to about 55°C. Cross-hybridizing species  
30 can thereby be readily identified as positively hybridizing signals with respect to control hybridizations. In any case, it is generally appreciated that conditions can be rendered

more stringent by the addition of increasing amounts of formamide, which serves to destabilize the hybrid duplex in the same manner as increased temperature. Thus, hybridization conditions can be readily manipulated, and thus will generally be a method of choice depending on the desired results.

5           According to another embodiment of the present invention, polynucleotide compositions comprising antisense oligonucleotides are provided. Antisense oligonucleotides have been demonstrated to be effective and targeted inhibitors of protein synthesis, and, consequently, provide a therapeutic approach by which a disease can be treated by inhibiting the synthesis of proteins that contribute to  
10 the disease. The efficacy of antisense oligonucleotides for inhibiting protein synthesis is well established. For example, the synthesis of polygalacturonase and the muscarine type 2 acetylcholine receptor are inhibited by antisense oligonucleotides directed to their respective mRNA sequences (U. S. Patent 5,739,119 and U. S. Patent 5,759,829). Further, examples of antisense inhibition have been demonstrated with the nuclear  
15 protein cyclin, the multiple drug resistance gene (MDG1), ICAM-1, E-selectin, STK-1, striatal GABA<sub>A</sub> receptor and human EGF (Jaskulski *et al.*, Science. 1988 Jun 10;240(4858):1544-6; Vasanthakumar and Ahmed, Cancer Commun. 1989;1(4):225-32; Peris *et al.*, Brain Res Mol Brain Res. 1998 Jun 15;57(2):310-20; U. S. Patent 5,801,154; U.S. Patent 5,789,573; U. S. Patent 5,718,709 and U.S. Patent 5,610,288).  
20 Antisense constructs have also been described that inhibit and can be used to treat a variety of abnormal cellular proliferations, *e.g.* cancer (U. S. Patent 5,747,470; U. S. Patent 5,591,317 and U. S. Patent 5,783,683).

          Therefore, in certain embodiments, the present invention provides oligonucleotide sequences that comprise all, or a portion of, any sequence that is  
25 capable of specifically binding to polynucleotide sequence described herein, or a complement thereof. In one embodiment, the antisense oligonucleotides comprise DNA or derivatives thereof. In another embodiment, the oligonucleotides comprise RNA or derivatives thereof. In a third embodiment, the oligonucleotides are modified DNAs comprising a phosphorothioated modified backbone. In a fourth embodiment, the  
30 oligonucleotide sequences comprise peptide nucleic acids or derivatives thereof. In each case, preferred compositions comprise a sequence region that is complementary,

and more preferably substantially-complementary, and even more preferably, completely complementary to one or more portions of polynucleotides disclosed herein. Selection of antisense compositions specific for a given gene sequence is based upon analysis of the chosen target sequence and determination of secondary structure,  $T_m$ ,  
5 binding energy, and relative stability. Antisense compositions may be selected based upon their relative inability to form dimers, hairpins, or other secondary structures that would reduce or prohibit specific binding to the target mRNA in a host cell. Highly preferred target regions of the mRNA, are those which are at or near the AUG translation initiation codon, and those sequences which are substantially complementary  
10 to 5' regions of the mRNA. These secondary structure analyses and target site selection considerations can be performed, for example, using v.4 of the OLIGO primer analysis software and/or the BLASTN 2.0.5 algorithm software (Altschul *et al.*, Nucleic Acids Res. 1997, 25(17):3389-402).

The use of an antisense delivery method employing a short peptide  
15 vector, termed MPG (27 residues), is also contemplated. The MPG peptide contains a hydrophobic domain derived from the fusion sequence of HIV gp41 and a hydrophilic domain from the nuclear localization sequence of SV40 T-antigen (Morris *et al.*, Nucleic Acids Res. 1997 Jul 15;25(14):2730-6). It has been demonstrated that several molecules of the MPG peptide coat the antisense oligonucleotides and can be delivered  
20 into cultured mammalian cells in less than 1 hour with relatively high efficiency (90%). Further, the interaction with MPG strongly increases both the stability of the oligonucleotide to nuclease and the ability to cross the plasma membrane.

According to another embodiment of the invention, the polynucleotide compositions described herein are used in the design and preparation of ribozyme  
25 molecules for inhibiting expression of the tumor polypeptides and proteins of the present invention in tumor cells. Ribozymes are RNA-protein complexes that cleave nucleic acids in a site-specific fashion. Ribozymes have specific catalytic domains that possess endonuclease activity (Kim and Cech, Proc Natl Acad Sci U S A. 1987 Dec;84(24):8788-92; Forster and Symons, Cell. 1987 Apr 24;49(2):211-20). For  
30 example, a large number of ribozymes accelerate phosphoester transfer reactions with a high degree of specificity, often cleaving only one of several phosphoesters in an

oligonucleotide substrate (Cech *et al.*, Cell. 1981 Dec;27(3 Pt 2):487-96; Michel and Westhof, J Mol Biol. 1990 Dec 5;216(3):585-610; Reinhold-Hurek and Shub, Nature. 1992 May 14;357(6374):173-6). This specificity has been attributed to the requirement that the substrate bind via specific base-pairing interactions to the internal guide  
5 sequence ("IGS") of the ribozyme prior to chemical reaction.

Six basic varieties of naturally-occurring enzymatic RNAs are known presently. Each can catalyze the hydrolysis of RNA phosphodiester bonds *in trans* (and thus can cleave other RNA molecules) under physiological conditions. In general, enzymatic nucleic acids act by first binding to a target RNA. Such binding occurs  
10 through the target binding portion of a enzymatic nucleic acid which is held in close proximity to an enzymatic portion of the molecule that acts to cleave the target RNA. Thus, the enzymatic nucleic acid first recognizes and then binds a target RNA through complementary base-pairing, and once bound to the correct site, acts enzymatically to cut the target RNA. Strategic cleavage of such a target RNA will destroy its ability to  
15 direct synthesis of an encoded protein. After an enzymatic nucleic acid has bound and cleaved its RNA target, it is released from that RNA to search for another target and can repeatedly bind and cleave new targets.

The enzymatic nature of a ribozyme is advantageous over many technologies, such as antisense technology (where a nucleic acid molecule simply binds  
20 to a nucleic acid target to block its translation) since the concentration of ribozyme necessary to affect a therapeutic treatment is lower than that of an antisense oligonucleotide. This advantage reflects the ability of the ribozyme to act enzymatically. Thus, a single ribozyme molecule is able to cleave many molecules of target RNA. In addition, the ribozyme is a highly specific inhibitor, with the specificity  
25 of inhibition depending not only on the base pairing mechanism of binding to the target RNA, but also on the mechanism of target RNA cleavage. Single mismatches, or base-substitutions, near the site of cleavage can completely eliminate catalytic activity of a ribozyme. Similar mismatches in antisense molecules do not prevent their action (Woelf *et al.*, Proc Natl Acad Sci U S A. 1992 Aug 15;89(16):7305-9). Thus, the  
30 specificity of action of a ribozyme is greater than that of an antisense oligonucleotide binding the same RNA site.

The enzymatic nucleic acid molecule may be formed in a hammerhead, hairpin, a hepatitis  $\delta$  virus, group I intron or RNaseP RNA (in association with an RNA guide sequence) or Neurospora VS RNA motif. Examples of hammerhead motifs are described by Rossi *et al.* Nucleic Acids Res. 1992 Sep 11;20(17):4559-65. Examples of hairpin motifs are described by Hampel *et al.* (Eur. Pat. Appl. Publ. No. EP 0360257), Hampel and Tritz, Biochemistry 1989 Jun 13;28(12):4929-33; Hampel *et al.*, Nucleic Acids Res. 1990 Jan 25;18(2):299-304 and U. S. Patent 5,631,359. An example of the hepatitis  $\delta$  virus motif is described by Perrotta and Been, Biochemistry. 1992 Dec 1;31(47):11843-52; an example of the RNaseP motif is described by Guerrier-Takada *et al.*, Cell. 1983 Dec;35(3 Pt 2):849-57; Neurospora VS RNA ribozyme motif is described by Collins (Saville and Collins, Cell. 1990 May 18;61(4):685-96; Saville and Collins, Proc Natl Acad Sci U S A. 1991 Oct 1;88(19):8826-30; Collins and Olive, Biochemistry. 1993 Mar 23;32(11):2795-9); and an example of the Group I intron is described in (U. S. Patent 4,987,071). All that is important in an enzymatic nucleic acid molecule of this invention is that it has a specific substrate binding site which is complementary to one or more of the target gene RNA regions, and that it have nucleotide sequences within or surrounding that substrate binding site which impart an RNA cleaving activity to the molecule. Thus the ribozyme constructs need not be limited to specific motifs mentioned herein.

Ribozymes may be designed as described in Int. Pat. Appl. Publ. No. WO 93/23569 and Int. Pat. Appl. Publ. No. WO 94/02595, each specifically incorporated herein by reference) and synthesized to be tested *in vitro* and *in vivo*, as described. Such ribozymes can also be optimized for delivery. While specific examples are provided, those in the art will recognize that equivalent RNA targets in other species can be utilized when necessary.

Ribozyme activity can be optimized by altering the length of the ribozyme binding arms, or chemically synthesizing ribozymes with modifications that prevent their degradation by serum ribonucleases (see *e.g.*, Int. Pat. Appl. Publ. No. WO 92/07065; Int. Pat. Appl. Publ. No. WO 93/15187; Int. Pat. Appl. Publ. No. WO 91/03162; Eur. Pat. Appl. Publ. No. 92110298.4; U. S. Patent 5,334,711; and Int. Pat. Appl. Publ. No. WO 94/13688, which describe various chemical modifications that can

be made to the sugar moieties of enzymatic RNA molecules), modifications which enhance their efficacy in cells, and removal of stem II bases to shorten RNA synthesis times and reduce chemical requirements.

Sullivan *et al.* (Int. Pat. Appl. Publ. No. WO 94/02595) describes the  
5 general methods for delivery of enzymatic RNA molecules. Ribozymes may be administered to cells by a variety of methods known to those familiar to the art, including, but not restricted to, encapsulation in liposomes, by iontophoresis, or by incorporation into other vehicles, such as hydrogels, cyclodextrins, biodegradable nanocapsules, and bioadhesive microspheres. For some indications, ribozymes may be  
10 directly delivered *ex vivo* to cells or tissues with or without the aforementioned vehicles. Alternatively, the RNA/vehicle combination may be locally delivered by direct inhalation, by direct injection or by use of a catheter, infusion pump or stent. Other routes of delivery include, but are not limited to, intravascular, intramuscular, subcutaneous or joint injection, aerosol inhalation, oral (tablet or pill form), topical,  
15 systemic, ocular, intraperitoneal and/or intrathecal delivery. More detailed descriptions of ribozyme delivery and administration are provided in Int. Pat. Appl. Publ. No. WO 94/02595 and Int. Pat. Appl. Publ. No. WO 93/23569, each specifically incorporated herein by reference.

Another means of accumulating high concentrations of a ribozyme(s)  
20 within cells is to incorporate the ribozyme-encoding sequences into a DNA expression vector. Transcription of the ribozyme sequences are driven from a promoter for eukaryotic RNA polymerase I (pol I), RNA polymerase II (pol II), or RNA polymerase III (pol III). Transcripts from pol II or pol III promoters will be expressed at high levels in all cells; the levels of a given pol II promoter in a given cell type will depend on the  
25 nature of the gene regulatory sequences (enhancers, silencers, *etc.*) present nearby. Prokaryotic RNA polymerase promoters may also be used, providing that the prokaryotic RNA polymerase enzyme is expressed in the appropriate cells. Ribozymes expressed from such promoters have been shown to function in mammalian cells. Such transcription units can be incorporated into a variety of vectors for introduction into  
30 mammalian cells, including but not restricted to, plasmid DNA vectors, viral DNA

vectors (such as adenovirus or adeno-associated vectors), or viral RNA vectors (such as retroviral, semliki forest virus, sindbis virus vectors).

In another embodiment of the invention, peptide nucleic acids (PNAs) compositions are provided. PNA is a DNA mimic in which the nucleobases are attached to a pseudopeptide backbone (Good and Nielsen, *Antisense Nucleic Acid Drug Dev.* 1997 7(4) 431-37). PNA is able to be utilized in a number methods that traditionally have used RNA or DNA. Often PNA sequences perform better in techniques than the corresponding RNA or DNA sequences and have utilities that are not inherent to RNA or DNA. A review of PNA including methods of making, characteristics of, and methods of using, is provided by Corey (*Trends Biotechnol* 1997 Jun;15(6):224-9). As such, in certain embodiments, one may prepare PNA sequences that are complementary to one or more portions of the ACE mRNA sequence, and such PNA compositions may be used to regulate, alter, decrease, or reduce the translation of ACE-specific mRNA, and thereby alter the level of ACE activity in a host cell to which such PNA compositions have been administered.

PNAs have 2-aminoethyl-glycine linkages replacing the normal phosphodiester backbone of DNA (Nielsen *et al.*, *Science* 1991 Dec 6;254(5037):1497-500; Hanvey *et al.*, *Science*. 1992 Nov 27;258(5087):1481-5; Hyrup and Nielsen, *Bioorg Med Chem.* 1996 Jan;4(1):5-23). This chemistry has three important consequences: firstly, in contrast to DNA or phosphorothioate oligonucleotides, PNAs are neutral molecules; secondly, PNAs are achiral, which avoids the need to develop a stereoselective synthesis; and thirdly, PNA synthesis uses standard Boc or Fmoc protocols for solid-phase peptide synthesis, although other methods, including a modified Merrifield method, have been used.

PNA monomers or ready-made oligomers are commercially available from PerSeptive Biosystems (Framingham, MA). PNA syntheses by either Boc or Fmoc protocols are straightforward using manual or automated protocols (Norton *et al.*, *Bioorg Med Chem.* 1995 Apr;3(4):437-45). The manual protocol lends itself to the production of chemically modified PNAs or the simultaneous synthesis of families of closely related PNAs.

As with peptide synthesis, the success of a particular PNA synthesis will depend on the properties of the chosen sequence. For example, while in theory PNAs can incorporate any combination of nucleotide bases, the presence of adjacent purines can lead to deletions of one or more residues in the product. In expectation of this  
5 difficulty, it is suggested that, in producing PNAs with adjacent purines, one should repeat the coupling of residues likely to be added inefficiently. This should be followed by the purification of PNAs by reverse-phase high-pressure liquid chromatography, providing yields and purity of product similar to those observed during the synthesis of peptides.

10 Modifications of PNAs for a given application may be accomplished by coupling amino acids during solid-phase synthesis or by attaching compounds that contain a carboxylic acid group to the exposed N-terminal amine. Alternatively, PNAs can be modified after synthesis by coupling to an introduced lysine or cysteine. The ease with which PNAs can be modified facilitates optimization for better solubility or  
15 for specific functional requirements. Once synthesized, the identity of PNAs and their derivatives can be confirmed by mass spectrometry. Several studies have made and utilized modifications of PNAs (for example, Norton *et al.*, Bioorg Med Chem. 1995 Apr;3(4):437-45; Petersen *et al.*, J Pept Sci. 1995 May-Jun;1(3):175-83; Orum *et al.*, Biotechniques. 1995 Sep;19(3):472-80; Footer *et al.*, Biochemistry. 1996 Aug  
20 20;35(33):10673-9; Griffith *et al.*, Nucleic Acids Res. 1995 Aug 11;23(15):3003-8; Pardridge *et al.*, Proc Natl Acad Sci U S A. 1995 Jun 6;92(12):5592-6; Boffa *et al.*, Proc Natl Acad Sci U S A. 1995 Mar 14;92(6):1901-5; Gambacorti-Passerini *et al.*, Blood. 1996 Aug 15;88(4):1411-7; Armitage *et al.*, Proc Natl Acad Sci U S A. 1997 Nov 11;94(23):12320-5; Seeger *et al.*, Biotechniques. 1997 Sep;23(3):512-7). U.S.  
25 Patent No. 5,700,922 discusses PNA-DNA-PNA chimeric molecules and their uses in diagnostics, modulating protein in organisms, and treatment of conditions susceptible to therapeutics.

Methods of characterizing the antisense binding properties of PNAs are discussed in Rose (Anal Chem. 1993 Dec 15;65(24):3545-9) and Jensen *et al.*  
30 (Biochemistry. 1997 Apr 22;36(16):5072-7). Rose uses capillary gel electrophoresis to determine binding of PNAs to their complementary oligonucleotide, measuring the

relative binding kinetics and stoichiometry. Similar types of measurements were made by Jensen *et al.* using BLAcore™ technology.

Other applications of PNAs that have been described and will be apparent to the skilled artisan include use in DNA strand invasion, antisense inhibition, 5 mutational analysis, enhancers of transcription, nucleic acid purification, isolation of transcriptionally active genes, blocking of transcription factor binding, genome cleavage, biosensors, *in situ* hybridization, and the like.

#### Polynucleotide Identification, Characterization and Expression

Polynucleotides compositions of the present invention may be identified, 10 prepared and/or manipulated using any of a variety of well established techniques (see generally, Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratories, Cold Spring Harbor, NY, 1989, and other like references). For example, a polynucleotide may be identified, as described in more detail below, by screening a microarray of cDNAs for tumor-associated expression (*i.e.*, expression that 15 is at least two fold greater in a tumor than in normal tissue, as determined using a representative assay provided herein). Such screens may be performed, for example, using the microarray technology of Affymetrix, Inc. (Santa Clara, CA) according to the manufacturer's instructions (and essentially as described by Schena et al., *Proc. Natl. Acad. Sci. USA* 93:10614-10619, 1996 and Heller et al., *Proc. Natl. Acad. Sci. USA* 20 94:2150-2155, 1997). Alternatively, polynucleotides may be amplified from cDNA prepared from cells expressing the proteins described herein, such as tumor cells.

Many template dependent processes are available to amplify a target sequences of interest present in a sample. One of the best known amplification methods is the polymerase chain reaction (PCR™) which is described in detail in U.S. Patent 25 Nos. 4,683,195, 4,683,202 and 4,800,159, each of which is incorporated herein by reference in its entirety. Briefly, in PCR™, two primer sequences are prepared which are complementary to regions on opposite complementary strands of the target sequence. An excess of deoxynucleoside triphosphates is added to a reaction mixture along with a DNA polymerase (*e.g.*, *Taq* polymerase). If the target sequence is present 30 in a sample, the primers will bind to the target and the polymerase will cause the

primers to be extended along the target sequence by adding on nucleotides. By raising and lowering the temperature of the reaction mixture, the extended primers will dissociate from the target to form reaction products, excess primers will bind to the target and to the reaction product and the process is repeated. Preferably reverse transcription and PCR<sup>TM</sup> amplification procedure may be performed in order to quantify the amount of mRNA amplified. Polymerase chain reaction methodologies are well known in the art.

Any of a number of other template dependent processes, many of which are variations of the PCR<sup>TM</sup> amplification technique, are readily known and available in the art. Illustratively, some such methods include the ligase chain reaction (referred to as LCR), described, for example, in Eur. Pat. Appl. Publ. No. 320,308 and U.S. Patent No. 4,883,750; Qbeta Replicase, described in PCT Intl. Pat. Appl. Publ. No. PCT/US87/00880; Strand Displacement Amplification (SDA) and Repair Chain Reaction (RCR). Still other amplification methods are described in Great Britain Pat. Appl. No. 2 202 328, and in PCT Intl. Pat. Appl. Publ. No. PCT/US89/01025. Other nucleic acid amplification procedures include transcription-based amplification systems (TAS) (PCT Intl. Pat. Appl. Publ. No. WO 88/10315), including nucleic acid sequence based amplification (NASBA) and 3SR. Eur. Pat. Appl. Publ. No. 329,822 describes a nucleic acid amplification process involving cyclically synthesizing single-stranded RNA ("ssRNA"), ssDNA, and double-stranded DNA (dsDNA). PCT Intl. Pat. Appl. Publ. No. WO 89/06700 describes a nucleic acid sequence amplification scheme based on the hybridization of a promoter/primer sequence to a target single-stranded DNA ("ssDNA") followed by transcription of many RNA copies of the sequence. Other amplification methods such as "RACE" (Frohman, 1990), and "one-sided PCR" (Ohara, 1989) are also well-known to those of skill in the art.

An amplified portion of a polynucleotide of the present invention may be used to isolate a full length gene from a suitable library (e.g., a tumor cDNA library) using well known techniques. Within such techniques, a library (cDNA or genomic) is screened using one or more polynucleotide probes or primers suitable for amplification. Preferably, a library is size-selected to include larger molecules. Random primed

libraries may also be preferred for identifying 5' and upstream regions of genes. Genomic libraries are preferred for obtaining introns and extending 5' sequences.

For hybridization techniques, a partial sequence may be labeled (*e.g.*, by nick-translation or end-labeling with  $^{32}\text{P}$ ) using well known techniques. A bacterial or bacteriophage library is then generally screened by hybridizing filters containing denatured bacterial colonies (or lawns containing phage plaques) with the labeled probe (see Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratories, Cold Spring Harbor, NY, 1989). Hybridizing colonies or plaques are selected and expanded, and the DNA is isolated for further analysis. cDNA clones may be analyzed to determine the amount of additional sequence by, for example, PCR using a primer from the partial sequence and a primer from the vector. Restriction maps and partial sequences may be generated to identify one or more overlapping clones. The complete sequence may then be determined using standard techniques, which may involve generating a series of deletion clones. The resulting overlapping sequences can then be assembled into a single contiguous sequence. A full length cDNA molecule can be generated by ligating suitable fragments, using well known techniques.

Alternatively, amplification techniques, such as those described above, can be useful for obtaining a full length coding sequence from a partial cDNA sequence. One such amplification technique is inverse PCR (see Triglia et al., *Nucl. Acids Res.* 16:8186, 1988), which uses restriction enzymes to generate a fragment in the known region of the gene. The fragment is then circularized by intramolecular ligation and used as a template for PCR with divergent primers derived from the known region. Within an alternative approach, sequences adjacent to a partial sequence may be retrieved by amplification with a primer to a linker sequence and a primer specific to a known region. The amplified sequences are typically subjected to a second round of amplification with the same linker primer and a second primer specific to the known region. A variation on this procedure, which employs two primers that initiate extension in opposite directions from the known sequence, is described in WO 96/38591. Another such technique is known as "rapid amplification of cDNA ends" or RACE. This technique involves the use of an internal primer and an external primer, which hybridizes to a polyA region or vector sequence, to identify sequences that are 5'

and 3' of a known sequence. Additional techniques include capture PCR (Lagerstrom et al., *PCR Methods Applic.* 1:111-19, 1991) and walking PCR (Parker et al., *Nucl. Acids. Res.* 19:3055-60, 1991). Other methods employing amplification may also be employed to obtain a full length cDNA sequence.

5                   In certain instances, it is possible to obtain a full length cDNA sequence by analysis of sequences provided in an expressed sequence tag (EST) database, such as that available from GenBank. Searches for overlapping ESTs may generally be performed using well known programs (e.g., NCBI BLAST searches), and such ESTs may be used to generate a contiguous full length sequence. Full length DNA sequences  
10 may also be obtained by analysis of genomic fragments.

                  In other embodiments of the invention, polynucleotide sequences or fragments thereof which encode polypeptides of the invention, or fusion proteins or functional equivalents thereof, may be used in recombinant DNA molecules to direct expression of a polypeptide in appropriate host cells. Due to the inherent degeneracy of  
15 the genetic code, other DNA sequences that encode substantially the same or a functionally equivalent amino acid sequence may be produced and these sequences may be used to clone and express a given polypeptide.

                  As will be understood by those of skill in the art, it may be advantageous in some instances to produce polypeptide-encoding nucleotide sequences possessing  
20 non-naturally occurring codons. For example, codons preferred by a particular prokaryotic or eukaryotic host can be selected to increase the rate of protein expression or to produce a recombinant RNA transcript having desirable properties, such as a half-life which is longer than that of a transcript generated from the naturally occurring sequence.

25                   Moreover, the polynucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter polypeptide encoding sequences for a variety of reasons, including but not limited to, alterations which modify the cloning, processing, and/or expression of the gene product. For example, DNA shuffling by random fragmentation and PCR reassembly of gene  
30 fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. In addition, site-directed mutagenesis may be used to insert new restriction

sites, alter glycosylation patterns, change codon preference, produce splice variants, or introduce mutations, and so forth.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences may be ligated to a heterologous sequence to  
5 encode a fusion protein. For example, to screen peptide libraries for inhibitors of polypeptide activity, it may be useful to encode a chimeric protein that can be recognized by a commercially available antibody. A fusion protein may also be engineered to contain a cleavage site located between the polypeptide-encoding sequence and the heterologous protein sequence, so that the polypeptide may be cleaved  
10 and purified away from the heterologous moiety.

Sequences encoding a desired polypeptide may be synthesized, in whole or in part, using chemical methods well known in the art (see Caruthers, M. H. et al. (1980) *Nucl. Acids Res. Symp. Ser.* 215-223, Horn, T. et al. (1980) *Nucl. Acids Res. Symp. Ser.* 225-232). Alternatively, the protein itself may be produced using chemical  
15 methods to synthesize the amino acid sequence of a polypeptide, or a portion thereof. For example, peptide synthesis can be performed using various solid-phase techniques (Roberge, J. Y. et al. (1995) *Science* 269:202-204) and automated synthesis may be achieved, for example, using the ABI 431A Peptide Synthesizer (Perkin Elmer, Palo Alto, CA).

20 A newly synthesized peptide may be substantially purified by preparative high performance liquid chromatography (e.g., Creighton, T. (1983) *Proteins, Structures and Molecular Principles*, WH Freeman and Co., New York, N.Y.) or other comparable techniques available in the art. The composition of the synthetic peptides may be confirmed by amino acid analysis or sequencing (e.g., the Edman degradation  
25 procedure). Additionally, the amino acid sequence of a polypeptide, or any part thereof, may be altered during direct synthesis and/or combined using chemical methods with sequences from other proteins, or any part thereof, to produce a variant polypeptide.

In order to express a desired polypeptide, the nucleotide sequences encoding the polypeptide, or functional equivalents, may be inserted into appropriate  
30 expression vector, i.e., a vector which contains the necessary elements for the transcription and translation of the inserted coding sequence. Methods which are well

known to those skilled in the art may be used to construct expression vectors containing sequences encoding a polypeptide of interest and appropriate transcriptional and translational control elements. These methods include *in vitro* recombinant DNA techniques, synthetic techniques, and *in vivo* genetic recombination. Such techniques  
5 are described, for example, in Sambrook, J. et al. (1989) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Press, Plainview, N.Y., and Ausubel, F. M. et al. (1989) Current Protocols in Molecular Biology, John Wiley & Sons, New York. N.Y.

A variety of expression vector/host systems may be utilized to contain  
10 and express polynucleotide sequences. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with virus expression vectors (*e.g.*, baculovirus); plant cell systems transformed with virus expression vectors (*e.g.*, cauliflower mosaic virus,  
15 CaMV; tobacco mosaic virus, TMV) or with bacterial expression vectors (*e.g.*, Ti or pBR322 plasmids); or animal cell systems.

The "control elements" or "regulatory sequences" present in an expression vector are those non-translated regions of the vector--enhancers, promoters, 5' and 3' untranslated regions--which interact with host cellular proteins to carry out  
20 transcription and translation. Such elements may vary in their strength and specificity. Depending on the vector system and host utilized, any number of suitable transcription and translation elements, including constitutive and inducible promoters, may be used. For example, when cloning in bacterial systems, inducible promoters such as the hybrid lacZ promoter of the PBLUESCRIPT phagemid (Stratagene, La Jolla, Calif.) or  
25 PSPORT1 plasmid (Gibco BRL, Gaithersburg, MD) and the like may be used. In mammalian cell systems, promoters from mammalian genes or from mammalian viruses are generally preferred. If it is necessary to generate a cell line that contains multiple copies of the sequence encoding a polypeptide, vectors based on SV40 or EBV may be advantageously used with an appropriate selectable marker.

30 In bacterial systems, any of a number of expression vectors may be selected depending upon the use intended for the expressed polypeptide. For example,

when large quantities are needed, for example for the induction of antibodies, vectors which direct high level expression of fusion proteins that are readily purified may be used. Such vectors include, but are not limited to, the multifunctional *E. coli* cloning and expression vectors such as BLUESCRIPT (Stratagene), in which the sequence  
5 encoding the polypeptide of interest may be ligated into the vector in frame with sequences for the amino-terminal Met and the subsequent 7 residues of .beta.-galactosidase so that a hybrid protein is produced; pIN vectors (Van Heeke, G. and S. M. Schuster (1989) *J. Biol. Chem.* 264:5503-5509); and the like. pGEX Vectors (Promega, Madison, Wis.) may also be used to express foreign polypeptides as fusion  
10 proteins with glutathione S-transferase (GST). In general, such fusion proteins are soluble and can easily be purified from lysed cells by adsorption to glutathione-agarose beads followed by elution in the presence of free glutathione. Proteins made in such systems may be designed to include heparin, thrombin, or factor XA protease cleavage sites so that the cloned polypeptide of interest can be released from the GST moiety at  
15 will.

In the yeast, *Saccharomyces cerevisiae*, a number of vectors containing constitutive or inducible promoters such as alpha factor, alcohol oxidase, and PGH may be used. For reviews, see Ausubel et al. (supra) and Grant et al. (1987) *Methods Enzymol.* 153:516-544.

20 In cases where plant expression vectors are used, the expression of sequences encoding polypeptides may be driven by any of a number of promoters. For example, viral promoters such as the 35S and 19S promoters of CaMV may be used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) *EMBO J.* 6:307-311. Alternatively, plant promoters such as the small subunit of  
25 RUBISCO or heat shock promoters may be used (Coruzzi, G. et al. (1984) *EMBO J.* 3:1671-1680; Broglie, R. et al. (1984) *Science* 224:838-843; and Winter, J. et al. (1991) *Results Probl. Cell Differ.* 17:85-105). These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. Such techniques are described in a number of generally available reviews (see, for example, Hobbs, S. or  
30 Murry, L. E. in McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York, N.Y.; pp. 191-196).

An insect system may also be used to express a polypeptide of interest. For example, in one such system, *Autographa californica* nuclear polyhedrosis virus (AcNPV) is used as a vector to express foreign genes in *Spodoptera frugiperda* cells or in *Trichoplusia* larvae. The sequences encoding the polypeptide may be cloned into a non-essential region of the virus, such as the polyhedrin gene, and placed under control of the polyhedrin promoter. Successful insertion of the polypeptide-encoding sequence will render the polyhedrin gene inactive and produce recombinant virus lacking coat protein. The recombinant viruses may then be used to infect, for example, *S. frugiperda* cells or *Trichoplusia* larvae in which the polypeptide of interest may be expressed (Engelhard, E. K. et al. (1994) *Proc. Natl. Acad. Sci.* 91 :3224-3227).

In mammalian host cells, a number of viral-based expression systems are generally available. For example, in cases where an adenovirus is used as an expression vector, sequences encoding a polypeptide of interest may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain a viable virus which is capable of expressing the polypeptide in infected host cells (Logan, J. and Shenk, T. (1984) *Proc. Natl. Acad. Sci.* 81:3655-3659). In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells.

Specific initiation signals may also be used to achieve more efficient translation of sequences encoding a polypeptide of interest. Such signals include the ATG initiation codon and adjacent sequences. In cases where sequences encoding the polypeptide, its initiation codon, and upstream sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a portion thereof, is inserted, exogenous translational control signals including the ATG initiation codon should be provided. Furthermore, the initiation codon should be in the correct reading frame to ensure translation of the entire insert. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers which are

appropriate for the particular cell system which is used, such as those described in the literature (Scharf, D. et al. (1994) *Results Probl. Cell Differ.* 20:125-162).

In addition, a host cell strain may be chosen for its ability to modulate the expression of the inserted sequences or to process the expressed protein in the  
5 desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" form of the protein may also be used to facilitate correct insertion, folding and/or function. Different host cells such as CHO, COS, HeLa, MDCK, HEK293, and WI38, which have specific cellular machinery  
10 and characteristic mechanisms for such post-translational activities, may be chosen to ensure the correct modification and processing of the foreign protein.

For long-term, high-yield production of recombinant proteins, stable expression is generally preferred. For example, cell lines which stably express a polynucleotide of interest may be transformed using expression vectors which may  
15 contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for 1-2 days in an enriched media before they are switched to selective media. The purpose of the selectable marker is to confer resistance to selection, and its presence allows growth and recovery of cells which  
20 successfully express the introduced sequences. Resistant clones of stably transformed cells may be proliferated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase (Wigler, M. et al. (1977) *Cell* 11:223-32) and adenine phosphoribosyltransferase  
25 (Lowy, I. et al. (1990) *Cell* 22:817-23) genes which can be employed in tk.sup.- or apt.sup.- cells, respectively. Also, antimetabolite, antibiotic or herbicide resistance can be used as the basis for selection; for example, dhfr which confers resistance to methotrexate (Wigler, M. et al. (1980) *Proc. Natl. Acad. Sci.* 77:3567-70); npt, which confers resistance to the aminoglycosides, neomycin and G-418 (Colbere-Garapin, F. et  
30 al (1981) *J. Mol. Biol.* 150:1-14); and als or pat, which confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively (Murry, *supra*).

Additional selectable genes have been described, for example, *trpB*, which allows cells to utilize indole in place of tryptophan, or *hisD*, which allows cells to utilize histinol in place of histidine (Hartman, S. C. and R. C. Mulligan (1988) *Proc. Natl. Acad. Sci.* 85:8047-51). The use of visible markers has gained popularity with such markers as  
5 anthocyanins, beta-glucuronidase and its substrate GUS, and luciferase and its substrate luciferin, being widely used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes, C. A. et al. (1995) *Methods Mol. Biol.* 55:121-131).

Although the presence/absence of marker gene expression suggests that  
10 the gene of interest is also present, its presence and expression may need to be confirmed. For example, if the sequence encoding a polypeptide is inserted within a marker gene sequence, recombinant cells containing sequences can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a polypeptide-encoding sequence under the control of a single promoter.  
15 Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

Alternatively, host cells that contain and express a desired polynucleotide sequence may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-  
20 RNA hybridizations and protein bioassay or immunoassay techniques which include, for example, membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein.

A variety of protocols for detecting and measuring the expression of polynucleotide-encoded products, using either polyclonal or monoclonal antibodies  
25 specific for the product are known in the art. Examples include enzyme-linked immunosorbent assay (ELISA), radioimmunoassay (RIA), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on a given polypeptide may be preferred for some applications, but a competitive binding assay may also be employed.  
30 These and other assays are described, among other places, in Hampton, R. et al. (1990;

Serological Methods, a Laboratory Manual, APS Press, St Paul, Minn.) and Maddox, D. E. et al. (1983; *J. Exp. Med.* 158:1211-1216).

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means  
5 for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides include oligolabeling, nick translation, end-labeling or PCR amplification using a labeled nucleotide. Alternatively, the sequences, or any portions thereof may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA  
10 probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits. Suitable reporter molecules or labels, which may be used include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

15 Host cells transformed with a polynucleotide sequence of interest may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a recombinant cell may be secreted or contained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides of the  
20 invention may be designed to contain signal sequences which direct secretion of the encoded polypeptide through a prokaryotic or eukaryotic cell membrane. Other recombinant constructions may be used to join sequences encoding a polypeptide of interest to nucleotide sequence encoding a polypeptide domain which will facilitate purification of soluble proteins. Such purification facilitating domains include, but are  
25 not limited to, metal chelating peptides such as histidine-tryptophan modules that allow purification on immobilized metals, protein A domains that allow purification on immobilized immunoglobulin, and the domain utilized in the FLAGS extension/affinity purification system (Immunex Corp., Seattle, Wash.). The inclusion of cleavable linker sequences such as those specific for Factor XA or enterokinase (Invitrogen, San Diego,  
30 Calif.) between the purification domain and the encoded polypeptide may be used to facilitate purification. One such expression vector provides for expression of a fusion

protein containing a polypeptide of interest and a nucleic acid encoding 6 histidine residues preceding a thioredoxin or an enterokinase cleavage site. The histidine residues facilitate purification on IMIAC (immobilized metal ion affinity chromatography) as described in Porath, J. et al. (1992, *Prot. Exp. Purif.* 3:263-281) while the enterokinase  
5 cleavage site provides a means for purifying the desired polypeptide from the fusion protein. A discussion of vectors which contain fusion proteins is provided in Kroll, D. J. et al. (1993; *DNA Cell Biol.* 12:441-453).

In addition to recombinant production methods, polypeptides of the invention, and fragments thereof, may be produced by direct peptide synthesis using  
10 solid-phase techniques (Merrifield J. (1963) *J. Am. Chem. Soc.* 85:2149-2154). Protein synthesis may be performed using manual techniques or by automation. Automated synthesis may be achieved, for example, using Applied Biosystems 431A Peptide Synthesizer (Perkin Elmer). Alternatively, various fragments may be chemically synthesized separately and combined using chemical methods to produce the full length  
15 molecule.

#### Antibody Compositions, Fragments Thereof and Other Binding Agents

According to another aspect, the present invention further provides binding agents, such as antibodies and antigen-binding fragments thereof, that exhibit immunological binding to a tumor polypeptide disclosed herein, or to a portion, variant  
20 or derivative thereof. An antibody, or antigen-binding fragment thereof, is said to "specifically bind," "immunologically bind," and/or is "immunologically reactive" to a polypeptide of the invention if it reacts at a detectable level (within, for example, an ELISA assay) with the polypeptide, and does not react detectably with unrelated polypeptides under similar conditions.

25 Immunological binding, as used in this context, generally refers to the non-covalent interactions of the type which occur between an immunoglobulin molecule and an antigen for which the immunoglobulin is specific. The strength, or affinity of immunological binding interactions can be expressed in terms of the dissociation constant ( $K_d$ ) of the interaction, wherein a smaller  $K_d$  represents a greater  
30 affinity. Immunological binding properties of selected polypeptides can be quantified

using methods well known in the art. One such method entails measuring the rates of antigen-binding site/antigen complex formation and dissociation, wherein those rates depend on the concentrations of the complex partners, the affinity of the interaction, and on geometric parameters that equally influence the rate in both directions. Thus, both the "on rate constant" ( $K_{on}$ ) and the "off rate constant" ( $K_{off}$ ) can be determined by calculation of the concentrations and the actual rates of association and dissociation. The ratio of  $K_{off}/K_{on}$  enables cancellation of all parameters not related to affinity, and is thus equal to the dissociation constant  $K_d$ . See, generally, Davies et al. (1990) Annual Rev. Biochem. 59:439-473.

10           An "antigen-binding site," or "binding portion" of an antibody refers to the part of the immunoglobulin molecule that participates in antigen binding. The antigen binding site is formed by amino acid residues of the N-terminal variable ("V") regions of the heavy ("H") and light ("L") chains. Three highly divergent stretches within the V regions of the heavy and light chains are referred to as "hypervariable regions" which are interposed between more conserved flanking stretches known as "framework regions," or "FRs". Thus the term "FR" refers to amino acid sequences which are naturally found between and adjacent to hypervariable regions in immunoglobulins. In an antibody molecule, the three hypervariable regions of a light chain and the three hypervariable regions of a heavy chain are disposed relative to each other in three dimensional space to form an antigen-binding surface. The antigen-binding surface is complementary to the three-dimensional surface of a bound antigen, and the three hypervariable regions of each of the heavy and light chains are referred to as "complementarity-determining regions," or "CDRs."

20           Binding agents may be further capable of differentiating between patients with and without a cancer, such as ovarian cancer, using the representative assays provided herein. For example, antibodies or other binding agents that bind to a tumor protein will preferably generate a signal indicating the presence of a cancer in at least about 20% of patients with the disease, more preferably at least about 30% of patients. Alternatively, or in addition, the antibody will generate a negative signal indicating the absence of the disease in at least about 90% of individuals without the cancer. To determine whether a binding agent satisfies this requirement, biological samples (e.g.,

blood, sera, sputum, urine and/or tumor biopsies) from patients with and without a cancer (as determined using standard clinical tests) may be assayed as described herein for the presence of polypeptides that bind to the binding agent. Preferably, a statistically significant number of samples with and without the disease will be assayed. Each  
5 binding agent should satisfy the above criteria; however, those of ordinary skill in the art will recognize that binding agents may be used in combination to improve sensitivity.

Any agent that satisfies the above requirements may be a binding agent. For example, a binding agent may be a ribosome, with or without a peptide component,  
10 an RNA molecule or a polypeptide. In a preferred embodiment, a binding agent is an antibody or an antigen-binding fragment thereof. Antibodies may be prepared by any of a variety of techniques known to those of ordinary skill in the art. See, e.g., Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In general, antibodies can be produced by cell culture techniques, including the generation  
15 of monoclonal antibodies as described herein, or via transfection of antibody genes into suitable bacterial or mammalian cell hosts, in order to allow for the production of recombinant antibodies. In one technique, an immunogen comprising the polypeptide is initially injected into any of a wide variety of mammals (e.g., mice, rats, rabbits, sheep or goats). In this step, the polypeptides of this invention may serve as the immunogen  
20 without modification. Alternatively, particularly for relatively short polypeptides, a superior immune response may be elicited if the polypeptide is joined to a carrier protein, such as bovine serum albumin or keyhole limpet hemocyanin. The immunogen is injected into the animal host, preferably according to a predetermined schedule incorporating one or more booster immunizations, and the animals are bled periodically.  
25 Polyclonal antibodies specific for the polypeptide may then be purified from such antisera by, for example, affinity chromatography using the polypeptide coupled to a suitable solid support.

Monoclonal antibodies specific for an antigenic polypeptide of interest may be prepared, for example, using the technique of Kohler and Milstein, *Eur. J.*  
30 *Immunol.* 6:511-519, 1976, and improvements thereto. Briefly, these methods involve the preparation of immortal cell lines capable of producing antibodies having the

desired specificity (*i.e.*, reactivity with the polypeptide of interest). Such cell lines may be produced, for example, from spleen cells obtained from an animal immunized as described above. The spleen cells are then immortalized by, for example, fusion with a myeloma cell fusion partner, preferably one that is syngeneic with the immunized animal. A variety of fusion techniques may be employed. For example, the spleen cells and myeloma cells may be combined with a nonionic detergent for a few minutes and then plated at low density on a selective medium that supports the growth of hybrid cells, but not myeloma cells. A preferred selection technique uses HAT (hypoxanthine, aminopterin, thymidine) selection. After a sufficient time, usually about 1 to 2 weeks, colonies of hybrids are observed. Single colonies are selected and their culture supernatants tested for binding activity against the polypeptide. Hybridomas having high reactivity and specificity are preferred.

Monoclonal antibodies may be isolated from the supernatants of growing hybridoma colonies. In addition, various techniques may be employed to enhance the yield, such as injection of the hybridoma cell line into the peritoneal cavity of a suitable vertebrate host, such as a mouse. Monoclonal antibodies may then be harvested from the ascites fluid or the blood. Contaminants may be removed from the antibodies by conventional techniques, such as chromatography, gel filtration, precipitation, and extraction. The polypeptides of this invention may be used in the purification process in, for example, an affinity chromatography step.

A number of therapeutically useful molecules are known in the art which comprise antigen-binding sites that are capable of exhibiting immunological binding properties of an antibody molecule. The proteolytic enzyme papain preferentially cleaves IgG molecules to yield several fragments, two of which (the "F(ab)" fragments) each comprise a covalent heterodimer that includes an intact antigen-binding site. The enzyme pepsin is able to cleave IgG molecules to provide several fragments, including the "F(ab)<sub>2</sub>" fragment which comprises both antigen-binding sites. An "Fv" fragment can be produced by preferential proteolytic cleavage of an IgM, and on rare occasions IgG or IgA immunoglobulin molecule. Fv fragments are, however, more commonly derived using recombinant techniques known in the art. The Fv fragment includes a non-covalent V<sub>H</sub>::V<sub>L</sub> heterodimer including an antigen-binding site which retains much

of the antigen recognition and binding capabilities of the native antibody molecule. Inbar et al. (1972) Proc. Nat. Acad. Sci. USA 69:2659-2662; Hochman et al. (1976) Biochem 15:2706-2710; and Ehrlich et al. (1980) Biochem 19:4091-4096.

A single chain Fv ("sFv") polypeptide is a covalently linked  $V_H::V_L$  heterodimer which is expressed from a gene fusion including  $V_H$ - and  $V_L$ -encoding genes linked by a peptide-encoding linker. Huston et al. (1988) Proc. Nat. Acad. Sci. USA 85(16):5879-5883. A number of methods have been described to discern chemical structures for converting the naturally aggregated--but chemically separated--light and heavy polypeptide chains from an antibody V region into an sFv molecule which will fold into a three dimensional structure substantially similar to the structure of an antigen-binding site. See, *e.g.*, U.S. Pat. Nos. 5,091,513 and 5,132,405, to Huston et al.; and U.S. Pat. No. 4,946,778, to Ladner et al.

Each of the above-described molecules includes a heavy chain and a light chain CDR set, respectively interposed between a heavy chain and a light chain FR set which provide support to the CDRs and define the spatial relationship of the CDRs relative to each other. As used herein, the term "CDR set" refers to the three hypervariable regions of a heavy or light chain V region. Proceeding from the N-terminus of a heavy or light chain, these regions are denoted as "CDR1," "CDR2," and "CDR3" respectively. An antigen-binding site, therefore, includes six CDRs, comprising the CDR set from each of a heavy and a light chain V region. A polypeptide comprising a single CDR, (*e.g.*, a CDR1, CDR2 or CDR3) is referred to herein as a "molecular recognition unit." Crystallographic analysis of a number of antigen-antibody complexes has demonstrated that the amino acid residues of CDRs form extensive contact with bound antigen, wherein the most extensive antigen contact is with the heavy chain CDR3. Thus, the molecular recognition units are primarily responsible for the specificity of an antigen-binding site.

As used herein, the term "FR set" refers to the four flanking amino acid sequences which frame the CDRs of a CDR set of a heavy or light chain V region. Some FR residues may contact bound antigen; however, FRs are primarily responsible for folding the V region into the antigen-binding site, particularly the FR residues directly adjacent to the CDRs. Within FRs, certain amino residues and certain structural

features are very highly conserved. In this regard, all V region sequences contain an internal disulfide loop of around 90 amino acid residues. When the V regions fold into a binding-site, the CDRs are displayed as projecting loop motifs which form an antigen-binding surface. It is generally recognized that there are conserved structural regions of FRs which influence the folded shape of the CDR loops into certain "canonical" structures--regardless of the precise CDR amino acid sequence. Further, certain FR residues are known to participate in non-covalent interdomain contacts which stabilize the interaction of the antibody heavy and light chains.

A number of "humanized" antibody molecules comprising an antigen-binding site derived from a non-human immunoglobulin have been described, including chimeric antibodies having rodent V regions and their associated CDRs fused to human constant domains (Winter et al. (1991) *Nature* 349:293-299; Lobuglio et al. (1989) *Proc. Nat. Acad. Sci. USA* 86:4220-4224; Shaw et al. (1987) *J Immunol.* 138:4534-4538; and Brown et al. (1987) *Cancer Res.* 47:3577-3583), rodent CDRs grafted into a human supporting FR prior to fusion with an appropriate human antibody constant domain (Riechmann et al. (1988) *Nature* 332:323-327; Verhoeyen et al. (1988) *Science* 239:1534-1536; and Jones et al. (1986) *Nature* 321:522-525), and rodent CDRs supported by recombinantly veneered rodent FRs (European Patent Publication No. 519,596, published Dec. 23, 1992). These "humanized" molecules are designed to minimize unwanted immunological response toward rodent antihuman antibody molecules which limits the duration and effectiveness of therapeutic applications of those moieties in human recipients.

As used herein, the terms "veneered FRs" and "recombinantly veneered FRs" refer to the selective replacement of FR residues from, *e.g.*, a rodent heavy or light chain V region, with human FR residues in order to provide a xenogeneic molecule comprising an antigen-binding site which retains substantially all of the native FR polypeptide folding structure. Veneering techniques are based on the understanding that the ligand binding characteristics of an antigen-binding site are determined primarily by the structure and relative disposition of the heavy and light chain CDR sets within the antigen-binding surface. Davies et al. (1990) *Ann. Rev. Biochem.* 59:439-473. Thus, antigen binding specificity can be preserved in a humanized antibody only wherein the

CDR structures, their interaction with each other, and their interaction with the rest of the V region domains are carefully maintained. By using veneering techniques, exterior (*e.g.*, solvent-accessible) FR residues which are readily encountered by the immune system are selectively replaced with human residues to provide a hybrid molecule that  
5 comprises either a weakly immunogenic, or substantially non-immunogenic veneered surface.

The process of veneering makes use of the available sequence data for human antibody variable domains compiled by Kabat et al., in Sequences of Proteins of Immunological Interest, 4th ed., (U.S. Dept. of Health and Human Services, U.S.  
10 Government Printing Office, 1987), updates to the Kabat database, and other accessible U.S. and foreign databases (both nucleic acid and protein). Solvent accessibilities of V region amino acids can be deduced from the known three-dimensional structure for human and murine antibody fragments. There are two general steps in veneering a murine antigen-binding site. Initially, the FRs of the variable domains of an antibody  
15 molecule of interest are compared with corresponding FR sequences of human variable domains obtained from the above-identified sources. The most homologous human V regions are then compared residue by residue to corresponding murine amino acids. The residues in the murine FR which differ from the human counterpart are replaced by the residues present in the human moiety using recombinant techniques well known in the  
20 art. Residue switching is only carried out with moieties which are at least partially exposed (solvent accessible), and care is exercised in the replacement of amino acid residues which may have a significant effect on the tertiary structure of V region domains, such as proline, glycine and charged amino acids.

In this manner, the resultant "veneered" murine antigen-binding sites are  
25 thus designed to retain the murine CDR residues, the residues substantially adjacent to the CDRs, the residues identified as buried or mostly buried (solvent inaccessible), the residues believed to participate in non-covalent (*e.g.*, electrostatic and hydrophobic) contacts between heavy and light chain domains, and the residues from conserved structural regions of the FRs which are believed to influence the "canonical" tertiary  
30 structures of the CDR loops. These design criteria are then used to prepare recombinant nucleotide sequences which combine the CDRs of both the heavy and light chain of a

murine antigen-binding site into human-appearing FRs that can be used to transfect mammalian cells for the expression of recombinant human antibodies which exhibit the antigen specificity of the murine antibody molecule.

In another embodiment of the invention, monoclonal antibodies of the present invention may be coupled to one or more therapeutic agents. Suitable agents in this regard include radionuclides, differentiation inducers, drugs, toxins, and derivatives thereof. Preferred radionuclides include  $^{90}\text{Y}$ ,  $^{123}\text{I}$ ,  $^{125}\text{I}$ ,  $^{131}\text{I}$ ,  $^{186}\text{Re}$ ,  $^{188}\text{Re}$ ,  $^{211}\text{At}$ , and  $^{212}\text{Bi}$ . Preferred drugs include methotrexate, and pyrimidine and purine analogs. Preferred differentiation inducers include phorbol esters and butyric acid. Preferred toxins include ricin, abrin, diphtheria toxin, cholera toxin, gelonin, *Pseudomonas* exotoxin, *Shigella* toxin, and pokeweed antiviral protein.

A therapeutic agent may be coupled (*e.g.*, covalently bonded) to a suitable monoclonal antibody either directly or indirectly (*e.g.*, via a linker group). A direct reaction between an agent and an antibody is possible when each possesses a substituent capable of reacting with the other. For example, a nucleophilic group, such as an amino or sulfhydryl group, on one may be capable of reacting with a carbonyl-containing group, such as an anhydride or an acid halide, or with an alkyl group containing a good leaving group (*e.g.*, a halide) on the other.

Alternatively, it may be desirable to couple a therapeutic agent and an antibody via a linker group. A linker group can function as a spacer to distance an antibody from an agent in order to avoid interference with binding capabilities. A linker group can also serve to increase the chemical reactivity of a substituent on an agent or an antibody, and thus increase the coupling efficiency. An increase in chemical reactivity may also facilitate the use of agents, or functional groups on agents, which otherwise would not be possible.

It will be evident to those skilled in the art that a variety of bifunctional or polyfunctional reagents, both homo- and hetero-functional (such as those described in the catalog of the Pierce Chemical Co., Rockford, IL), may be employed as the linker group. Coupling may be effected, for example, through amino groups, carboxyl groups, sulfhydryl groups or oxidized carbohydrate residues. There are numerous references describing such methodology, *e.g.*, U.S. Patent No. 4,671,958, to Rodwell et al.

Where a therapeutic agent is more potent when free from the antibody portion of the immunoconjugates of the present invention, it may be desirable to use a linker group which is cleavable during or upon internalization into a cell. A number of different cleavable linker groups have been described. The mechanisms for the intracellular release of an agent from these linker groups include cleavage by reduction of a disulfide bond (*e.g.*, U.S. Patent No. 4,489,710, to Spitler), by irradiation of a photolabile bond (*e.g.*, U.S. Patent No. 4,625,014, to Senter et al.), by hydrolysis of derivatized amino acid side chains (*e.g.*, U.S. Patent No. 4,638,045, to Kohn et al.), by serum complement-mediated hydrolysis (*e.g.*, U.S. Patent No. 4,671,958, to Rodwell et al.), and acid-catalyzed hydrolysis (*e.g.*, U.S. Patent No. 4,569,789, to Blattler et al.).

It may be desirable to couple more than one agent to an antibody. In one embodiment, multiple molecules of an agent are coupled to one antibody molecule. In another embodiment, more than one type of agent may be coupled to one antibody. Regardless of the particular embodiment, immunoconjugates with more than one agent may be prepared in a variety of ways. For example, more than one agent may be coupled directly to an antibody molecule, or linkers that provide multiple sites for attachment can be used. Alternatively, a carrier can be used.

A carrier may bear the agents in a variety of ways, including covalent bonding either directly or via a linker group. Suitable carriers include proteins such as albumins (*e.g.*, U.S. Patent No. 4,507,234, to Kato et al.), peptides and polysaccharides such as aminodextran (*e.g.*, U.S. Patent No. 4,699,784, to Shih et al.). A carrier may also bear an agent by noncovalent bonding or by encapsulation, such as within a liposome vesicle (*e.g.*, U.S. Patent Nos. 4,429,008 and 4,873,088). Carriers specific for radionuclide agents include radiohalogenated small molecules and chelating compounds. For example, U.S. Patent No. 4,735,792 discloses representative radiohalogenated small molecules and their synthesis. A radionuclide chelate may be formed from chelating compounds that include those containing nitrogen and sulfur atoms as the donor atoms for binding the metal, or metal oxide, radionuclide. For example, U.S. Patent No. 4,673,562, to Davison et al. discloses representative chelating compounds and their synthesis.

### T Cell Compositions

The present invention, in another aspect, provides T cells specific for a tumor polypeptide disclosed herein, or for a variant or derivative thereof. Such cells may generally be prepared *in vitro* or *ex vivo*, using standard procedures. For example,  
5 T cells may be isolated from bone marrow, peripheral blood, or a fraction of bone marrow or peripheral blood of a patient, using a commercially available cell separation system, such as the Isolex™ System, available from Nexell Therapeutics, Inc. (Irvine, CA; see also U.S. Patent No. 5,240,856; U.S. Patent No. 5,215,926; WO 89/06280; WO 91/16116 and WO 92/07243). Alternatively, T cells may be derived from related or  
10 unrelated humans, non-human mammals, cell lines or cultures.

T cells may be stimulated with a polypeptide, polynucleotide encoding a polypeptide and/or an antigen presenting cell (APC) that expresses such a polypeptide. Such stimulation is performed under conditions and for a time sufficient to permit the generation of T cells that are specific for the polypeptide of interest. Preferably, a tumor  
15 polypeptide or polynucleotide of the invention is present within a delivery vehicle, such as a microsphere, to facilitate the generation of specific T cells.

T cells are considered to be specific for a polypeptide of the present invention if the T cells specifically proliferate, secrete cytokines or kill target cells coated with the polypeptide or expressing a gene encoding the polypeptide. T cell  
20 specificity may be evaluated using any of a variety of standard techniques. For example, within a chromium release assay or proliferation assay, a stimulation index of more than two fold increase in lysis and/or proliferation, compared to negative controls, indicates T cell specificity. Such assays may be performed, for example, as described in Chen et al., *Cancer Res.* 54:1065-1070, 1994. Alternatively, detection of the  
25 proliferation of T cells may be accomplished by a variety of known techniques. For example, T cell proliferation can be detected by measuring an increased rate of DNA synthesis (e.g., by pulse-labeling cultures of T cells with tritiated thymidine and measuring the amount of tritiated thymidine incorporated into DNA). Contact with a  
tumor polypeptide (100 ng/ml - 100 µg/ml, preferably 200 ng/ml - 25 µg/ml) for 3 - 7  
30 days will typically result in at least a two fold increase in proliferation of the T cells. Contact as described above for 2-3 hours should result in activation of the T cells, as

measured using standard cytokine assays in which a two fold increase in the level of cytokine release (e.g., TNF or IFN- $\gamma$ ) is indicative of T cell activation (*see* Coligan et al., Current Protocols in Immunology, vol. 1, Wiley Interscience (Greene 1998)). T cells that have been activated in response to a tumor polypeptide, polynucleotide or polypeptide-expressing APC may be CD4<sup>+</sup> and/or CD8<sup>+</sup>. Tumor polypeptide-specific T cells may be expanded using standard techniques. Within preferred embodiments, the T cells are derived from a patient, a related donor or an unrelated donor, and are administered to the patient following stimulation and expansion.

For therapeutic purposes, CD4<sup>+</sup> or CD8<sup>+</sup> T cells that proliferate in response to a tumor polypeptide, polynucleotide or APC can be expanded in number either *in vitro* or *in vivo*. Proliferation of such T cells *in vitro* may be accomplished in a variety of ways. For example, the T cells can be re-exposed to a tumor polypeptide, or a short peptide corresponding to an immunogenic portion of such a polypeptide, with or without the addition of T cell growth factors, such as interleukin-2, and/or stimulator cells that synthesize a tumor polypeptide. Alternatively, one or more T cells that proliferate in the presence of the tumor polypeptide can be expanded in number by cloning. Methods for cloning cells are well known in the art, and include limiting dilution.

#### Pharmaceutical Compositions

In additional embodiments, the present invention concerns formulation of one or more of the polynucleotide, polypeptide, T-cell and/or antibody compositions disclosed herein in pharmaceutically-acceptable carriers for administration to a cell or an animal, either alone, or in combination with one or more other modalities of therapy.

It will be understood that, if desired, a composition as disclosed herein may be administered in combination with other agents as well, such as, e.g., other proteins or polypeptides or various pharmaceutically-active agents. In fact, there is virtually no limit to other components that may also be included, given that the additional agents do not cause a significant adverse effect upon contact with the target cells or host tissues. The compositions may thus be delivered along with various other agents as required in the particular instance. Such compositions may be purified from

host cells or other biological sources, or alternatively may be chemically synthesized as described herein. Likewise, such compositions may further comprise substituted or derivatized RNA or DNA compositions.

Therefore, in another aspect of the present invention, pharmaceutical  
5 compositions are provided comprising one or more of the polynucleotide, polypeptide, antibody, and/or T-cell compositions described herein in combination with a physiologically acceptable carrier. In certain preferred embodiments, the pharmaceutical compositions of the invention comprise immunogenic polynucleotide and/or polypeptide compositions of the invention for use in prophylactic and therapeutic  
10 vaccine applications. Vaccine preparation is generally described in, for example, M.F. Powell and M.J. Newman, eds., "Vaccine Design (the subunit and adjuvant approach)," Plenum Press (NY, 1995). Generally, such compositions will comprise one or more polynucleotide and/or polypeptide compositions of the present invention in combination with one or more immunostimulants.

15 It will be apparent that any of the pharmaceutical compositions described herein can contain pharmaceutically acceptable salts of the polynucleotides and polypeptides of the invention. Such salts can be prepared, for example, from pharmaceutically acceptable non-toxic bases, including organic bases (*e.g.*, salts of primary, secondary and tertiary amines and basic amino acids) and inorganic bases (*e.g.*,  
20 sodium, potassium, lithium, ammonium, calcium and magnesium salts).

In another embodiment, illustrative immunogenic compositions, *e.g.*, vaccine compositions, of the present invention comprise DNA encoding one or more of the polypeptides as described above, such that the polypeptide is generated *in situ*. As noted above, the polynucleotide may be administered within any of a variety of delivery  
25 systems known to those of ordinary skill in the art. Indeed, numerous gene delivery techniques are well known in the art, such as those described by Rolland, *Crit. Rev. Therap. Drug Carrier Systems* 15:143-198, 1998, and references cited therein. Appropriate polynucleotide expression systems will, of course, contain the necessary regulatory DNA regulatory sequences for expression in a patient (such as a suitable  
30 promoter and terminating signal). Alternatively, bacterial delivery systems may involve

the administration of a bacterium (such as *Bacillus-Calmette-Guerrin*) that expresses an immunogenic portion of the polypeptide on its cell surface or secretes such an epitope.

Therefore, in certain embodiments, polynucleotides encoding immunogenic polypeptides described herein are introduced into suitable mammalian host cells for expression using any of a number of known viral-based systems. In one illustrative embodiment, retroviruses provide a convenient and effective platform for gene delivery systems. A selected nucleotide sequence encoding a polypeptide of the present invention can be inserted into a vector and packaged in retroviral particles using techniques known in the art. The recombinant virus can then be isolated and delivered to a subject. A number of illustrative retroviral systems have been described (*e.g.*, U.S. Pat. No. 5,219,740; Miller and Rosman (1989) *BioTechniques* 7:980-990; Miller, A. D. (1990) *Human Gene Therapy* 1:5-14; Scarpa et al. (1991) *Virology* 180:849-852; Burns et al. (1993) *Proc. Natl. Acad. Sci. USA* 90:8033-8037; and Boris-Lawrie and Temin (1993) *Cur. Opin. Genet. Develop.* 3:102-109.

In addition, a number of illustrative adenovirus-based systems have also been described. Unlike retroviruses which integrate into the host genome, adenoviruses persist extrachromosomally thus minimizing the risks associated with insertional mutagenesis (Haj-Ahmad and Graham (1986) *J. Virol.* 57:267-274; Bett et al. (1993) *J. Virol.* 67:5911-5921; Mittereder et al. (1994) *Human Gene Therapy* 5:717-729; Seth et al. (1994) *J. Virol.* 68:933-940; Barr et al. (1994) *Gene Therapy* 1:51-58; Berkner, K. L. (1988) *BioTechniques* 6:616-629; and Rich et al. (1993) *Human Gene Therapy* 4:461-476).

Various adeno-associated virus (AAV) vector systems have also been developed for polynucleotide delivery. AAV vectors can be readily constructed using techniques well known in the art. See, *e.g.*, U.S. Pat. Nos. 5,173,414 and 5,139,941; International Publication Nos. WO 92/01070 and WO 93/03769; Lebkowski et al. (1988) *Molec. Cell. Biol.* 8:3988-3996; Vincent et al. (1990) *Vaccines 90* (Cold Spring Harbor Laboratory Press); Carter, B. J. (1992) *Current Opinion in Biotechnology* 3:533-539; Muzyczka, N. (1992) *Current Topics in Microbiol. and Immunol.* 158:97-129; Kotin, R. M. (1994) *Human Gene Therapy* 5:793-801; Shelling and Smith (1994) *Gene Therapy* 1:165-169; and Zhou et al. (1994) *J. Exp. Med.* 179:1867-1875.

Additional viral vectors useful for delivering the polynucleotides encoding polypeptides of the present invention by gene transfer include those derived from the pox family of viruses, such as vaccinia virus and avian poxvirus. By way of example, vaccinia virus recombinants expressing the novel molecules can be  
5 constructed as follows. The DNA encoding a polypeptide is first inserted into an appropriate vector so that it is adjacent to a vaccinia promoter and flanking vaccinia DNA sequences, such as the sequence encoding thymidine kinase (TK). This vector is then used to transfect cells which are simultaneously infected with vaccinia. Homologous recombination serves to insert the vaccinia promoter plus the gene  
10 encoding the polypeptide of interest into the viral genome. The resulting TK.sup.(-) recombinant can be selected by culturing the cells in the presence of 5-bromodeoxyuridine and picking viral plaques resistant thereto.

A vaccinia-based infection/transfection system can be conveniently used to provide for inducible, transient expression or coexpression of one or more  
15 polypeptides described herein in host cells of an organism. In this particular system, cells are first infected in vitro with a vaccinia virus recombinant that encodes the bacteriophage T7 RNA polymerase. This polymerase displays exquisite specificity in that it only transcribes templates bearing T7 promoters. Following infection, cells are transfected with the polynucleotide or polynucleotides of interest, driven by a T7  
20 promoter. The polymerase expressed in the cytoplasm from the vaccinia virus recombinant transcribes the transfected DNA into RNA which is then translated into polypeptide by the host translational machinery. The method provides for high level, transient, cytoplasmic production of large quantities of RNA and its translation products. See, *e.g.*, Elroy-Stein and Moss, Proc. Natl. Acad. Sci. USA (1990) 87:6743-  
25 6747; Fuerst et al. Proc. Natl. Acad. Sci. USA (1986) 83:8122-8126.

Alternatively, avipoxviruses, such as the fowlpox and canarypox viruses, can also be used to deliver the coding sequences of interest. Recombinant avipox viruses, expressing immunogens from mammalian pathogens, are known to confer protective immunity when administered to non-avian species. The use of an Avipox  
30 vector is particularly desirable in human and other mammalian species since members of the Avipox genus can only productively replicate in susceptible avian species and

therefore are not infective in mammalian cells. Methods for producing recombinant Avipoxviruses are known in the art and employ genetic recombination, as described above with respect to the production of vaccinia viruses. See, e.g., WO 91/12882; WO 89/03429; and WO 92/03545.

5           Any of a number of alphavirus vectors can also be used for delivery of polynucleotide compositions of the present invention, such as those vectors described in U.S. Patent Nos. 5,843,723; 6,015,686; 6,008,035 and 6,015,694. Certain vectors based on Venezuelan Equine Encephalitis (VEE) can also be used, illustrative examples of which can be found in U.S. Patent Nos. 5,505,947 and 5,643,576.

10           Moreover, molecular conjugate vectors, such as the adenovirus chimeric vectors described in Michael et al. J. Biol. Chem. (1993) 268:6866-6869 and Wagner et al. Proc. Natl. Acad. Sci. USA (1992) 89:6099-6103, can also be used for gene delivery under the invention.

          Additional illustrative information on these and other known viral-based  
15 delivery systems can be found, for example, in Fisher-Hoch et al., *Proc. Natl. Acad. Sci. USA* 86:317-321, 1989; Flexner et al., *Ann. N.Y. Acad. Sci.* 569:86-103, 1989; Flexner et al., *Vaccine* 8:17-21, 1990; U.S. Patent Nos. 4,603,112, 4,769,330, and 5,017,487; WO 89/01973; U.S. Patent No. 4,777,127; GB 2,200,651; EP 0,345,242; WO 91/02805; Berkner, *Biotechniques* 6:616-627, 1988; Rosenfeld et al., *Science*  
20 252:431-434, 1991; Kolls et al., *Proc. Natl. Acad. Sci. USA* 91:215-219, 1994; Kass-Eisler et al., *Proc. Natl. Acad. Sci. USA* 90:11498-11502, 1993; Guzman et al., *Circulation* 88:2838-2848, 1993; and Guzman et al., *Cir. Res.* 73:1202-1207, 1993.

          In certain embodiments, a polynucleotide may be integrated into the genome of a target cell. This integration may be in the specific location and orientation  
25 via homologous recombination (gene replacement) or it may be integrated in a random, non-specific location (gene augmentation). In yet further embodiments, the polynucleotide may be stably maintained in the cell as a separate, episomal segment of DNA. Such polynucleotide segments or "episomes" encode sequences sufficient to permit maintenance and replication independent of or in synchronization with the host  
30 cell cycle. The manner in which the expression construct is delivered to a cell and

where in the cell the polynucleotide remains is dependent on the type of expression construct employed.

In another embodiment of the invention, a polynucleotide is administered/delivered as "naked" DNA, for example as described in Ulmer et al.,  
5 *Science* 259:1745-1749, 1993 and reviewed by Cohen, *Science* 259:1691-1692, 1993. The uptake of naked DNA may be increased by coating the DNA onto biodegradable beads, which are efficiently transported into the cells.

In still another embodiment, a composition of the present invention can be delivered via a particle bombardment approach, many of which have been described.  
10 In one illustrative example, gas-driven particle acceleration can be achieved with devices such as those manufactured by Powderject Pharmaceuticals PLC (Oxford, UK) and Powderject Vaccines Inc. (Madison, WI), some examples of which are described in U.S. Patent Nos. 5,846,796; 6,010,478; 5,865,796; 5,584,807; and EP Patent No. 0500 799. This approach offers a needle-free delivery approach wherein a dry powder  
15 formulation of microscopic particles, such as polynucleotide or polypeptide particles, are accelerated to high speed within a helium gas jet generated by a hand held device, propelling the particles into a target tissue of interest.

In a related embodiment, other devices and methods that may be useful for gas-driven needle-less injection of compositions of the present invention include  
20 those provided by Bioject, Inc. (Portland, OR), some examples of which are described in U.S. Patent Nos. 4,790,824; 5,064,413; 5,312,335; 5,383,851; 5,399,163; 5,520,639 and 5,993,412.

According to another embodiment, the pharmaceutical compositions described herein will comprise one or more immunostimulants in addition to the  
25 immunogenic polynucleotide, polypeptide, antibody, T-cell and/or APC compositions of this invention. An immunostimulant refers to essentially any substance that enhances or potentiates an immune response (antibody and/or cell-mediated) to an exogenous antigen. One preferred type of immunostimulant comprises an adjuvant. Many adjuvants contain a substance designed to protect the antigen from rapid catabolism,  
30 such as aluminum hydroxide or mineral oil, and a stimulator of immune responses, such as lipid A, *Bordetella pertussis* or *Mycobacterium tuberculosis* derived proteins.

Certain adjuvants are commercially available as, for example, Freund's Incomplete Adjuvant and Complete Adjuvant (Difco Laboratories, Detroit, MI); Merck Adjuvant 65 (Merck and Company, Inc., Rahway, NJ); AS-2 (SmithKline Beecham, Philadelphia, PA); aluminum salts such as aluminum hydroxide gel (alum) or aluminum phosphate; salts of calcium, iron or zinc; an insoluble suspension of acylated tyrosine; acylated sugars; cationically or anionically derivatized polysaccharides; polyphosphazenes; biodegradable microspheres; monophosphoryl lipid A and quil A. Cytokines, such as GM-CSF, interleukin-2, -7, -12, and other like growth factors, may also be used as adjuvants.

10                Within certain embodiments of the invention, the adjuvant composition is preferably one that induces an immune response predominantly of the Th1 type. High levels of Th1-type cytokines (*e.g.*, IFN- $\gamma$ , TNF $\alpha$ , IL-2 and IL-12) tend to favor the induction of cell mediated immune responses to an administered antigen. In contrast, high levels of Th2-type cytokines (*e.g.*, IL-4, IL-5, IL-6 and IL-10) tend to favor the  
15 induction of humoral immune responses. Following application of a vaccine as provided herein, a patient will support an immune response that includes Th1- and Th2-type responses. Within a preferred embodiment, in which a response is predominantly Th1-type, the level of Th1-type cytokines will increase to a greater extent than the level of Th2-type cytokines. The levels of these cytokines may be readily assessed using  
20 standard assays. For a review of the families of cytokines, see Mosmann and Coffman, *Ann. Rev. Immunol.* 7:145-173, 1989.

Certain preferred adjuvants for eliciting a predominantly Th1-type response include, for example, a combination of monophosphoryl lipid A, preferably 3-de-O-acylated monophosphoryl lipid A, together with an aluminum salt. MPL<sup>®</sup>  
25 adjuvants are available from Corixa Corporation (Seattle, WA; *see*, for example, US Patent Nos. 4,436,727; 4,877,611; 4,866,034 and 4,912,094). CpG-containing oligonucleotides (in which the CpG dinucleotide is unmethylated) also induce a predominantly Th1 response. Such oligonucleotides are well known and are described, for example, in WO 96/02555, WO 99/33488 and U.S. Patent Nos. 6,008,200 and  
30 5,856,462. Immunostimulatory DNA sequences are also described, for example, by Sato et al., *Science* 273:352, 1996. Another preferred adjuvant comprises a saponin,

such as Quil A, or derivatives thereof, including QS21 and QS7 (Aquila Biopharmaceuticals Inc., Framingham, MA); Escin; Digitonin; or *Gypsophila* or *Chenopodium quinoa* saponins. Other preferred formulations include more than one saponin in the adjuvant combinations of the present invention, for example  
5 combinations of at least two of the following group comprising QS21, QS7, Quil A,  $\beta$ -escin, or digitonin.

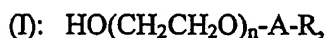
Alternatively the saponin formulations may be combined with vaccine vehicles composed of chitosan or other polycationic polymers, polylactide and polylactide-co-glycolide particles, poly-N-acetyl glucosamine-based polymer matrix,  
10 particles composed of polysaccharides or chemically modified polysaccharides, liposomes and lipid-based particles, particles composed of glycerol monoesters, etc. The saponins may also be formulated in the presence of cholesterol to form particulate structures such as liposomes or ISCOMs. Furthermore, the saponins may be formulated together with a polyoxyethylene ether or ester, in either a non-particulate solution or  
15 suspension, or in a particulate structure such as a paucilamellar liposome or ISCOM. The saponins may also be formulated with excipients such as Carbopol<sup>R</sup> to increase viscosity, or may be formulated in a dry powder form with a powder excipient such as lactose.

In one preferred embodiment, the adjuvant system includes the  
20 combination of a monophosphoryl lipid A and a saponin derivative, such as the combination of QS21 and 3D-MPL<sup>®</sup> adjuvant, as described in WO 94/00153, or a less reactogenic composition where the QS21 is quenched with cholesterol, as described in WO 96/33739. Other preferred formulations comprise an oil-in-water emulsion and tocopherol. Another particularly preferred adjuvant formulation employing QS21, 3D-  
25 MPL<sup>®</sup> adjuvant and tocopherol in an oil-in-water emulsion is described in WO 95/17210.

Another enhanced adjuvant system involves the combination of a CpG-containing oligonucleotide and a saponin derivative particularly the combination of CpG and QS21 is disclosed in WO 00/09159. Preferably the formulation additionally  
30 comprises an oil in water emulsion and tocopherol.

Additional illustrative adjuvants for use in the pharmaceutical compositions of the invention include Montanide ISA 720 (Seppic, France), SAF (Chiron, California, United States), ISCOMS (CSL), MF-59 (Chiron), the SBAS series of adjuvants (e.g., SBAS-2 or SBAS-4, available from SmithKline Beecham, Rixensart, Belgium), Detox (Enhanzyn<sup>®</sup>) (Corixa, Hamilton, MT), RC-529 (Corixa, Hamilton, MT) and other aminoalkyl glucosaminide 4-phosphates (AGPs), such as those described in pending U.S. Patent Application Serial Nos. 08/853,826 and 09/074,720, the disclosures of which are incorporated herein by reference in their entireties, and polyoxyethylene ether adjuvants such as those described in WO 99/52549A1.

Other preferred adjuvants include adjuvant molecules of the general formula



wherein,  $n$  is 1-50,  $A$  is a bond or  $-\text{C}(\text{O})-$ ,  $R$  is  $\text{C}_{1-50}$  alkyl or Phenyl  $\text{C}_{1-50}$  alkyl.

One embodiment of the present invention consists of a vaccine formulation comprising a polyoxyethylene ether of general formula (I), wherein  $n$  is between 1 and 50, preferably 4-24, most preferably 9; the  $R$  component is  $\text{C}_{1-50}$ , preferably  $\text{C}_4\text{-C}_{20}$  alkyl and most preferably  $\text{C}_{12}$  alkyl, and  $A$  is a bond. The concentration of the polyoxyethylene ethers should be in the range 0.1-20%, preferably from 0.1-10%, and most preferably in the range 0.1-1%. Preferred polyoxyethylene ethers are selected from the following group: polyoxyethylene-9-lauryl ether, polyoxyethylene-9-stearyl ether, polyoxyethylene-8-stearyl ether, polyoxyethylene-4-lauryl ether, polyoxyethylene-35-lauryl ether, and polyoxyethylene-23-lauryl ether. Polyoxyethylene ethers such as polyoxyethylene lauryl ether are described in the Merck index (12<sup>th</sup> edition: entry 7717). These adjuvant molecules are described in WO 99/52549.

The polyoxyethylene ether according to the general formula (I) above may, if desired, be combined with another adjuvant. For example, a preferred adjuvant combination is preferably with CpG as described in the pending UK patent application GB 9820956.2.

According to another embodiment of this invention, an immunogenic composition described herein is delivered to a host via antigen presenting cells (APCs),

such as dendritic cells, macrophages, B cells, monocytes and other cells that may be engineered to be efficient APCs. Such cells may, but need not, be genetically modified to increase the capacity for presenting the antigen, to improve activation and/or maintenance of the T cell response, to have anti-tumor effects *per se* and/or to be  
5 immunologically compatible with the receiver (*i.e.*, matched HLA haplotype). APCs may generally be isolated from any of a variety of biological fluids and organs, including tumor and peritumoral tissues, and may be autologous, allogeneic, syngeneic or xenogeneic cells.

Certain preferred embodiments of the present invention use dendritic  
10 cells or progenitors thereof as antigen-presenting cells. Dendritic cells are highly potent APCs (Banchereau and Steinman, *Nature* 392:245-251, 1998) and have been shown to be effective as a physiological adjuvant for eliciting prophylactic or therapeutic antitumor immunity (*see* Timmerman and Levy, *Ann. Rev. Med.* 50:507-529, 1999). In general, dendritic cells may be identified based on their typical shape (stellate *in situ*,  
15 with marked cytoplasmic processes (dendrites) visible *in vitro*), their ability to take up, process and present antigens with high efficiency and their ability to activate naïve T cell responses. Dendritic cells may, of course, be engineered to express specific cell-surface receptors or ligands that are not commonly found on dendritic cells *in vivo* or *ex vivo*, and such modified dendritic cells are contemplated by the present invention. As  
20 an alternative to dendritic cells, secreted vesicles antigen-loaded dendritic cells (called exosomes) may be used within a vaccine (*see* Zitvogel et al., *Nature Med.* 4:594-600, 1998).

Dendritic cells and progenitors may be obtained from peripheral blood, bone marrow, tumor-infiltrating cells, peritumoral tissues-infiltrating cells, lymph  
25 nodes, spleen, skin, umbilical cord blood or any other suitable tissue or fluid. For example, dendritic cells may be differentiated *ex vivo* by adding a combination of cytokines such as GM-CSF, IL-4, IL-13 and/or TNF $\alpha$  to cultures of monocytes harvested from peripheral blood. Alternatively, CD34 positive cells harvested from peripheral blood, umbilical cord blood or bone marrow may be differentiated into  
30 dendritic cells by adding to the culture medium combinations of GM-CSF, IL-3, TNF $\alpha$ ,

CD40 ligand, LPS, flt3 ligand and/or other compound(s) that induce differentiation, maturation and proliferation of dendritic cells.

Dendritic cells are conveniently categorized as "immature" and "mature" cells, which allows a simple way to discriminate between two well characterized phenotypes. However, this nomenclature should not be construed to exclude all possible intermediate stages of differentiation. Immature dendritic cells are characterized as APC with a high capacity for antigen uptake and processing, which correlates with the high expression of Fcγ receptor and mannose receptor. The mature phenotype is typically characterized by a lower expression of these markers, but a high expression of cell surface molecules responsible for T cell activation such as class I and class II MHC, adhesion molecules (*e.g.*, CD54 and CD11) and costimulatory molecules (*e.g.*, CD40, CD80, CD86 and 4-1BB).

APCs may generally be transfected with a polynucleotide of the invention (or portion or other variant thereof) such that the encoded polypeptide, or an immunogenic portion thereof, is expressed on the cell surface. Such transfection may take place *ex vivo*, and a pharmaceutical composition comprising such transfected cells may then be used for therapeutic purposes, as described herein. Alternatively, a gene delivery vehicle that targets a dendritic or other antigen presenting cell may be administered to a patient, resulting in transfection that occurs *in vivo*. *In vivo* and *ex vivo* transfection of dendritic cells, for example, may generally be performed using any methods known in the art, such as those described in WO 97/24447, or the gene gun approach described by Mahvi et al., *Immunology and cell Biology* 75:456-460, 1997. Antigen loading of dendritic cells may be achieved by incubating dendritic cells or progenitor cells with the tumor polypeptide, DNA (naked or within a plasmid vector) or RNA; or with antigen-expressing recombinant bacterium or viruses (*e.g.*, vaccinia, fowlpox, adenovirus or lentivirus vectors). Prior to loading, the polypeptide may be covalently conjugated to an immunological partner that provides T cell help (*e.g.*, a carrier molecule). Alternatively, a dendritic cell may be pulsed with a non-conjugated immunological partner, separately or in the presence of the polypeptide.

While any suitable carrier known to those of ordinary skill in the art may be employed in the pharmaceutical compositions of this invention, the type of carrier

will typically vary depending on the mode of administration. Compositions of the present invention may be formulated for any appropriate manner of administration, including for example, topical, oral, nasal, mucosal, intravenous, intracranial, intraperitoneal, subcutaneous and intramuscular administration.

5 Carriers for use within such pharmaceutical compositions are biocompatible, and may also be biodegradable. In certain embodiments, the formulation preferably provides a relatively constant level of active component release. In other embodiments, however, a more rapid rate of release immediately upon administration may be desired. The formulation of such compositions is well within the  
10 level of ordinary skill in the art using known techniques. Illustrative carriers useful in this regard include microparticles of poly(lactide-co-glycolide), polyacrylate, latex, starch, cellulose, dextran and the like. Other illustrative delayed-release carriers include supramolecular biovectors, which comprise a non-liquid hydrophilic core (*e.g.*, a cross-linked polysaccharide or oligosaccharide) and, optionally, an external layer  
15 comprising an amphiphilic compound, such as a phospholipid (*see e.g.*, U.S. Patent No. 5,151,254 and PCT applications WO 94/20078, WO/94/23701 and WO 96/06638). The amount of active compound contained within a sustained release formulation depends upon the site of implantation, the rate and expected duration of release and the nature of the condition to be treated or prevented.

20 In another illustrative embodiment, biodegradable microspheres (*e.g.*, polylactate polyglycolate) are employed as carriers for the compositions of this invention. Suitable biodegradable microspheres are disclosed, for example, in U.S. Patent Nos. 4,897,268; 5,075,109; 5,928,647; 5,811,128; 5,820,883; 5,853,763; 5,814,344, 5,407,609 and 5,942,252. Modified hepatitis B core protein carrier systems.  
25 such as described in WO/99 40934, and references cited therein, will also be useful for many applications. Another illustrative carrier/delivery system employs a carrier comprising particulate-protein complexes, such as those described in U.S. Patent No. 5,928,647, which are capable of inducing a class I-restricted cytotoxic T lymphocyte responses in a host.

30 The pharmaceutical compositions of the invention will often further comprise one or more buffers (*e.g.*, neutral buffered saline or phosphate buffered

saline), carbohydrates (e.g., glucose, mannose, sucrose or dextrans), mannitol, proteins, polypeptides or amino acids such as glycine, antioxidants, bacteriostats, chelating agents such as EDTA or glutathione, adjuvants (e.g., aluminum hydroxide), solutes that render the formulation isotonic, hypotonic or weakly hypertonic with the blood of a recipient, suspending agents, thickening agents and/or preservatives. Alternatively, compositions of the present invention may be formulated as a lyophilizate.

The pharmaceutical compositions described herein may be presented in unit-dose or multi-dose containers, such as sealed ampoules or vials. Such containers are typically sealed in such a way to preserve the sterility and stability of the formulation until use. In general, formulations may be stored as suspensions, solutions or emulsions in oily or aqueous vehicles. Alternatively, a pharmaceutical composition may be stored in a freeze-dried condition requiring only the addition of a sterile liquid carrier immediately prior to use.

The development of suitable dosing and treatment regimens for using the particular compositions described herein in a variety of treatment regimens, including e.g., oral, parenteral, intravenous, intranasal, and intramuscular administration and formulation, is well known in the art, some of which are briefly discussed below for general purposes of illustration.

In certain applications, the pharmaceutical compositions disclosed herein may be delivered *via* oral administration to an animal. As such, these compositions may be formulated with an inert diluent or with an assimilable edible carrier, or they may be enclosed in hard- or soft-shell gelatin capsule, or they may be compressed into tablets, or they may be incorporated directly with the food of the diet.

The active compounds may even be incorporated with excipients and used in the form of ingestible tablets, buccal tables, troches, capsules, elixirs, suspensions, syrups, wafers, and the like (see, for example, Mathiowitz *et al.*, Nature 1997 Mar 27;386(6623):410-4; Hwang *et al.*, Crit Rev Ther Drug Carrier Syst 1998;15(3):243-84; U. S. Patent 5,641,515; U. S. Patent 5,580,579 and U. S. Patent 5,792,451). Tablets, troches, pills, capsules and the like may also contain any of a variety of additional components, for example, a binder, such as gum tragacanth, acacia, cornstarch, or gelatin; excipients, such as dicalcium phosphate; a disintegrating agent,

such as corn starch, potato starch, alginic acid and the like; a lubricant, such as magnesium stearate; and a sweetening agent, such as sucrose, lactose or saccharin may be added or a flavoring agent, such as peppermint, oil of wintergreen, or cherry flavoring. When the dosage unit form is a capsule, it may contain, in addition to materials of the above type, a liquid carrier. Various other materials may be present as coatings or to otherwise modify the physical form of the dosage unit. For instance, tablets, pills, or capsules may be coated with shellac, sugar, or both. Of course, any material used in preparing any dosage unit form should be pharmaceutically pure and substantially non-toxic in the amounts employed. In addition, the active compounds may be incorporated into sustained-release preparation and formulations.

Typically, these formulations will contain at least about 0.1% of the active compound or more, although the percentage of the active ingredient(s) may, of course, be varied and may conveniently be between about 1 or 2% and about 60% or 70% or more of the weight or volume of the total formulation. Naturally, the amount of active compound(s) in each therapeutically useful composition may be prepared in such a way that a suitable dosage will be obtained in any given unit dose of the compound. Factors such as solubility, bioavailability, biological half-life, route of administration, product shelf life, as well as other pharmacological considerations will be contemplated by one skilled in the art of preparing such pharmaceutical formulations, and as such, a variety of dosages and treatment regimens may be desirable.

For oral administration the compositions of the present invention may alternatively be incorporated with one or more excipients in the form of a mouthwash, dentifrice, buccal tablet, oral spray, or sublingual orally-administered formulation. Alternatively, the active ingredient may be incorporated into an oral solution such as one containing sodium borate, glycerin and potassium bicarbonate, or dispersed in a dentifrice, or added in a therapeutically-effective amount to a composition that may include water, binders, abrasives, flavoring agents, foaming agents, and humectants. Alternatively the compositions may be fashioned into a tablet or solution form that may be placed under the tongue or otherwise dissolved in the mouth.

In certain circumstances it will be desirable to deliver the pharmaceutical compositions disclosed herein parenterally, intravenously, intramuscularly, or even

intraperitoneally. Such approaches are well known to the skilled artisan, some of which are further described, for example, in U. S. Patent 5,543,158; U. S. Patent 5,641,515 and U. S. Patent 5,399,363. In certain embodiments, solutions of the active compounds as free base or pharmacologically acceptable salts may be prepared in water suitably  
5 mixed with a surfactant, such as hydroxypropylcellulose. Dispersions may also be prepared in glycerol, liquid polyethylene glycols, and mixtures thereof and in oils. Under ordinary conditions of storage and use, these preparations generally will contain a preservative to prevent the growth of microorganisms.

Illustrative pharmaceutical forms suitable for injectable use include  
10 sterile aqueous solutions or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersions (for example, see U. S. Patent 5,466,468). In all cases the form must be sterile and must be fluid to the extent that easy syringability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms,  
15 such as bacteria and fungi. The carrier can be a solvent or dispersion medium containing, for example, water, ethanol, polyol (*e.g.*, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), suitable mixtures thereof, and/or vegetable oils. Proper fluidity may be maintained, for example, by the use of a coating, such as lecithin, by the maintenance of the required particle size in the case of dispersion and/or  
20 by the use of surfactants. The prevention of the action of microorganisms can be facilitated by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in  
25 the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

In one embodiment, for parenteral administration in an aqueous solution, the solution should be suitably buffered if necessary and the liquid diluent first rendered isotonic with sufficient saline or glucose. These particular aqueous solutions are  
30 especially suitable for intravenous, intramuscular, subcutaneous and intraperitoneal administration. In this connection, a sterile aqueous medium that can be employed will

be known to those of skill in the art in light of the present disclosure. For example, one dosage may be dissolved in 1 ml of isotonic NaCl solution and either added to 1000 ml of hypodermoclysis fluid or injected at the proposed site of infusion, (see for example, "Remington's Pharmaceutical Sciences" 15th Edition, pages 1035-1038 and 1570-  
5 1580). Some variation in dosage will necessarily occur depending on the condition of the subject being treated. Moreover, for human administration, preparations will of course preferably meet sterility, pyrogenicity, and the general safety and purity standards as required by FDA Office of Biologics standards.

In another embodiment of the invention, the compositions disclosed  
10 herein may be formulated in a neutral or salt form. Illustrative pharmaceutically-acceptable salts include the acid addition salts (formed with the free amino groups of the protein) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups can also be  
15 derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, histidine, procaine and the like. Upon formulation, solutions will be administered in a manner compatible with the dosage formulation and in such amount as is therapeutically effective.

20 The carriers can further comprise any and all solvents, dispersion media, vehicles, coatings, diluents, antibacterial and antifungal agents, isotonic and absorption delaying agents, buffers, carrier solutions, suspensions, colloids, and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active  
25 ingredient, its use in the therapeutic compositions is contemplated. Supplementary active ingredients can also be incorporated into the compositions. The phrase "pharmaceutically-acceptable" refers to molecular entities and compositions that do not produce an allergic or similar untoward reaction when administered to a human.

In certain embodiments, the pharmaceutical compositions may be  
30 delivered by intranasal sprays, inhalation, and/or other aerosol delivery vehicles. Methods for delivering genes, nucleic acids, and peptide compositions directly to the

lungs *via* nasal aerosol sprays has been described, *e.g.*, in U. S. Patent 5,756,353 and U. S. Patent 5,804,212. Likewise, the delivery of drugs using intranasal microparticle resins (Takenaga *et al.*, J Controlled Release 1998 Mar 2;52(1-2):81-7) and lysophosphatidyl-glycerol compounds (U. S. Patent 5,725,871) are also well-known in  
5 the pharmaceutical arts. Likewise, illustrative transmucosal drug delivery in the form of a polytetrafluoroethylene support matrix is described in U. S. Patent 5,780,045.

In certain embodiments, liposomes, nanocapsules, microparticles, lipid particles, vesicles, and the like, are used for the introduction of the compositions of the present invention into suitable host cells/organisms. In particular, the compositions of  
10 the present invention may be formulated for delivery either encapsulated in a lipid particle, a liposome, a vesicle, a nanosphere, or a nanoparticle or the like. Alternatively, compositions of the present invention can be bound, either covalently or non-covalently, to the surface of such carrier vehicles.

The formation and use of liposome and liposome-like preparations as  
15 potential drug carriers is generally known to those of skill in the art (see for example, Lasic, Trends Biotechnol 1998 Jul;16(7):307-21; Takakura, Nippon Rinsho 1998 Mar;56(3):691-5; Chandran *et al.*, Indian J Exp Biol. 1997 Aug;35(8):801-9; Margalit, Crit Rev Ther Drug Carrier Syst. 1995;12(2-3):233-61; U.S. Patent 5,567,434; U.S. Patent 5,552,157; U.S. Patent 5,565,213; U.S. Patent 5,738,868 and U.S. Patent  
20 5,795,587, each specifically incorporated herein by reference in its entirety).

Liposomes have been used successfully with a number of cell types that are normally difficult to transfect by other procedures, including T cell suspensions, primary hepatocyte cultures and PC 12 cells (Renneisen *et al.*, J Biol Chem. 1990 Sep 25;265(27):16337-42; Muller *et al.*, DNA Cell Biol. 1990 Apr;9(3):221-9). In addition,  
25 liposomes are free of the DNA length constraints that are typical of viral-based delivery systems. Liposomes have been used effectively to introduce genes, various drugs, radiotherapeutic agents, enzymes, viruses, transcription factors, allosteric effectors and the like, into a variety of cultured cell lines and animals. Furthermore, the use of liposomes does not appear to be associated with autoimmune responses or unacceptable  
30 toxicity after systemic delivery.

In certain embodiments, liposomes are formed from phospholipids that are dispersed in an aqueous medium and spontaneously form multilamellar concentric bilayer vesicles (also termed multilamellar vesicles (MLVs)).

Alternatively, in other embodiments, the invention provides for  
5 pharmaceutically-acceptable nanocapsule formulations of the compositions of the present invention. Nanocapsules can generally entrap compounds in a stable and reproducible way (see, for example, Quintanar-Guerrero *et al.*, Drug Dev Ind Pharm. 1998 Dec;24(12):1113-28). To avoid side effects due to intracellular polymeric overloading, such ultrafine particles (sized around 0.1  $\mu\text{m}$ ) may be designed using  
10 polymers able to be degraded *in vivo*. Such particles can be made as described, for example, by Couvreur *et al.*, Crit Rev Ther Drug Carrier Syst. 1988;5(1):1-20; zur Muhlen *et al.*, Eur J Pharm Biopharm. 1998 Mar;45(2):149-55; Zambaux *et al.* J Controlled Release. 1998 Jan 2;50(1-3):31-40; and U. S. Patent 5,145,684.

#### Cancer Therapeutic Methods

15 In further aspects of the present invention, the pharmaceutical compositions described herein may be used for the treatment of cancer, particularly for the immunotherapy of ovarian cancer. Within such methods, the pharmaceutical compositions described herein are administered to a patient, typically a warm-blooded animal, preferably a human. A patient may or may not be afflicted with cancer.  
20 Accordingly, the above pharmaceutical compositions may be used to prevent the development of a cancer or to treat a patient afflicted with a cancer. Pharmaceutical compositions and vaccines may be administered either prior to or following surgical removal of primary tumors and/or treatment such as administration of radiotherapy or conventional chemotherapeutic drugs. As discussed above, administration of the  
25 pharmaceutical compositions may be by any suitable method, including administration by intravenous, intraperitoneal, intramuscular, subcutaneous, intranasal, intradermal, anal, vaginal, topical and oral routes.

Within certain embodiments, immunotherapy may be active immunotherapy, in which treatment relies on the *in vivo* stimulation of the endogenous  
30 host immune system to react against tumors with the administration of immune

response-modifying agents (such as polypeptides and polynucleotides as provided herein).

Within other embodiments, immunotherapy may be passive immunotherapy, in which treatment involves the delivery of agents with established  
5 tumor-immune reactivity (such as effector cells or antibodies) that can directly or indirectly mediate antitumor effects and does not necessarily depend on an intact host immune system. Examples of effector cells include T cells as discussed above, T lymphocytes (such as CD8<sup>+</sup> cytotoxic T lymphocytes and CD4<sup>+</sup> T-helper tumor-infiltrating lymphocytes), killer cells (such as Natural Killer cells and lymphokine-  
10 activated killer cells), B cells and antigen-presenting cells (such as dendritic cells and macrophages) expressing a polypeptide provided herein. T cell receptors and antibody receptors specific for the polypeptides recited herein may be cloned, expressed and transferred into other vectors or effector cells for adoptive immunotherapy. The polypeptides provided herein may also be used to generate antibodies or anti-idiotypic  
15 antibodies (as described above and in U.S. Patent No. 4,918,164) for passive immunotherapy.

Effector cells may generally be obtained in sufficient quantities for adoptive immunotherapy by growth *in vitro*, as described herein. Culture conditions for expanding single antigen-specific effector cells to several billion in number with  
20 retention of antigen recognition *in vivo* are well known in the art. Such *in vitro* culture conditions typically use intermittent stimulation with antigen, often in the presence of cytokines (such as IL-2) and non-dividing feeder cells. As noted above, immunoreactive polypeptides as provided herein may be used to rapidly expand antigen-specific T cell cultures in order to generate a sufficient number of cells for  
25 immunotherapy. In particular, antigen-presenting cells, such as dendritic, macrophage, monocyte, fibroblast and/or B cells, may be pulsed with immunoreactive polypeptides or transfected with one or more polynucleotides using standard techniques well known in the art. For example, antigen-presenting cells can be transfected with a polynucleotide having a promoter appropriate for increasing expression in a  
30 recombinant virus or other expression system. Cultured effector cells for use in therapy must be able to grow and distribute widely, and to survive long term *in vivo*. Studies

have shown that cultured effector cells can be induced to grow *in vivo* and to survive long term in substantial numbers by repeated stimulation with antigen supplemented with IL-2 (*see, for example, Cheever et al., Immunological Reviews 157:177, 1997*).

Alternatively, a vector expressing a polypeptide recited herein may be  
5 introduced into antigen presenting cells taken from a patient and clonally propagated *ex vivo* for transplant back into the same patient. Transfected cells may be reintroduced into the patient using any means known in the art, preferably in sterile form by intravenous, intracavitary, intraperitoneal or intratumor administration.

Routes and frequency of administration of the therapeutic compositions  
10 described herein, as well as dosage, will vary from individual to individual, and may be readily established using standard techniques. In general, the pharmaceutical compositions and vaccines may be administered by injection (*e.g.*, intracutaneous, intramuscular, intravenous or subcutaneous), intranasally (*e.g.*, by aspiration) or orally. Preferably, between 1 and 10 doses may be administered over a 52 week period.  
15 Preferably, 6 doses are administered, at intervals of 1 month, and booster vaccinations may be given periodically thereafter. Alternate protocols may be appropriate for individual patients. A suitable dose is an amount of a compound that, when administered as described above, is capable of promoting an anti-tumor immune response, and is at least 10-50% above the basal (*i.e.*, untreated) level. Such response  
20 can be monitored by measuring the anti-tumor antibodies in a patient or by vaccine-dependent generation of cytolytic effector cells capable of killing the patient's tumor cells *in vitro*. Such vaccines should also be capable of causing an immune response that leads to an improved clinical outcome (*e.g.*, more frequent remissions, complete or partial or longer disease-free survival) in vaccinated patients as compared to non-  
25 vaccinated patients. In general, for pharmaceutical compositions and vaccines comprising one or more polypeptides, the amount of each polypeptide present in a dose ranges from about 25  $\mu$ g to 5 mg per kg of host. Suitable dose sizes will vary with the size of the patient, but will typically range from about 0.1 mL to about 5 mL.

In general, an appropriate dosage and treatment regimen provides the  
30 active compound(s) in an amount sufficient to provide therapeutic and/or prophylactic benefit. Such a response can be monitored by establishing an improved clinical

outcome (e.g., more frequent remissions, complete or partial, or longer disease-free survival) in treated patients as compared to non-treated patients. Increases in preexisting immune responses to a tumor protein generally correlate with an improved clinical outcome. Such immune responses may generally be evaluated using standard  
5 proliferation, cytotoxicity or cytokine assays, which may be performed using samples obtained from a patient before and after treatment.

#### Cancer Detection and Diagnostic Compositions, Methods and Kits

In general, a cancer may be detected in a patient based on the presence of one or more ovarian tumor proteins and/or polynucleotides encoding such proteins in a  
10 biological sample (for example, blood, sera, sputum urine and/or tumor biopsies) obtained from the patient. In other words, such proteins may be used as markers to indicate the presence or absence of a cancer such as ovarian cancer. In addition, such proteins may be useful for the detection of other cancers. The binding agents provided herein generally permit detection of the level of antigen that binds to the agent in the  
15 biological sample. Polynucleotide primers and probes may be used to detect the level of mRNA encoding an ovarian tumor protein, which is also indicative of the presence or absence of a cancer. In general, a ovarian tumor sequence should be present at a level that is at least three fold higher in tumor tissue than in normal tissue

There are a variety of assay formats known to those of ordinary skill in  
20 the art for using a binding agent to detect polypeptide markers in a sample. See, e.g., Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, 1988. In general, the presence or absence of a cancer in a patient may be determined by (a) contacting a biological sample obtained from a patient with a binding agent; (b) detecting in the sample a level of polypeptide that binds to the binding agent; and (c)  
25 comparing the level of polypeptide with a predetermined cut-off value.

In a preferred embodiment, the assay involves the use of binding agent immobilized on a solid support to bind to and remove the polypeptide from the remainder of the sample. The bound polypeptide may then be detected using a detection reagent that contains a reporter group and specifically binds to the binding  
30 agent/polypeptide complex. Such detection reagents may comprise, for example, a

binding agent that specifically binds to the polypeptide or an antibody or other agent that specifically binds to the binding agent, such as an anti-immunoglobulin, protein G, protein A or a lectin. Alternatively, a competitive assay may be utilized, in which a polypeptide is labeled with a reporter group and allowed to bind to the immobilized binding agent after incubation of the binding agent with the sample. The extent to which components of the sample inhibit the binding of the labeled polypeptide to the binding agent is indicative of the reactivity of the sample with the immobilized binding agent. Suitable polypeptides for use within such assays include full length ovarian tumor proteins and polypeptide portions thereof to which the binding agent binds, as described above.

The solid support may be any material known to those of ordinary skill in the art to which the tumor protein may be attached. For example, the solid support may be a test well in a microtiter plate or a nitrocellulose or other suitable membrane. Alternatively, the support may be a bead or disc, such as glass, fiberglass, latex or a plastic material such as polystyrene or polyvinylchloride. The support may also be a magnetic particle or a fiber optic sensor, such as those disclosed, for example, in U.S. Patent No. 5,359,681. The binding agent may be immobilized on the solid support using a variety of techniques known to those of skill in the art, which are amply described in the patent and scientific literature. In the context of the present invention, the term "immobilization" refers to both noncovalent association, such as adsorption, and covalent attachment (which may be a direct linkage between the agent and functional groups on the support or may be a linkage by way of a cross-linking agent). Immobilization by adsorption to a well in a microtiter plate or to a membrane is preferred. In such cases, adsorption may be achieved by contacting the binding agent, in a suitable buffer, with the solid support for a suitable amount of time. The contact time varies with temperature, but is typically between about 1 hour and about 1 day. In general, contacting a well of a plastic microtiter plate (such as polystyrene or polyvinylchloride) with an amount of binding agent ranging from about 10 ng to about 10  $\mu$ g, and preferably about 100 ng to about 1  $\mu$ g, is sufficient to immobilize an adequate amount of binding agent.

Covalent attachment of binding agent to a solid support may generally be achieved by first reacting the support with a bifunctional reagent that will react with both the support and a functional group, such as a hydroxyl or amino group, on the binding agent. For example, the binding agent may be covalently attached to supports  
5 having an appropriate polymer coating using benzoquinone or by condensation of an aldehyde group on the support with an amine and an active hydrogen on the binding partner (*see, e.g.*, Pierce Immunotechnology Catalog and Handbook, 1991, at A12-A13).

In certain embodiments, the assay is a two-antibody sandwich assay.  
10 This assay may be performed by first contacting an antibody that has been immobilized on a solid support, commonly the well of a microtiter plate, with the sample, such that polypeptides within the sample are allowed to bind to the immobilized antibody. Unbound sample is then removed from the immobilized polypeptide-antibody complexes and a detection reagent (preferably a second antibody capable of binding to a  
15 different site on the polypeptide) containing a reporter group is added. The amount of detection reagent that remains bound to the solid support is then determined using a method appropriate for the specific reporter group.

More specifically, once the antibody is immobilized on the support as described above, the remaining protein binding sites on the support are typically  
20 blocked. Any suitable blocking agent known to those of ordinary skill in the art, such as bovine serum albumin or Tween 20™ (Sigma Chemical Co., St. Louis, MO). The immobilized antibody is then incubated with the sample, and polypeptide is allowed to bind to the antibody. The sample may be diluted with a suitable diluent, such as phosphate-buffered saline (PBS) prior to incubation. In general, an appropriate contact  
25 time (*i.e.*, incubation time) is a period of time that is sufficient to detect the presence of polypeptide within a sample obtained from an individual with ovarian cancer. Preferably, the contact time is sufficient to achieve a level of binding that is at least about 95% of that achieved at equilibrium between bound and unbound polypeptide. Those of ordinary skill in the art will recognize that the time necessary to achieve  
30 equilibrium may be readily determined by assaying the level of binding that occurs over

a period of time. At room temperature, an incubation time of about 30 minutes is generally sufficient.

Unbound sample may then be removed by washing the solid support with an appropriate buffer, such as PBS containing 0.1% Tween 20™. The second  
5 antibody, which contains a reporter group, may then be added to the solid support. Preferred reporter groups include those groups recited above.

The detection reagent is then incubated with the immobilized antibody-polypeptide complex for an amount of time sufficient to detect the bound polypeptide. An appropriate amount of time may generally be determined by assaying the level of  
10 binding that occurs over a period of time. Unbound detection reagent is then removed and bound detection reagent is detected using the reporter group. The method employed for detecting the reporter group depends upon the nature of the reporter group. For radioactive groups, scintillation counting or autoradiographic methods are generally appropriate. Spectroscopic methods may be used to detect dyes, luminescent groups  
15 and fluorescent groups. Biotin may be detected using avidin, coupled to a different reporter group (commonly a radioactive or fluorescent group or an enzyme). Enzyme reporter groups may generally be detected by the addition of substrate (generally for a specific period of time), followed by spectroscopic or other analysis of the reaction products.

20 To determine the presence or absence of a cancer, such as ovarian cancer, the signal detected from the reporter group that remains bound to the solid support is generally compared to a signal that corresponds to a predetermined cut-off value. In one preferred embodiment, the cut-off value for the detection of a cancer is the average mean signal obtained when the immobilized antibody is incubated with  
25 samples from patients without the cancer. In general, a sample generating a signal that is three standard deviations above the predetermined cut-off value is considered positive for the cancer. In an alternate preferred embodiment, the cut-off value is determined using a Receiver Operator Curve, according to the method of Sackett et al., *Clinical Epidemiology: A Basic Science for Clinical Medicine*, Little Brown and Co., 1985,  
30 p. 106-7. Briefly, in this embodiment, the cut-off value may be determined from a plot of pairs of true positive rates (*i.e.*, sensitivity) and false positive rates (100%-specificity)

that correspond to each possible cut-off value for the diagnostic test result. The cut-off value on the plot that is the closest to the upper left-hand corner (*i.e.*, the value that encloses the largest area) is the most accurate cut-off value, and a sample generating a signal that is higher than the cut-off value determined by this method may be considered  
5 positive. Alternatively, the cut-off value may be shifted to the left along the plot, to minimize the false positive rate, or to the right, to minimize the false negative rate. In general, a sample generating a signal that is higher than the cut-off value determined by this method is considered positive for a cancer.

In a related embodiment, the assay is performed in a flow-through or  
10 strip test format, wherein the binding agent is immobilized on a membrane, such as nitrocellulose. In the flow-through test, polypeptides within the sample bind to the immobilized binding agent as the sample passes through the membrane. A second, labeled binding agent then binds to the binding agent-polypeptide complex as a solution containing the second binding agent flows through the membrane. The detection of  
15 bound second binding agent may then be performed as described above. In the strip test format, one end of the membrane to which binding agent is bound is immersed in a solution containing the sample. The sample migrates along the membrane through a region containing second binding agent and to the area of immobilized binding agent. Concentration of second binding agent at the area of immobilized antibody indicates the  
20 presence of a cancer. Typically, the concentration of second binding agent at that site generates a pattern, such as a line, that can be read visually. The absence of such a pattern indicates a negative result. In general, the amount of binding agent immobilized on the membrane is selected to generate a visually discernible pattern when the biological sample contains a level of polypeptide that would be sufficient to generate a  
25 positive signal in the two-antibody sandwich assay, in the format discussed above. Preferred binding agents for use in such assays are antibodies and antigen-binding fragments thereof. Preferably, the amount of antibody immobilized on the membrane ranges from about 25 ng to about 1 $\mu$ g, and more preferably from about 50 ng to about 500 ng. Such tests can typically be performed with a very small amount of biological  
30 sample.

Of course, numerous other assay protocols exist that are suitable for use with the tumor proteins or binding agents of the present invention. The above descriptions are intended to be exemplary only. For example, it will be apparent to those of ordinary skill in the art that the above protocols may be readily modified to use  
5 tumor polypeptides to detect antibodies that bind to such polypeptides in a biological sample. The detection of such tumor protein specific antibodies may correlate with the presence of a cancer.

A cancer may also, or alternatively, be detected based on the presence of T cells that specifically react with a tumor protein in a biological sample. Within  
10 certain methods, a biological sample comprising CD4<sup>+</sup> and/or CD8<sup>+</sup> T cells isolated from a patient is incubated with a tumor polypeptide, a polynucleotide encoding such a polypeptide and/or an APC that expresses at least an immunogenic portion of such a polypeptide, and the presence or absence of specific activation of the T cells is detected. Suitable biological samples include, but are not limited to, isolated T cells. For  
15 example, T cells may be isolated from a patient by routine techniques (such as by Ficoll/Hypaque density gradient centrifugation of peripheral blood lymphocytes). T cells may be incubated *in vitro* for 2-9 days (typically 4 days) at 37°C with polypeptide (e.g., 5 - 25 µg/ml). It may be desirable to incubate another aliquot of a T cell sample in the absence of tumor polypeptide to serve as a control. For CD4<sup>+</sup> T cells, activation is  
20 preferably detected by evaluating proliferation of the T cells. For CD8<sup>+</sup> T cells, activation is preferably detected by evaluating cytolytic activity. A level of proliferation that is at least two fold greater and/or a level of cytolytic activity that is at least 20% greater than in disease-free patients indicates the presence of a cancer in the patient.

As noted above, a cancer may also, or alternatively, be detected based on  
25 the level of mRNA encoding a tumor protein in a biological sample. For example, at least two oligonucleotide primers may be employed in a polymerase chain reaction (PCR) based assay to amplify a portion of a tumor cDNA derived from a biological sample, wherein at least one of the oligonucleotide primers is specific for (*i.e.*, hybridizes to) a polynucleotide encoding the tumor protein. The amplified cDNA is  
30 then separated and detected using techniques well known in the art, such as gel electrophoresis. Similarly, oligonucleotide probes that specifically hybridize to a

polynucleotide encoding a tumor protein may be used in a hybridization assay to detect the presence of polynucleotide encoding the tumor protein in a biological sample.

To permit hybridization under assay conditions, oligonucleotide primers and probes should comprise an oligonucleotide sequence that has at least about 60%,  
5 preferably at least about 75% and more preferably at least about 90%, identity to a portion of a polynucleotide encoding a tumor protein of the invention that is at least 10 nucleotides, and preferably at least 20 nucleotides, in length. Preferably, oligonucleotide primers and/or probes hybridize to a polynucleotide encoding a polypeptide described herein under moderately stringent conditions, as defined above.  
10 Oligonucleotide primers and/or probes which may be usefully employed in the diagnostic methods described herein preferably are at least 10-40 nucleotides in length. In a preferred embodiment, the oligonucleotide primers comprise at least 10 contiguous nucleotides, more preferably at least 15 contiguous nucleotides, of a DNA molecule having a sequence as disclosed herein. Techniques for both PCR based assays and  
15 hybridization assays are well known in the art (*see, for example, Mullis et al., Cold Spring Harbor Symp. Quant. Biol.*, 51:263, 1987; Erlich ed., *PCR Technology*, Stockton Press, NY, 1989).

One preferred assay employs RT-PCR, in which PCR is applied in conjunction with reverse transcription. Typically, RNA is extracted from a biological  
20 sample, such as biopsy tissue, and is reverse transcribed to produce cDNA molecules. PCR amplification using at least one specific primer generates a cDNA molecule, which may be separated and visualized using, for example, gel electrophoresis. Amplification may be performed on biological samples taken from a test patient and from an individual who is not afflicted with a cancer. The amplification reaction may be  
25 performed on several dilutions of cDNA spanning two orders of magnitude. A two-fold or greater increase in expression in several dilutions of the test patient sample as compared to the same dilutions of the non-cancerous sample is typically considered positive.

In another embodiment, the compositions described herein may be used  
30 as markers for the progression of cancer. In this embodiment, assays as described above for the diagnosis of a cancer may be performed over time, and the change in the level of

reactive polypeptide(s) or polynucleotide(s) evaluated. For example, the assays may be performed every 24-72 hours for a period of 6 months to 1 year, and thereafter performed as needed. In general, a cancer is progressing in those patients in whom the level of polypeptide or polynucleotide detected increases over time. In contrast, the  
5 cancer is not progressing when the level of reactive polypeptide or polynucleotide either remains constant or decreases with time.

Certain *in vivo* diagnostic assays may be performed directly on a tumor. One such assay involves contacting tumor cells with a binding agent. The bound binding agent may then be detected directly or indirectly via a reporter group. Such  
10 binding agents may also be used in histological applications. Alternatively, polynucleotide probes may be used within such applications.

As noted above, to improve sensitivity, multiple tumor protein markers may be assayed within a given sample. It will be apparent that binding agents specific for different proteins provided herein may be combined within a single assay. Further,  
15 multiple primers or probes may be used concurrently. The selection of tumor protein markers may be based on routine experiments to determine combinations that results in optimal sensitivity. In addition, or alternatively, assays for tumor proteins provided herein may be combined with assays for other known tumor antigens.

The present invention further provides kits for use within any of the  
20 above diagnostic methods. Such kits typically comprise two or more components necessary for performing a diagnostic assay. Components may be compounds, reagents, containers and/or equipment. For example, one container within a kit may contain a monoclonal antibody or fragment thereof that specifically binds to a tumor protein. Such antibodies or fragments may be provided attached to a support material, as  
25 described above. One or more additional containers may enclose elements, such as reagents or buffers, to be used in the assay. Such kits may also, or alternatively, contain a detection reagent as described above that contains a reporter group suitable for direct or indirect detection of antibody binding.

Alternatively, a kit may be designed to detect the level of mRNA  
30 encoding a tumor protein in a biological sample. Such kits generally comprise at least one oligonucleotide probe or primer, as described above, that hybridizes to a

polynucleotide encoding a tumor protein. Such an oligonucleotide may be used, for example, within a PCR or hybridization assay. Additional components that may be present within such kits include a second oligonucleotide and/or a diagnostic reagent or container to facilitate the detection of a polynucleotide encoding a tumor protein.

5           The following Examples are offered by way of illustration and not by way of limitation.

## EXAMPLES

### EXAMPLE 1

#### 10           IDENTIFICATION OF REPRESENTATIVE OVARIAN CARCINOMA cDNA SEQUENCES

Primary ovarian tumor and metastatic ovarian tumor cDNA libraries were each constructed in kanamycin resistant pZErO™-2 vector (Invitrogen) from pools of three different ovarian tumor RNA samples. For the primary ovarian tumor library, the following RNA samples were used: (1) a moderately differentiated papillary serous carcinoma of a 41 year old, (2) a stage IIIC ovarian tumor and (3) a papillary serous  
15           adenocarcinoma for a 50 year old caucasian. For the metastatic ovarian tumor library, the RNA samples used were omentum tissue from: (1) a metastatic poorly differentiated papillary adenocarcinoma with psammoma bodies in a 73 year old, (2) a metastatic poorly differentiated adenocarcinoma in a 74 year old and (3) a metastatic  
20           poorly differentiated papillary adenocarcinoma in a 68 year old.

The number of clones in each library was estimated by plating serial dilutions of unamplified libraries. Insert data were determined from 32 primary ovarian tumor clones and 32 metastatic ovarian tumor clones. The library characterization results are shown in Table I.

25

Table I  
Characterization of cDNA Libraries

Library	# Clones in Library	Clones with Insert (%)	Insert Size Range (bp)	Ave. Insert Size (bp)
Primary Ovarian Tumor	1,258,000	97	175 - 8000	2356
Metastatic Ovarian Tumor	1,788,000	100	150 - 4300	1755

5

Four subtraction libraries were constructed in ampicillin resistant pcDNA3.1 vector (Invitrogen). Two of the libraries were from primary ovarian tumors and two were from metastatic ovarian tumors. In each case, the number of restriction enzyme cuts within inserts was minimized to generate full length subtraction libraries.

10 The subtractions were each done with slightly different protocols, as described in more detail below.

A. POTS 2 Library: Primary Ovarian Tumor Subtraction Library

Tracer: 10 µg primary ovarian tumor library, digested with Not I

15 Driver: 35 µg normal pancreas in pcDNA3.1(+)

20 µg normal PBMC in pcDNA3.1(+)

10 µg normal skin in pcDNA3.1(+)

35 µg normal bone marrow in pZErO™-2

Digested with Bam HI/Xho I/Sca I

20 Two hybridizations were performed, and Not I-cut pcDNA3.1(+) was the cloning vector for the subtracted library. Sequence results for previously unidentified clones that were randomly picked from the subtracted library are presented in Table II.

Table II  
Ovarian Carcinoma Sequences

25

Sequence	SEQ ID NO
21907	1
21909	2
21911	5
21920	9
21921	10

Sequence	SEQ ID NO
25099	143
25101	144
25103	145
25107	146
25111	148
25113	149
25115	150
25116	151
25752	156
25757	158
25763	160
25769	161
25770	162

**B. POTS 7 Library: Primary Ovarian Tumor Subtraction Library**

- Tracer: 10 µg primary ovarian tumor library, digested with Not I
- 5 Driver: 35 µg normal pancreas in pcDNA3.1(+)
- 20 µg normal PBMC in pcDNA3.1(+)
- 10 µg normal skin in pcDNA3.1(+)
- 35 µg normal bone marrow in pZErO™-2
- Digested with Bam HI/Xho I/Sca I
- 10 ~25 µg pZErO™-2, digested with Bam HI and Xho I

Two hybridizations were performed, and Not I-cut pcDNA3.1(+) was the cloning vector for the subtracted library. Sequence results for previously unidentified clones that were randomly picked from the subtracted library are presented in Table III.

15

**Table III**  
**Ovarian Carcinoma Sequences**

Sequence	SEQ ID NO
24937	125
24940	128
24946	132
24950	133
24951	134
24955	136
24956	137
25791	166

Sequence	SEQ ID NO
25796	167
25797	168
25804	171

### C. OS1D Library: Metastatic Ovarian Tumor Subtraction Library

5	Tracer:	10µg metastatic ovarian library in pZErO™-2, digested with Not I
	Driver:	24.5µg normal pancreas in pcDNA3.1
		14µg normal PBMC in pcDNA3.1
		14µg normal skin in pcDNA3.1
		24.5µg normal bone marrow in pZErO™-2
10		50µg pZErO™-2, digested with Bam HI/Xho I/Sfu I

Three hybridizations were performed, and the last two hybridizations were done with an additional 15µg of biotinylated pZErO™-2 to remove contaminating pZErO™-2 vectors. The cloning vector for the subtracted library was pcDNA3.1/Not I cut. Sequence results for previously unidentified clones that were randomly picked from the subtracted library are presented in Table IV.

**Table IV**  
**Ovarian Carcinoma Sequences**

Sequence	SEQ ID NO
23645.1	13
23660.1	16
23666.1	19
23679.1	23
24635	57
24647	63
24651	65
24661	69
24663	70
24664	71
24670	72
24675	75
24683	78

D. OSIF Library: Metastatic Ovarian Tumor Subtraction Library

Tracer: 10µg metastatic ovarian tumor library, digested with Not I

Driver: 12.8µg normal pancreas in pcDNA3.1

5 7.3µg normal PBMC in pcDNA3.1

7.3µg normal skin in pcDNA3.1

12.8µg normal bone marrow in pZErO™-2

25µg pZErO™-2, digested with Bam HI/Xho I/Sfu I

One hybridization was performed. The cloning vector for the subtracted  
10 library was pcDNA3.1/Not I cut. Sequence results for previously unidentified clones  
that were randomly picked from the subtracted library are presented in Table V.

Table V

Ovarian Carcinoma Sequences

Sequence	SEQ ID NO
24336 (79% with H. sapiens mitochondrial genome (consensus sequence))	27
24337	28
24341 (91% Homo sapiens chromosome 5, BAC clone 249h5 (LBNL H149))	32
24344	33
24348	35
24351	38
24355 (91% Homo sapiens chromosome 17, clone hCIT.91 J 4)	41
24356	42
24357 (87% S. scrofa mRNA for UDP glucose pyrophosphorylase)	43
24358	44
24359 (78% Human mRNA for KIAA0111 gene, complete cds)	45
24360	46
24361	47
24362 (88% Homo sapiens Chromosome 16 BAC clone CIT987SK-A-233A7)	48
24363 (87% Homo sapiens eukaryotic translation elongation factor 1 alpha 1 (EEF1A1))	49
24364 (89% Human DNA sequence from PAC 27K14 on chromosome Xp11.3-Xp11.4)	50
24367 (89% Homo sapiens 12p13.3 BAC RCPII1-	52

Sequence	SEQ ID NO
935C2)	
24368	53
24690	81
24692	82
24694	84
24696	86
24699	89
24701	90
24703	91
24704 (88% Homo sapiens chromosome 9, clone hRPK.401_G_18)	92
24705	93
24707	95
24709	97
24711	98
24713	99
24714 (91% Human DNA sequence from clone 125N5 on chromosome 6q26-27)	100
24717 (89% Homo sapiens proliferation-associated gene A (natural killer-enhancing factor A) (PAGA)	103
24727	107
24732	111
24737 (84% Human ADP/ATP translocase mRNA)	114
24741	117
24745	120
24746	121

The sequences in Table VI, which correspond to known sequences, were also identified in the above libraries.

5

Table VI  
Ovarian Carcinoma Sequences

Identity	SEQ ID NO	Sequence	Library
H.sapiens DNA for muscle nicotinic acetylcholine receptor gene promotor, clone ICRFc105F02104	3	21910	POTS2
Homo sapiens complement component 3 (C3) gene, exons 1-30.	4	21913	POTS2
Homo sapiens SWI/SNF related, matrix associated, actin dependent regulator of chromatin, subfamily a, member 4 (SMARCA4)	6	21914	POTS2
Human ferritin Heavy subunit mRNA, complete cds.	7	21915	POTS2

Identity	SEQ ID NO	Sequence	Library
Homo sapiens CGI-151 protein mRNA, complete cds	8	21916	POTS2
Human BAC clone GS055K18 from 7p15-p21	11	23636.1	OS1D
HUMGFIBPA Human growth hormone-dependent insulin-like growth factor-binding protein	12	23637.1	OS1D
Homo sapiens ribosomal protein, large, P0 (RPLP0) mRNA	14	23647.1	OS1D
HUMTRPM2A Human TRPM-2 mRNA	15	23657.1	OS1D
HUMMTA Homo sapiens mitochondrial DNA	17	23661.1	OS1D
HSU78095 Homo sapiens placental bikunin mRNA	18	23662.1	OS1D
HUMTI227HC Human mRNA for TI-227H	20	23669.1	OS1D
HUMMTCG Human mitochondrion	21	23673.1	OS1D
Homo sapiens FK506-binding protein 1A (12kD) (FKBP1A) mRNA	22	23677.1	OS1D
Homo sapiens mRNA for zinc-finger DNA-binding protein, complete cds	24	24333	OS1F
Homo sapiens mRNA; cDNA DKFZp564E1962 (from clone DKFZp564E1962)	25	24334	OS1F
Homo sapiens tumor protein, translationally-controlled 1 (TPT1) mRNA.	26	24335	OS1F
Homo sapiens interleukin 1 receptor accessory protein (IL1RAP) mRNA.	29	24338	OS1F
Human mRNA for KIAA0026 gene	30	24339	OS1F
Homo sapiens K-Cl cotransporter KCC4 mRNA, complete cds	31	24340	OS1F
Homo sapiens nuclear chloride ion channel protein (NCC27) mRNA	34	24345	OS1F
Homo sapiens mRNA for DEPP (decidual protein induced by progesterone)	36	24349	OS1F
Homo sapiens atrophin-1 interacting protein 4 (AIP4) mRNA	37	24350	OS1F
Human collagenase type IV mRNA, 3' end.	39	24352	OS1F
Human mRNA for T-cell cyclophilin	40	24354	OS1F
Homo sapiens tumor suppressing subtransferable candidate 1 (TSSC1)	51	24366	OS1F
Homo sapiens clone 24452 mRNA sequence	54	24374	OS1F
Homo sapiens eukaryotic translation elongation factor 1 alpha 1 (EEF1A1)	55	24627	OS1D
Genomic sequence from Human 9q34	56	24634	OS1D
Human insulin-like growth factor-binding protein-3 gene	58	24636	OS1D
Human ribosomal protein L3 mRNA, 3' end	59	24638	OS1D
Homo sapiens annexin II (lipocortin II) (ANX2) mRNA	60	24640	OS1D
Homo sapiens tubulin, alpha, ubiquitous (K-ALPHA-1)	61	24642	OS1D

Identity	SEQ ID NO	Sequence	Library
Human non-histone chromosomal protein HMG-14 mRNA	62	24645	OS1D
Homo sapiens ferritin, heavy polypeptide 1 (FTH1)	64	24648	OS1D
Homo sapiens 12p13.3 PAC RPCII-96H9 (Roswell Park Cancer Institute Human PACLibrary)	66	24653	OS1D
Homo sapiens T cell-specific tyrosine kinase mRNA	67	24655	OS1D
Homo sapiens keratin 18 (KRT18) mRNA	68	24657	OS1D
Homo sapiens growth arrest specific transcript 5 gene	73	24671	OS1D
Homo sapiens ribosomal protein S7 (RPS7)	74	24673	OS1D
Homo sapiens mRNA; cDNA DKFZp564H182	76	24677	OS1D
Human TSC-22 protein mRNA	77	24679	OS1D
Human mRNA for ribosomal protein	79	24687	OS1D
Genomic sequence from Human 13	80	24689	OS1F
Homo sapiens clone IMAGE 286356	83	24693	OS1F
Homo sapiens v-fos FBJ murine osteosarcoma viral oncogene homolog(FOS) mRNA	85	24695	OS1F
Homo sapiens hypothetical 43.2 Kd protein mRNA	87	24697	OS1F
Human heat shock protein 27 (HSPB1) gene exons 1-3	88	24698	OS1F
Homo sapiens senescence-associated epithelial membrane protein (SEMP1)	94	24706	OS1F
Human ferritin H chain mRNA	96	24708	OS1F
Homo sapiens mRNA for KIAA0287 gene	101	24715	OS1F
Homo sapiens CGI-08 protein mRNA	102	24716	OS1F
H.sapiens CpG island DNA genomic MseI fragment, clone 84a5	104	24719	OS1F
Human clone 23722 mRNA	105	24721	OS1F
Homo sapiens zinc finger protein slug (SLUG) gene	106	24722	OS1F
Homo sapiens (clone L6) E-cadherin (CDH1) gene	108	24728	OS1F
Homo sapiens ribosomal protein L13 (RPL13)	109	24729	OS1F
H.sapiens RNA for snRNP protein B	110	24730	OS1F
Homo sapiens mRNA; cDNA DKFZp434K114	112	24734	OS1F
Homo sapiens cornichon protein mRNA	113	24735	OS1F
Homo sapiens keratin 8 (KRT8) mRNA	115	24739	OS1F
Human DNA sequence from PAC 29K1 on chromosome 6p21.3-22.2.	116	24740	OS1F
Homo sapiens mRNA for KIAA0762 protein	118	24742	OS1F
Human clones 23667 and 23775 zinc finger protein mRNA	119	24744	OS1F
Human H19 RNA gene, complete cds.	122	24933	POTS7
Human triosephosphate isomerase mRNA, complete cds.	123	24934	POTS7
Human cyclooxygenase-1 (PTSG1) mRNA, partial cds	124	24935	POTS7
Homo sapiens megakaryocyte potentiating factor	126	24938	POTS7

Identity	SEQ ID NO	Sequence	Library
(MPF) mRNA.			
Human mRNA for Apo1_Human (MER5(Aop1-Mouse)-like protein), complete cds	127	24939	POTS7
Homo sapiens arylacetamide deacetylase (esterase) (AADAC) mRNA.	129	24942	POTS7
Homo sapiens echinoderm microtubule-associated protein-like EMAP2 mRNA, complete cds	130	24943	POTS7
Homo sapiens podocalyxin-like (PODXL) mRNA.	131	24944	POTS7
Homo sapiens synaptogyrin 2 (SYNGR2) mRNA.	135	24952	POTS7
Homo sapiens amyloid beta precursor protein-binding protein 1, 59kD (APPBP1) mRNA.	138	24959	POTS7
Human aldose reductase mRNA, complete cds.	139	24969	POTS7
Genomic sequence from Human 9q34, complete sequence [Homo sapiens]	140	25092	POTS2
Human glyceraldehyde-3-phosphate dehydrogenase (GAPDH) mRNA, complete cds.	141	25093	POTS2
Homo sapiens breast cancer suppressor candidate 1 (bcsc-1) mRNA, complete cds	142	25098	POTS2
Homo sapiens SKB1 (S. cerevisiae) homolog (SKB1) mRNA.	147	25110	POTS2
Homo sapiens prepro dipeptidyl peptidase I (DPP-I) gene, complete cds	152	25117	POTS2
Homo sapiens preferentially expressed antigen of melanoma (PRAME) mRNA	153	25745	POTS2
Human translocated t(8;14) c-myc (MYC) oncogene, exon 3 and complete cds	154	25746	POTS2
Human 12S RNA induced by poly(rI), poly(rC) and Newcastle disease virus	155	25749	POTS2
Human mRNA for fibronectin (FN precursor)	157	25755	POTS2
Homo sapiens mRNA for hepatocyte growth factor activator inhibitor type 2, complete cds	159	25758	POTS2
Homo sapiens mRNA for KIAA0552 protein, complete cds	163	25771	POTS7
Homo sapiens IMP (inosine monophosphate) dehydrogenase 2 (IMPDH2) mRNA	164	25775	POTS7
Homo sapiens clone 23942 alpha enolase mRNA, partial cds	165	25787	POTS7
H.sapiens vegf gene, 3'UTR	169	25799	POTS7
Homo sapiens 30S ribosomal protein S7 homolog mRNA, complete cds	170	25802	POTS7
Homo sapiens acetyl-Coenzyme A acetyltransferase 2 (acetoacetyl Coenzyme A thiolase) (ACAT2) mRNA	172	25808	POTS7
Homo sapiens Norrie disease protein (NDP) mRNA	173	25809	POTS7

Still further ovarian carcinoma polynucleotide and/or polypeptide sequences identified from the above libraries are provided below in Table VII. Sequences O574S (SEQ ID NO:183 & 185), O584S (SEQ ID NO:193) and O585S (SEQ ID NO:194) represent novel sequences. The remaining sequences exhibited at least some homology with known genomic and/or EST sequences.

Table VII

SEQ ID:	Sequence	Library
174 :	O565S CRABP	OS1D
175 :	O566S Ceruloplasmin	POTS2
176 :	O567S 41191.SEQ(1>487)	POTS2
177 :	O568S KIAA0762.seq(1>3999)	POTS7
178 :	O569S 41220.seq(1>1069)	POTS7
179 :	O570S 41215.seq(1>1817)	POTS2
180:	O571S 41213.seq(1>2382)	POTS2
181 :	O572S 41208.seq(1>2377)	POTS2
182 :	O573S 41177.seq(1>1370)	OS1F
183 :	O574S 47807.seq(1>2060)	n/a
184 :	O568S/VSGF DNA seq	n/a
185:	O574S 47807.seq(1>3000)	n/a
186:	O568S/VSGF protein seq	n/a
187 :	449H1(57581)	OS1D
188:	451E12(57582)	OS1D
189 :	453C7 3'(57583.1)Osteonectin	OS1D
190 :	453C7 5'(57583.2)	OS1D
191:	456G1 3'(57584.1)Neurotensin	OS1F
192:	456G1 5'(57584.2)	OS1F
193:	O584S 465G5(57585)	OS1F
194:	O585S 469B12(57586)	POTS2
195:	O569S 474C3(57587)	POTS7
196:	483B1 3'(24934.1)Triosephosphate	POTS7
197:	57885 Human preferentially expressed antigen of melanoma	POTS2
198:	57886 Chromosome 22q12.1 clone CTA-723E4	POTS2
199:	57887 Homologous to mouse brain cDNA clone MNCb-0671	POTS2

Further studies on the clone of SEQ ID NO:182 (also referred to as O573S) led to the identification of multiple open reading frames that encode the amino acid sequences of SEQ ID NO:200-202.

## EXAMPLE 2

## ANALYSIS OF CDNA EXPRESSION USING MICROARRAY TECHNOLOGY

In additional studies, sequences disclosed herein were found to be overexpressed in specific tumor tissues as determined by microarray analysis. Using this approach, cDNA sequences are PCR amplified and their mRNA expression profiles in tumor and normal tissues are examined using cDNA microarray technology essentially as described (Shena et al., 1995). In brief, the clones are arrayed onto glass slides as multiple replicas, with each location corresponding to a unique cDNA clone (as many as 5500 clones can be arrayed on a single slide or chip). Each chip is hybridized with a pair of cDNA probes that are fluorescence-labeled with Cy3 and Cy5 respectively. Typically, 1 $\mu$ g of polyA<sup>+</sup> RNA is used to generate each cDNA probe. After hybridization, the chips are scanned and the fluorescence intensity recorded for both Cy3 and Cy5 channels. There are multiple built-in quality control steps. First, the probe quality is monitored using a panel of ubiquitously expressed genes. Secondly, the control plate also can include yeast DNA fragments of which complementary RNA may be spiked into the probe synthesis for measuring the quality of the probe and the sensitivity of the analysis. Currently, the technology offers a sensitivity of 1 in 100,000 copies of mRNA. Finally, the reproducibility of this technology can be ensured by including duplicated control cDNA elements at different locations.

The microarray results for clones 57885 (SEQ ID NO:197), 57886 (SEQ ID NO:198) and 57887 (SEQ ID NO:199) are as follows.

Clone 57885: 16/38 (42%) of ovarian tumors showed an expression signal value of >0.4. The mean value for all ovary tumors was 0.662 with a mean value of 0.187 for all normal tissues, which yields a 3.64 fold overexpression level in ovary tumor relative to essential normal tissues. Normal tissue expression was elevated (>0.4) in peritoneum, skin and thymus.

Clone 57886: 16/38 (42%) of ovarian tumors showed an expression signal value of >0.4. The mean value for all ovary tumors was 0.574 with a mean value of 0.166 for all normal tissues which yields a 3.46 fold overexpression level in ovary tumor relative to essential normal tissues. Normal tissue expression was elevated (>0.4) in heart, pancreas and small intestine.

Clone 57887: 17/38 (44%) of ovarian tumors showed an expression signal value of  $>0.4$ . The mean value for all ovary tumors is 0.744 with a mean value of 0.184 for all normal tissues which yields a 4.04 fold overexpression level in ovary tumor relative to essential normal tissues. Normal tissue expression was elevated ( $>0.4$ ) in esophagus.

### EXAMPLE 3

#### EXPRESSION OF RECOMBINANT ANTIGEN O568S IN E. COLI

This example describes the expression of recombinant antigen O568S (SEQ ID NO:177) in E. coli. This sequence was identified in Example 1 from the POTS 7 subtraction library using primary ovarian tumor cDNA as the tracer. PCR primers specific for the open reading frame of O568S were designed and used in the specific amplification of O568S. The PCR product was enzymatically digested with EcoRI and ligated into pPDM, a modified pET28 vector which had been cut with the restriction enzymes EcoRI and Eco72I. The construct sequence and orientation was confirmed through sequence analysis, the sequence of which is shown in SEQ ID NO:206. The vector was then transformed into the expression hosts, BLR (DE3) and HMS 174 (DE3) pLys S. Protein expression was confirmed, the sequence of which is provided in SEQ ID NO:207.

20

### EXAMPLE 4

#### ADDITIONAL SEQUENCE OBTAINED FOR CLONE O591S

The sequence of O591S (clone identifier 57887) was used to search public sequence databases. It was found that the reverse strand showed some degree of identity to the C-terminal end of GPR39. The cDNA for the coding region of GPR39 is disclosed in SEQ ID NO:208 and the corresponding amino acid sequence in SEQ ID NO:209. The GPR39 coding region contains two exons. Both O591S and GPR39, encoded by the complementary strand of O591S, are located on chromosome 2.

25

## EXAMPLE 5

## FURTHER CHARACTERIZATION OF O591S AND IDENTIFICATION OF EXTENDED SEQUENCE

O1034C is an ovary specific gene identified by electronic subtraction. Briefly, electronic subtraction involves an analysis of EST database sequences to  
5 identify ovarian-specific genes. In the electronic subtraction method used to identify O1034C, sequences of EST clones derived from ovary libraries (normal and tumor) were obtained from the GenBank public human EST database. Each ovary sequence was used as a "seed" query in a BLASTN search of the total human EST database to identify other EST clones that share sequence with the seed sequence (clones that potentially  
10 originated from the same mRNA). EST clones with shared sequence were grouped into clusters, and clusters that shared sequence with other clusters were grouped into superclusters. The tissue source of each EST within each supercluster was noted, and superclusters were ranked based on the distribution of the tissues from which the ESTs originated. Superclusters that comprise primarily, or solely, EST clones from ovary  
15 libraries were considered to represent genes that were differentially expressed in ovary tissue, relative to all other normal adult tissue.

This clone was identified from the public EST databases as Integrated Molecular Analysis of Genomics and their Expression (IMAGE) clone number 595449 (the IMAGE consortium is a repository of EST clones and cDNA clones) and is  
20 disclosed as SEQ ID NO:210. Accession numbers AA173739 and AA173383 represents the sequence of the identified EST in Genbank. This clone is part of Unigene cluster HS.85339 (Unigene is an experimental system for automatically partitioning Genbank sequences into a non-redundant set of gene-orientated clusters) and was annotated as encoding a neurotensin-like G protein coupled receptor (GRP39).  
25 However, the inventors have discovered that IMAGE#595449 encodes a novel protein derived from the complementary strand to that which encodes the potential GPR39.

Microarray analysis of the clone using a series of ovary tumor specific probes indicated that this clone was over expressed 4.95-fold in a group of ovary tumor and normal ovary samples as compared to a group of essential normal tissue samples.

30 IMAGE#59449 was subjected to a Blast A search of the EST database and Genbank and an electronic full length clone contig (O1034C) was generated by

extending IMAGE#595449 and its resulting contigs to completion. This process was repeated to completion when no further EST sequences were identified to extend the consensus sequence. This electronically derived clone was identified as coding a previously described clone, O591S, the sequence of which is disclosed in SEQ ID NO:211. The discovery of this ovary specific candidate is described in more detail in Example 4.

The consensus sequence for O1034C extended further 5' than O591S due to the additional sequences derived from two EST clones, accession numbers BF345141 and BE336607, the sequences for which are disclosed in SEQ ID NO:212 and 213 respectively. Although BF345141 diverges from the O1034C/O591S consensus at its 3'-end (possibly representing a different splice form), and from BE336607 at several bases at its 5'-end, the two ESTs were compared to the available matching chromosome sequence. They were found on human chromosome 2, clone RP11-159N20:htgs database accession number AC010974. These sequences were used to extend O1034C/O591S to form a final consensus sequence for O1034C/O591S of 1897 base pairs, disclosed in SEQ ID NO:214.

An open reading frame (ORF) was identified within the O1034C/O591S consensus sequence (nucleotides 260-682), the predicted translation of which is disclosed in SEQ ID NO:215. A BLASTx database search against the Genbank database indicated that this ORF had no identity (E value  $<1e-25$ ) with any known human protein. The only match was with the G protein-coupled receptors, including GPR39, which the inventors have shown to be encoded at the 3'-end of O1034C/O591S on the complementary strand. However, the ORF did encode a protein that had 93% similarity (131/141 amino acids) and 91% identity (129/141 amino acids) with an unnamed murine product (Accession #BAA95101), suggesting that this is a real translation product that represents a novel human ovary-specific antigen.

The novelty of O1034C/O591S was confirmed by Northern Blot analysis using single stranded probes that complement either GPR39 or O1034C/O591S. The strand-specific O1034C/O591S probe specifically hybridized to the ovary tumor samples probed on the Northern blot, whilst all samples were negative when probed with GPR39. In addition real-time PCR was performed using primers specific for either

GPR39 or O1034C/O591S. These results further demonstrated the differential expression profiles of the two sequences. This protein is a putative membrane protein as determined from Corixa's Tmpred protein prediction algorithm.

5

## EXAMPLE 6

EXPRESSION ANALYSIS AND FURTHER CHARACTERIZATION OF  
OVARIAN SEQUENCE O568S

The ovarian sequence O568S was originally identified as cDNA clone 24742 (SEQ ID NO:118). Using clone 24742 as a query sequence to search public  
10 sequence databases, the sequence was found to have a high degree of homology with KIAA0762 (SEQ ID NO:177) and with VSGF. The DNA sequence for VSGF is provided in SEQ ID 184 and the VSGF protein sequence is provided in SEQ ID NO:186.

Real-time PCR (*see* Gibson et al., *Genome Research* 6:995-1001, 1996;  
15 Heid et al., *Genome Research* 6:986-994, 1996) is a technique that evaluates the level of PCR product accumulation during amplification. This technique permits quantitative evaluation of mRNA levels in multiple samples. Briefly, mRNA is extracted from tumor and normal tissue and cDNA is prepared using standard techniques. Real-time PCR is performed, for example, using a Perkin Elmer/Applied Biosystems (Foster City,  
20 CA) 7700 Prism instrument. Matching primers and fluorescent probes are designed for genes of interest using, for example, the primer express program provided by Perkin Elmer/Applied Biosystems (Foster City, CA). Optimal concentrations of primers and probes are initially determined by those of ordinary skill in the art, and control (*e.g.*,  $\beta$ -actin) primers and probes are obtained commercially from, for example, Perkin  
25 Elmer/Applied Biosystems (Foster City, CA). To quantitate the amount of specific RNA in a sample, a standard curve is generated using a plasmid containing the gene of interest. Standard curves are generated using the Ct values determined in the real-time PCR, which are related to the initial cDNA concentration used in the assay. Standard dilutions ranging from  $10^{-10}$  to  $10^{-6}$  copies of the gene of interest are generally sufficient. In  
30 addition, a standard curve is generated for the control sequence. This permits

standardization of initial RNA content of a tissue sample to the amount of control for comparison purposes.

By RealTime PCR analysis, O568 was highly overexpressed in the majority of ovary tumors and ovary tumor metastases tested relative to normal ovary tissue and relative to an extensive normal tissue panel. Little or no expression was observed in normal esophagus, spinal cord, bladder, colon, liver, PBMC (activated or resting), lung, skin, small intestine, stomach, skeletal muscle, pancreas, dendritic cells, heart, spleen bone marrow, thyroid, trachea, thymus, bronchia, cerebellum, ureter, uterus and peritoneum epithelium. Some low level expression was observed in normal breast, brain, bone, kidney, adrenal gland and salivary gland, but the expression levels in these normal tissues were generally at least several fold less than the levels observed in ovary tumors overexpressing O568S.

Moreover, a series of Northern blots was performed which also demonstrated that the ORF region of O568S is specifically overexpressed in ovary tumors. The initial blot contained RNA from a series of normal tissues as well as from ovary tumors. This blot was probed using, as a labeled probe, DNA from O568S that corresponded to the 3'UTR of the VSGF sequence disclosed in SEQ ID NO:184. This blot revealed an ovary tumor-specific 5.0Kb message as well as a potential 3.5Kb brain specific message and a ubiquitously expressed 1.35 Kb message.

Another Northern blot was performed with RNAs from a number of different brain tissues and probed with the 3'UTR region as above. Five of eleven brain samples showed overexpression of the 3.5Kb message. In order to determine whether the ORF region of O568S was specifically overexpressed in ovary tumors, a series of three blots was carried out using three separate probes designed from within the VSGF ORF of O568S. Results from these experiments clearly indicated that only the 5.0Kb message is expressed in ovary tumor.

## EXAMPLE 7

### SYNTHESIS OF POLYPEPTIDES

Polypeptides are synthesized on a Perkin Elmer/Applied Biosystems Division 430A peptide synthesizer using FMOC chemistry with HPTU (O-

Benzotriazole-N,N,N',N'-tetramethyluronium hexafluorophosphate) activation. A Gly-Cys-Gly sequence is attached to the amino terminus of the peptide to provide a method of conjugation, binding to an immobilized surface, or labeling of the peptide. Cleavage of the peptides from the solid support is carried out using the following cleavage mixture: trifluoroacetic acid:ethanedithiol:thioanisole:water:phenol (40:1:2:2:3). After cleaving for 2 hours, the peptides are precipitated in cold methyl-t-butyl-ether. The peptide pellets are then dissolved in water containing 0.1% trifluoroacetic acid (TFA) and lyophilized prior to purification by C18 reverse phase HPLC. A gradient of 0%-60% acetonitrile (containing 0.1% TFA) in water (containing 0.1% TFA) is used to elute the peptides. Following lyophilization of the pure fractions, the peptides are characterized using electrospray or other types of mass spectrometry and by amino acid analysis.

#### EXAMPLE 8

15

##### O568S NORTHERN BLOT ANALYSIS

As described in Example 6, Northern blot analysis demonstrated that the ORF region of O568S was specifically over expressed in ovarian tumors. The original probe used corresponded to the 3'UTR of the VSGF sequence disclosed in SEQ ID NO:184. The results from these Northern blots revealed an ovarian tumor-specific 5.0 Kb message as well as a potential 3.5 Kb brain specific message. To confirm that the entire region covered by the ORF yields a single 5.0 Kb ovarian tumor-specific message, two additional probes were designed. The probes were located at the 5' and 3' regions of the ORF. Northern blot analysis using these two probes demonstrated that both probes hybridized to a 5.0 Kb product present only in ovarian tumor samples. Both probes failed to hybridize with RNA derived from multiple brain samples.

#### EXAMPLE 9

##### REAL TIME PCR AND NORTHERN BLOT ANALYSIS OF O590S

Real time PCR analysis of ovarian tumor antigen O590S was performed essentially as described in Example 6. O590S specific primers and probe were designed and quantitative Real Time PCR was performed on a panel of cDNAs prepared from a

variety of tissues including ovarian tumor samples and a panel of normal tissues. This analysis revealed that O590S-specific mRNA was over expressed in approximately 65% of ovarian tumor samples tested, 100% tumor samples derived from SCID mice, and 100% ovarian tumor cell lines tested, when compared to normal ovarian tissue. No  
5 detectable expression was observed in normal tissues.

In addition to Real Time PCR, Northern blot analysis was performed to determine to transcript size of O590S. The Northern blot was probed with a 537 bp PCR product specific for O590S, which was designed to avoid regions of repeat sequences. This probe revealed a smeared band that was approximately 9.0 Kb in size,  
10 which was present in the majority of ovarian tumor samples tested.

From the foregoing it will be appreciated that, although specific embodiments of the invention have been described herein for purposes of illustration, various modifications may be made without deviating from the spirit and scope of the invention. Accordingly, the invention is not limited except as by the appended claims.

## CLAIMS

## What is Claimed:

1. An isolated polypeptide comprising an amino acid sequence of an ovarian tumor protein selected from the group consisting of:
  - (a) sequences provided in SEQ ID NOs: 215, 200-202, 207, and 209;
  - (b) sequences having at least 70% identity to a sequence provided in SEQ ID NOs: 215, 200-202, 207, and 209; and
  - (c) sequences having at least 90% identity to a sequence provided in SEQ ID NOs: 215, 200-202, 207, and 209.
2. An isolated polynucleotide comprising a sequence selected from the group consisting of:
  - (a) sequences provided in SEQ ID NO: 214, 203-206, 208, and 210-213;
  - (b) complements of the sequences provided in SEQ ID NO: 214, 203-206, 208, and 210-213;
  - (c) sequences consisting of at least 20 contiguous residues of a sequence provided in SEQ ID NO: 214, 203-206, 208, and 210-213;
  - (d) sequences that hybridize to a sequence provided in SEQ ID NO: 214, 203-206, 208, and 210-213 under moderately stringent conditions;
  - (e) sequences having at least 75% identity to a sequence provided in SEQ ID NO: 214, 203-206, 208, and 210-213;
  - (f) sequences having at least 90% identity to a sequence provided in SEQ ID NO: 214, 203-206, 208, and 210-213 and
  - (g) degenerate variants of a sequence provided in SEQ ID NO: 214, 203-206, 208, and 210-213.

3. An isolated polypeptide comprising an amino acid sequence of an ovarian tumor protein selected from the group consisting of:

- (a) sequences encoded by a polynucleotide of claim 2;
- (b) sequences having at least 70% identity to a sequence encoded by a polynucleotide of claim 2; and
- (c) sequences having at least 90% identity to a sequence encoded by a polynucleotide of claim 2.

4. An expression vector comprising a polynucleotide of claim 2 operably linked to an expression control sequence.

5. A host cell transformed or transfected with an expression vector according to claim 4.

6. An isolated antibody, or antigen-binding fragment thereof, that specifically binds to a polypeptide of claim 1 or claim 3.

7. A method for detecting the presence of an ovarian cancer in a patient, comprising the steps of:

- (a) obtaining a biological sample from the patient;
- (b) contacting the biological sample with a binding agent that binds to a polypeptide of claim 1 or claim 3;
- (c) detecting in the sample an amount of polypeptide that binds to the binding agent; and
- (d) comparing the amount of polypeptide to a predetermined cut-off value and therefrom determining the presence of a cancer in the patient.

8. A fusion protein comprising at least one polypeptide according to claim 1 or claim 3.

9. An oligonucleotide that hybridizes to a sequence recited in SEQ ID NO: 214, 203-206, 208, and 210-213 under moderately stringent conditions.

10. A method for stimulating and/or expanding T cells specific for a tumor protein, comprising contacting T cells with at least one component selected from the group consisting of:

- (a) polypeptides according to claim 1 or claim 3;
- (b) polynucleotides according to claim 2; and
- (c) antigen-presenting cells that express a polypeptide according to claim 1 or claim 3,

under conditions and for a time sufficient to permit the stimulation and/or expansion of T cells.

11. An isolated T cell population, comprising T cells prepared according to the method of claim 10.

12. A composition comprising a first component selected from the group consisting of physiologically acceptable carriers and immunostimulants, and a second component selected from the group consisting of:

- (a) polypeptides according to claim 1 or claim 3;
- (b) polynucleotides according to claim 2;
- (c) antibodies according to claim 6;
- (d) fusion proteins according to claim 8; and
- (e) antigen presenting cells that express a polypeptide according to claim 2 or claim 3.

13. A method for stimulating an immune response in a patient, comprising administering to the patient a composition of claim 12.

14. A method for the treatment of a cancer in a patient, comprising administering to the patient a composition of claim 12.

15. A method for determining the presence of a cancer in a patient, comprising the steps of:

- (a) obtaining a biological sample from the patient;
- (b) contacting the biological sample with an oligonucleotide according to claim 9;
- (c) detecting in the sample an amount of a polynucleotide that hybridizes to the oligonucleotide; and
- (d) compare the amount of polynucleotide that hybridizes to the oligonucleotide to a predetermined cut-off value, and therefrom determining the presence of the cancer in the patient.

16. A diagnostic kit comprising at least one oligonucleotide according to claim 9.

17. A diagnostic kit comprising at least one antibody according to claim 6 and a detection reagent, wherein the detection reagent comprises a reporter group.

18. A method for inhibiting the development of a cancer in a patient, comprising the steps of:

- (a) incubating CD4+ and/or CD8+ T cells isolated from a patient with at least one component selected from the group consisting of: (i) polypeptides according to claim 1 or claim 3; (ii) polynucleotides according to claim 2; and (iii) antigen presenting cells that express a polypeptide of claim 1 or claim 3, such that T cell proliferate;
- (b) administering to the patient an effective amount of the proliferated T cells,  
and thereby inhibiting the development of a cancer in the patient.

## SEQUENCE LISTING

<110> Corixa Corporation  
 Xu, Jiangchun  
 Stolk, John A.  
 Algate, Paul A.  
 Fling, Steven P.  
 Molesh, David Alan

<120> COMPOSITIONS AND METHODS FOR THE THERAPY  
 AND DIAGNOSIS OF OVARIAN CANCER

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&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 13

```

accacaggct ggccacaaga agcgctggag tgtgctggcg gctgcaggcc tacggggcct 60
ggtccggctg ctgcacgtgc gtgccggctt ctgctgcggg gtcacccgag cccacaagaa 120
ggccatcgcc accctgtgct tcagccccgc ccacgagacc catctcttca cggcctccta 180
tgacaagcgg atcatcctct gggacatcgg ggtgcccaac caggactacg aattccaggc 240
cagccagctg ctacactgg acaccacctc tatccccctg cgcctctgcc ctgtcgctc 300
ctgccgggac gccgcctgc tggccggctg cgagggcggc tgctgctgct gggacgtgcg 360
gctggaccag ccccaaaaga ggagggtgtg tgaagt 396

```

&lt;210&gt; 14

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 14

```

acggcgctct cgtggaagtg acatcgtctt taaaccctgc gtggcaatcc ctgacgcacc 60
gccgtgatgc ccagggaaga caggcgacc tggaggtcca actacttcct taagatcatc 120
caactattgg atgattatcc gaaatgttc attgtgggag cagacaatgt gggctccaag 180
cagatgcagc agatccgcat gtcccttcgc gggaaggctg tggctgctgat gggcaagaac 240
accatgatgc gcaaggccat ccgagggcac ctggaaaaca acccagctct ggagaaactg 300
ctgcctcata tccgggggaa tgtgggcttt gtgttcacca aggaggacct cactgagatc 360
agggacatgt tgctggccaa taagtgcca gctgct 396

```

&lt;210&gt; 15

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; 333

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 15

```

accgcgcggg cacagggtgc cgctgaccga ggcgtgcaaa gactccagaa ttggaggcat 60
gatgaagact ctgctgctgt ttgtggggct gctgctgacc tgggagagtg gccaggtcct 120
gggggaccag acggtctcag acaatgagct ccaggaaatg tccaatcagg gaagtaagta 180
cgtcaataag gaaattcaaa atgcttgtca acggggtgaa acagataaag actctcatag 240
aaaaa caaa cgaagagcgc aagacactgc tcagcaacct agaagaagcc aagaagaaga 300
aagaggatgc cctaaatgag accagggaat canagacaaa gctgaaggag ctcccaggag 360
tgtgcaatga gaccatgatg gccctctggg aagagt 396

```

&lt;210&gt; 16

&lt;211&gt; 396

&lt;212&gt; DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 114, 121, 122, 123, 127, 134, 136, 138, 140, 141, 142, 143,  
144, 148, 163, 166, 172, 173, 174, 176, 177, 183, 184, 185,  
187, 195, 196, 198, 199, 202, 203, 206, 213, 214, 215, 216,  
217, 218, 219, 223, 225, 226, 227, 229, 230, 236, 238

<223> n = A,T,C or G

<221> misc\_feature

<222> 239, 252, 256, 257, 261, 262, 268, 269, 273, 278, 280, 288,  
289, 290, 292, 293, 303, 312, 325, 327, 333, 335, 336, 341,  
342, 347, 354, 359, 365, 371, 383, 384, 386, 388, 391

<223> n = A,T,C or G

<400> 16

```
tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt 60
tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt tttngggggg 120
nnnaaaanttt ttntnanan nnnngggnaa aaaaaaaaaa aanaangggg gnnntnnggc 180
ccnnnanaaa aaaanngnna annaancccc ccnnnnnnnc ccncnnntnn ggaaananna 240
aaaccccccc cngggngggg nnaaaaannc ccnggggnan tttttatnnn annccccccc 300
ccnggggggg gnggaaaaaa aaaantnccc ccnannaaaa nngggggnccc ccctttttnc 360
aaaanggggg nccgggcccc ccnnantntt ngggggg 396
```

<210> 17

<211> 396

<212> DNA

<213> Homo sapiens

<400> 17

```
accacactaa ccatatacca atgatggcgc gatgtaacac gagaaagcac ataccaaggc 60
caccacacac cacctgtcca aaaaggcctt cgatacggga taatcctatt tattacctca 120
gaagtttttt tcttcgcagg atttttctga gccttttacc actccagcct agcccctacc 180
ccccaactag gagggcactg gccccaaca ggcatacccc cgctaaatcc cctagaagtc 240
ccactcctaa acacatccgt attactcgca tcaggagtat caatcacctg agctcaccat 300
agtctaatag aaaacaaccg aaaccaaata attcaagcac tgcttattac aattttactg 360
ggtctctatt ttaccctcct acaagcctca gaggtag 396
```

<210> 18

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 51, 54, 66, 81, 86, 98, 106, 111, 117, 124, 129, 133, 135,  
150, 151, 154, 159, 161, 172, 179, 181, 183, 185, 220, 223,  
229, 238, 258, 259, 264, 282, 289, 292, 294, 299, 303, 311,  
315, 329, 343, 349, 351, 353, 361, 369, 370, 389, 392

<223> n = A,T,C or G

<221> misc\_feature

<222> 396

<223> n = A,T,C or G

<400> 18

```
tttttttttt tttttttttt tttttttttt tttttttttt ttttttttta ntcnaaaggg 60
```

```

gaaggncctt ttttattaaa nttggncatt ttacttttct tttttnaaaa ngctaanaaa 120
aaanttttnt tttncttaaa aaaaaccctn natntcacna ncaaaaaaaaa cnattcccnc 180
ntnctttttg tgataaaaaa aaaggcaatg gaattcaacn tancctaana aaactttnc 240
tgaggaggaaa aaaaatttnt ccgngggaaa cacttggggc tntccaaant gnanccatnc 300
tangaggacc ntctntaaga tttccaaang aaacccttc ctnccaaang nantaccccg 360
ntgcctacnn ccataaaaa aaacctcanc cntaan 396

```

&lt;210&gt; 19

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

```

<222> 47, 69, 75, 80, 83, 87, 88, 90, 92, 102, 104, 108, 116, 121,
130, 138, 139, 142, 153, 156, 158, 162, 165, 166, 180, 192,
193, 195, 201, 224, 226, 232, 235, 237, 241, 248, 251, 253,
256, 269, 272, 274, 277, 284, 287, 290, 292, 297

```

&lt;223&gt; n = A,T,C or G

&lt;221&gt; misc\_feature

```

<222> 299, 305, 306, 315, 323, 324, 326, 332, 351, 368, 377, 380,
383, 387, 392

```

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 19

```

tttttttttt tttttttttt tttttttttt tttttttttt tttttntgg tctgggcttt 60
tatttttacna aaaaactaan gnaaaanntn cnttaacta antngaanac aaagtnttaa 120
ngaaaaagggn ctgggggnnt cntttacaaa aanggnccng gncanntttg ggcttaaaan 180
ttcaaaaagg gnnctcaaaa ngggtttgca ttgcatgtt tcancnctaa ancgngangaa 240
naaaaccngg ngncnctgg gaaaagtnt tnanctncca aaanatnaaa tntttgnanc 300
aggggnntttt tgggnaaaaa aannanttcc anaaactttc catccctgg ntttgggttc 360
ggccttgngt tttcggnatn atntccntta angggg 396

```

&lt;210&gt; 20

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

```

<222> 29, 43, 49, 53, 55, 75, 81, 100, 110, 111, 125, 129, 160,
162, 168, 246, 277

```

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 20

```

tttttttttt tttttttttt tttttctna aaaaaccctg ttnttggng gngnggggta 60
taatactaag ttganatgat ntcatttacg ggggaaggcn ctttgtgaan naggccttat 120
ttctnttgnc ctttcgtaca gggaggaatt tgaagttaa anaaaccnac ctggattact 180
ccggtctgaa ctcaaatcac gtaggacttt aatcgttgaa caaacaaacc tttaatagcg 240
gctgcncat tgggatgtcc tgatccaaca tcgaggncgt aaaccctatt gttgatatgg 300
actctaaaaa taggattgag ctgttatccc tagggtaact tgttcccggt gtcaaagtta 360
ttgatcaat tgagtataag tagttcgctt tgactg 396

```

&lt;210&gt; 21

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

<222> 6, 9, 18, 23, 37, 43, 48, 55, 65, 73, 75, 103, 110, 117,  
123, 125, 134, 153, 182, 195, 202, 205, 213, 216, 223, 239,  
249, 276, 293, 294, 302, 307, 344, 356, 359, 369, 374, 381,  
392

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 21

```
acatanatnt tatactancca ttnaccatct cacttgnagg aanactanta tatecnctcac 60
acctnatatc ctncntacta tgcctagaag gaataatact atngctgttn attatancta 120
ctntnataac cctnaacacc cactccctct tanccaatat tgtgcctatt gccatactag 180
tntttgccgc ctgcnaagca gnggngggcc tanccntact agnctcaatc tccaacacnt 240
atggcctana ctacgtacat aacctaacc tactcnaatg ctaaaaactaa tcnncccaac 300
anttatntta ctaccactga catgactttc caaaaaaacac atantttgaa tcaacncanc 360
caccacanc ctanttatta ncatcatccc cntact 396
```

&lt;210&gt; 22

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; 17, 244

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 22

```
tttttttttt ttttganaaa agccggcata aagcactttt attgcaataa taaaacttga 60
gactcataaa tgggtgctggg ggaagggtgc agcaacgatt tctcaccaa tcactacaca 120
ggacagcaaa ggggtgagaa ggggctgagg gaggaaaagc caggaaactg agatcagcag 180
agggagccaa gcatcaaaaa acaggagatg ctgaagctgc gatgaccagc atcattttct 240
taanagaaca ttcaaggatt tgtcatgatg gctgggcttt cactgggtgt taagtctaca 300
aacagcacct tcaattgaaa ctgtcaatta aagtctctaa gatttaggaa gtggtggagc 360
ttggaaagtt atgagattac aaaattcctg aaagtc 396
```

&lt;210&gt; 23

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 23

```
acaaaggcgg ttccaagcta aggaattcca tcagtgtttt tttcgcagcc accaaattta 60
gcaggcctgt gaggttttca tatcctgaag agatgtatnt taaagctttt tttttttaat 120
gaaaaaatgt cagacacaca caaaagtaga atagtaacct ggagtcccca cgtacccagc 180
ctgcagcttc aacagttacc acatttgcca accggagaga ctgccaaggc aggaaaaagc 240
cctggaaaagc ccacggcccc tttttccctt gggtcagagg ccttagagct ggctgcaaaa 300
gcagccaacc aaaggggcag ctgagctcct tcgtggcacc agcagtgttc ctgatgcagt 360
tgaagagttg atgtctttga caacatacgg acactg 396
```

&lt;210&gt; 24

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

<222> 313, 337, 340, 350, 351, 352, 353, 354, 355, 356, 366, 376, 377, 378, 382, 384, 385, 387, 389, 390, 392, 393, 396

<223> n = A,T,C or G

<400> 24

```
cgactatcct ctcagattct tatctggcac taatttataa ctattatatt atcagagact 60
atgtagcaat atatcagtgac acaggcgcat cccaggcctg tacagatgta tgtctacacg 120
taagtataaa tgaatttgca taccagggtt tacacttgca tctctaatag agattaaaaa 180
caacaaattg gcctcttcct aagtatatta atatcattta tccttacatt ttatgcctcc 240
ccctaaatta atgactgagt tgggtgaaag cggctagggt ttattcatac tgtttttgt 300
tctcaacttc aanagtaatc tacctctgaa aaatttntan tttaatattn nnnnnnagga 360
atttgnacca ctttannnct tncnntntnn tnnccn 396
```

<210> 25

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 90, 125, 136, 278, 299, 301, 305, 344, 347, 353, 355, 356, 357, 359, 360, 361, 365, 369, 378, 380, 381, 382, 383, 384, 385, 386, 391, 392, 393, 395, 396

<223> n = A,T,C or G

<400> 25

```
tttttttttt tttttttttt gtctttttaa aaatataaaa gtgttattat tttaaaacat 60
caagcattac agactgtaaa atcaattaan aactttctgt atatgaggac aaaaatacat 120
ttaanacata tacaanaaga tgctttttcc tgagtagaat gcaaactttt atattaagct 180
tctttgaatt ttcaaaatgt aaaataccaa ggctttttca catcagacaa aaatcaggaa 240
tgttcacctt cacatccaaa aagaaaaaaa aaaaaaanc aattttcaag ttgaagttna 300
ncaanaatga tgtaaaatct gaaaaaagtg gccaaaattt taanttncaa canannngnn 360
ncagnttttna tggatctntn nnnnnncttc nnnntnn 396
```

<210> 26

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 313, 314, 316, 318, 321, 343, 344, 352, 353, 356, 363, 366, 370, 372, 373, 374, 375, 377, 378, 379, 383, 384, 385, 386, 387, 391, 393, 394, 395, 396

<223> n = A,T,C or G

<400> 26

```
gacgctcccc cctccccccg agcgccgctc cggctgcacc gcgctcgctc cgagtttcag 60
gtcgtgctga agctagcgcc gtcgtcgctc cccttcagtc gccatcatga ttatctaccg 120
ggacctcatc agccacgatg agatgttctc cgacatctac aagatccggg agatcgcgga 180
cgggttgtgc ctggaggtgg aggggaagat ggtcagtagg acagaaggta acattgatga 240
ctcgctcatt ggtggaaatg cctccgctga aggccccgag ggcgaaggta cccgaaagca 300
cagtaatcac tgnngncnat nttgtcatga accatcacct gcnnгааааа annttnacaa 360
aanaancctn cnnnnannnc ctnnnnnatt ncnnnn 396
```

<210> 27

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 49, 61, 66, 73, 75, 99, 102, 103, 105, 107, 120, 124, 126,  
129, 138, 139, 141, 147, 155, 157, 162, 165, 175, 187, 191,  
193, 198, 207, 217, 218, 220, 221, 223, 226, 231, 232, 245,  
257, 259, 260, 263, 266, 271, 287, 305, 306, 307, 308

<223> n = A,T,C or G

<221> misc\_feature

<222> 321, 330, 332, 335, 342, 343, 344, 345, 349, 350, 351, 352,  
354, 355, 356, 357, 365, 366, 367, 370, 371, 372, 373, 374,  
375, 376, 377, 378, 379, 380, 381, 382, 383, 386, 387, 388,  
389, 391, 392, 393, 394, 395, 396

<223> n = A,T,C or G

<400> 27

```

tttttttttt tttttttttt tttttttttt tttttttttt tggctaaant ttatgtatac 60
nggttnttca aangnggggg aggggggggg gcatccatnt annncnccca ggtttatggn 120
gggntnttnt actattanna nttttcnctt caaancnaag gnttntcaaa tcatnaaaat 180
tattaanatt ncngctgnta aaaaaangaa tgaaccnncn nanganagga nntttcatgg 240
ggggnatgca tcggggnann ccnaanaacc ncggggccat tcccganagg cccaaaaaat 300
gtttnnnnna aaagggtaaa nttaccccn tnaantttat annnnaaann nnannnnnagc 360
ccaannnttn nnnnnnnnnn nnnccnnnna nnnnnn 396

```

<210> 28

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 278, 283, 298, 309, 326, 331, 338, 351, 355, 356, 357, 358,  
360, 371, 377, 378, 383, 386, 387, 391, 393, 394, 395

<223> n = A,T,C or G

<400> 28

```

cgaccttttt tttttttttt atagatgaaa gagggtttat ttattaatat atgatagcct 60
tggctcaaaa aagacaaatg agggctcaaa aaggaattac agtaacttta aaaaatatat 120
taaacatatc caagatccta aatatattat tctcccaaaa agctagctgc ttccaaactt 180
gatttgatat ttgcatgtt ttccctacgt tgcttggtta atatatgtgc ttctcctttc 240
tgcaatcgac gtctgacagc tgatttttgc tgttttgnca acntgacgtt tcaccttntg 300
tttcaccant tctggaggaa ttgttnaaca ncttacaanca ctgccttgaa naaannnnan 360
gcctcaaaaag ntcttgnnct atnctnnttc nttnnt 396

```

<210> 29

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 329, 334, 361, 386, 390

<223> n = A,T,C or G

<400> 29

```

gacttgctca tttagagttt gcaggaggct ccatactagg ttcagtctga aagaaatctc 60

```

```

ctaattggtgc tatagagagg gaggtaacag aaagactctt ttagggcatt tttctgactc 120
atgaaaagag cacagaaaag gatgtttggc aatttgtctt ttaagtctta accttgctaa 180
tgtgaatact gggaaagtga tttttttctc actcgttttt gttgctccat tgtaaagggc 240
ggaggtcagt cttagtggcc ttgagagttg cttttggcat ttaaattttc taagagaatt 300
aactgtattt cctgtcacct attcactant gcangaaata tacttgctcc aaataagtc 360
ntatgagaag tcactgtcaa tgaaanttgn tttgtt 396

```

```

<210> 30
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 28, 83, 126, 138, 254, 275, 298, 310, 311, 353, 363, 374,
379, 393
<223> n = A,T,C or G

```

```

<400> 30
tttttttttt tttttttttg aaatttanaa acaaatttta ttttaagatct gaaatacaat 60
tcctaaaata tcaacttttc canaaaaaccg tggctacaca ataatgcatt gcctctatca 120
tggtanaacg tgcattanac tcaaatacaa aaaccatgaa acaaatcacc atccttcaac 180
aatttgagca aagatagaat gcctaagaac aacatagatg gacttgacaga ggatgggctg 240
ttttacttca agcnccataa aaaaaaaaaa gagcncaaatt gcattgggtt ttcaggnta 300
tacattaagn ngaacctttg gcactaggaa tcagggcgtt ttgtcacata gcnttaaac 360
atnttaaaaa attntgtant gtcaaaggga tangaa 396

```

```

<210> 31
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 285, 287, 350, 362, 365, 377, 378, 382, 388, 390, 393
<223> n = A,T,C or G

```

```

<400> 31
gacggggccag ggccatctgg aaagggaact cggtttttcc agaacgtggt ggatcatctg 60
tcgggtgtgt ggtgaacacg ttcagttcat cagggcctac gctccgggaa ggggccccca 120
gctgtggctc tgccatgccg ggctgtgttt gcagctgtcc gactctccat ccgccttttag 180
aaaaccagcc acttcttttc ataagcactg acaggggcca gccacagcc acaggtgcga 240
tcagtgcctc acgcaggcaa atgcactgaa acccaggggc acacncncgc agagtgaaca 300
gtgagttccc ccgacagccc acgacagcca ggactgccct cccaccccn ccccgacccc 360
angancacgg cacacanntc ancctctnan ctngct 396

```

```

<210> 32
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 341
<223> n = A,T,C or G

```

```

<400> 32
cgactggcct cataccttgt ctacacagtc cctgcacagg gttcctaacc tgtggtagt 60

```

```
aaagaatgtc acttttctaac aggtctggaa gctccgagtt tatcttggga actcaagagg 120
agaggatcac ccagttcaca ggtatttgag gatacaaac cattgctggg ctgggcttta 180
aaagtcttat ctgaaattcc ttgtgaaaca gagtttcac aaagccaatc caaaaggcct 240
atgtaaaaat aaccattctt gctgcacttt atgcaataa tcaggccaaa tataagacta 300
cagtttattt acaatttggt tttacccaaa atgaggacta nagagaaaaa tgggtgctcca 360
aagcttatca tacattgtc attaagtcct agtctc 396
```

<210> 33  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 121, 122, 124, 125, 126, 128, 130, 131, 132, 133, 134, 136,  
137, 153, 154, 155, 156, 157, 158, 159, 168, 169, 170, 171,  
172, 173, 174, 175, 176, 177, 178, 179, 184, 185, 192, 197,  
199, 200, 202, 204, 205, 208, 209, 210, 211, 214, 215  
<223> n = A,T,C or G

<221> misc\_feature  
<222> 216, 217, 218, 222, 227, 228, 229, 233, 234, 241, 242, 244,  
245, 246, 247, 248, 249, 252, 260, 261, 262, 263, 264, 265,  
270, 272, 273, 274, 275, 279, 282, 284, 288, 290, 291, 292,  
293, 294, 299, 300, 301, 302, 303, 306, 313, 314, 319  
<223> n = A,T,C or G

<221> misc\_feature  
<222> 327, 328, 330, 331, 332, 333, 334, 335, 343, 349, 350, 351,  
352, 355, 360, 369, 370, 371, 375, 379, 387, 388, 390, 391,  
392, 393, 394, 395, 396  
<223> n = A,T,C or G

```
<400> 33
cctttttttt tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt 60
tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt 120
nngnnntntn nnnnannaaa aaaaaaaaaa aannnnnnna aaaaaaannn nnnnnnnnnt 180
tttnnggggg gnttttnann gnannttnnn nttnnnnnna ancccccnnng ggnggggggg 240
nntnnnnnng gnaaaaaaan nnnnnnggggn cnnnnngggnc cncncccnan nnnnaaaann 300
nnnggntttt ttnnttttna aaaaaaannn nnnnnaacaa aanttttttn nnaanttttn 360
gggggaaann ncccntttnt ttttttnnan nnnnnn 396
```

<210> 34  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 8, 60, 72, 123, 128, 155, 172, 198, 207, 246, 305, 325, 348,  
349, 369, 371, 380, 393, 394  
<223> n = A,T,C or G

```
<400> 34
acggaccnag ctggaggagc tgggtgtggg gtgcgttggg ctggtgggga ggcctagttn 60
gggtgcaagt angctgtatt gagcttgtgt tgtgctgaag ggacagccct gggtctaggg 120
ganagagncc ctgagtgtga gacccacctt cccngtccc agccctccc anttccccca 180
gggacggcca ctctctgntc cccgacncaa ccatggctga agaacaaccg caggtcgaat 240
```

```

tggtcntgaa ggctggcagt gatggggcca agattgggaa ctgcccattc tcccacagac 300
tgtnatggt actgtggctc aaggnaagtc ccttcaatgt taccacnnt gacacaaaa 360
ggcggaccna nacagtgcac aagctgtgcc canngg 396

```

```

<210> 35
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<400> 35
tcgacaaaaa tcaaactctg cactcacaag ccctggccga cccccaatgg gttttaccac 60
tccccctcta gaccctgtct tgcaaaatcc tctccctagc cagctagtat tttctgggct 120
aaagactgta caaccagttc ctccatttta tagaagttta ctactccag gggaaatggt 180
gagtccctca acctcccttt caaccagtcc catattcca accagtggta ccatagagca 240
gcaccccccg ccaccctctg agccagttagt gccagcagtg atgatggcca cccatgagcc 300
cagtgtctgac ctggcaccca agaaaaagcc caggaagtca agcatgcctg tgaagattga 360
gaaggaaatt attgataacc ccgatgagtt tgatga 396

```

```

<210> 36
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<400> 36
tcgacgggaa gagcctgcta cgggtggactg tgagactcag tgcactgtcc tcctcccagc 60
gacccacgc tggacccoct gccggaccct ccacccttcg gcccccaagc ttcccagggg 120
cttccttttg actggactgt ccctgtctcat ccattctcct gccaccccca gacctcctca 180
gctccagggt gccacctcct ctgcgcagag tgatgaggtc ccggcttctg ctctccgtgg 240
cccatctgcc cacaattcgg gagaccacgg aggagatgct gcttgggggt cctggcacagg 300
agccccacc ctctcctagc ctggatgact acgtgaggtc tataatctcga ctggcacagc 360
ccacctctgt gctggacaag gccacggccc agggcc 396

```

```

<210> 37
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 376
<223> n = A,T,C or G

```

```

<400> 37
cgacgggtgc agcaactggc catgccacag cacataaaga ttacagtgc aagaaaaaca 60
ttgtttgagg attcctttca acagataatg agcttcagtc cccaagatct gcgaagacgt 120
ttgtgggtga tttttccagg agaagaaggt ttagattatg gaggtgtagc aagagaatgg 180
ttctttcttt tgtcacatga agtgttgaa ccaatgtatt gcctgtttga atatgcaggg 240
aaggataact actgcttgca gataaacccc gcttcttaca tcaatccaga tcacctgaaa 300
tattttcggt ttattggcag atttattgcc atggctctgt tccatgggaa aattcataga 360
cacgggtttt tctttncat tctataagcg tatctt 396

```

```

<210> 38
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<400> 38
cgacaaaaat gataaatagc tttaagaatg tgctaagatg aaatgattac atgtcaattt 60

```

```

aatgtactta atgtttaata ccttatttga ataattacct gaagaatata ttttttagta 120
ctgcatttca ttgattctaa gttgcacttt ttaccccat actgttaaca tatctgaaat 180
cagaatgtgt cttacaatca gtgatcgttt aacattgtga caaagtttaa tggacagttt 240
tttcccatat gtatatataa aataatgtgt tttacaatca gtggcttaga ttcagtgaat 300
tacagtaatt cattcaatta tgatagtatc tttacagaca ttttaaaaat aagttatttt 360
tatatgctaa tattctatgt tcaagtggaa tttgga 396

```

```

<210> 39
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<400> 39
tcgaccaaga atagatgctg actgtactcc tcccaggcgc cccttccccc tccaatccca 60
ccaaccctca gagccacccc taaagagata ctttgatatt ttcaacgcag ccctgctttg 120
ggctgccttg gtgctgccac acttcaggct cttctccttt cacaaccttc tgtggctcac 180
agaacccttg gagccaatgg agactgtctc aagagggcac tgggtggccg acagcctggc 240
acagggcaag tgggacaggg catggccagg tggccactcc agaccctgg cttttcactg 300
ctggctgcct tagaaccttt cttacattag cagtttgctt tgtatgcact ttgttttttt 360
ctttgggtct tgtttttttt ttccacttag aaattg 396

```

```

<210> 40
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 200, 375
<223> n = A,T,C or G

```

```

<400> 40
tttttttttt ttttgttatt tagtttttat ttcataatca taaacttaac tctgcaatcc 60
agctaggcat gggagggaac aaggaaaaca tggaacccaa agggaactgc agcgagagca 120
caaagattct aggatactgc gagcaaatgg ggtggagggg tgctctcctg agctacagaa 180
ggaatgatct ggtgggtaan ataaaacaca agtcaaactt attcgagttg tccacagtca 240
gcaatggtga tcttcttgct ggtcttgcca ttctggacc caaagcgctc catggcctcc 300
acaatattca tgccttcttt cactttgcca aacaccacat gcttgccatc caaccactca 360
gtcttggcag tgcanatgaa aaactgggaa ccattt 396

```

```

<210> 41
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 288
<223> n = A,T,C or G

```

```

<400> 41
tcgacctctt gtgtagtcac ttctgattct gacaatcaat caatcaatgg cctagagcac 60
tgactgttaa cacaaacgtc actagcaaag tagcaacagc tttaagtcta aatacaaagc 120
tgttctgtgt gagaattttt taaaaggcta cttgtataat aacccttgct atttttaatg 180
tacaaaacgc tattaagtgg cttagaattt gaacatttgt ggtctttatt tactttgctt 240
cgtgtgtggg caaagcaaca tcttccttaa atatataatta cccaaagnaa aagcaagaag 300
ccagattagg tttttgacaa aacaaacagg ccaaaagggg gctgacctgg agcagagcat 360
ggtgagaggc aaggcatgag agggcaagtt tgttgt 396

```

<210> 42  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> 65, 68, 69, 71, 72, 75, 77, 79, 82, 85, 86, 87, 89, 90, 97,  
 98, 105, 107, 109, 112, 117, 121, 122, 124, 126, 149, 152,  
 153, 155, 157, 161, 163, 167, 168, 169, 174, 177, 178, 179,  
 180, 186, 188, 192, 201, 202, 207, 208, 215, 217, 220  
 <223> n = A,T,C or G

<221> misc\_feature  
 <222> 225, 230, 242, 243, 247, 250, 259, 263, 271, 272, 279, 284,  
 295, 298, 299, 308, 309, 312, 323, 342, 348, 351, 363, 366,  
 370, 386, 390, 392  
 <223> n = A,T,C or G

<400> 42  
 cttttttttt tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt 60  
 aaaanccnna nnaanang gnaannnann aaaaaannca aaccncntnt anaaaangcc 120  
 nntntnaggg ggggggttca aaaccaaang gnnngntngga ngnaaannna aaanttnnnn 180  
 ggggggnanaa anaaaaaggg nngaaanntg acccnanaan gaccngaaan cccgggaaac 240  
 cnngggntan aaaaaaagnt ganccctaaa ncccccgna aaanggggga agggnaannc 300  
 caaatccnnt gnggggttggg gngggggaaa aaaaaaaccc cnaaaaantg naaaaaaccg 360  
 ggnttnaaan atttgggttc gggggnnttn tnttaa 396

<210> 43  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> 108, 195, 213, 279, 287, 349  
 <223> n = A,T,C or G

<400> 43  
 tttttttttt ttttgcttca ctgctttatt tttgaaatca caagcaattc aaagtgatca 60  
 tcattgaggc ttctgttaaa agttcttcca aagttgccca gttttaanat taaacaatat 120  
 tgcactttaa gatgaactaa cttttgggat tctcttcaaa gaaggaaagt attgctccat 180  
 ctgtgctttt cttnactaa aagcatactg canaaaactc tattttaaaa atcaacactg 240  
 cagggtacag taacatagta aagtacctgc ctattttana atcctanaga acatttcatt 300  
 gtaagaaact agccattat ttaagtgtcc acagtatttt tcatttcant ggtccaagat 360  
 gccagggtt ccaaacacaa tcttggttct taatac 396

<210> 44  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<400> 44  
 gacctagttt tacctcttaa atatctctgt tcccttctaa gttgtttgct gtgttttctt 60  
 cagagcaaga aggttatatt ttttaaaatt tacttagtaa tgcacattca aaacacacat 120  
 caagtcttca ggataaagtt caaaaccgct gtcattggccc catgtgatct ctccctcccc 180  
 taccctctta tcatttagtt tcttctgcgc aagccactct ggcttccttt cagttttgtg 240

```
gttccccgttt ttagctagtt cagtgggttt caatgggcat ttcttgccctt tttttttcta 300
aacgacaaat agaaatacat cttctttatt atcctccaaa tccaattcag aggtaatatg 360
ctccacctac acacaatttt agaaataaat taaaaa 396
```

```
<210> 45
<211> 396
<212> DNA
<213> Homo sapiens
```

```
<220>
<221> misc_feature
<222> 18, 19, 22, 39, 40, 43, 62, 84, 90, 99, 103, 104, 105, 117,
120, 123, 128, 134, 139, 141, 142, 143, 144, 145, 182, 187,
207, 218, 219, 242, 247, 257, 260, 263, 272, 276, 277, 279,
284, 288, 294, 296, 297, 305, 310, 314, 319, 320, 322
<223> n = A,T,C or G
```

```
<221> misc_feature
<222> 364, 366, 376, 378, 381, 387, 388, 396
<223> n = A,T,C or G
```

```
<400> 45
tttttttttt ttttaaannt tntaaatttt taatgaaann ganttagaac aatgtattat 60
tnacatgtaa ataaaaaaag agancataaa ccccatatnc tcnnnaaagg aaggganacn 120
gcnggccntt tatnagaana nnnnncatat aagaccccat taagaagaat ctggatctaa 180
anacttncaa acaggagttc acagtangtg aacagcannc cctaatacca ctgatgtgat 240
gnttcanaaa aaatcancan cgntgatcgg gnacnnanc aatntgancg gaanannact 300
gctcnatatn ttnaggann cngatgtggt catTTTTTtac aaagataatg gccacaccct 360
tccngnccga atcgancnga nctcccnntt ctgtgn 396
```

```
<210> 46
<211> 396
<212> DNA
<213> Homo sapiens
```

```
<220>
<221> misc_feature
<222> 24, 105, 144, 188, 190, 214, 317, 369, 371, 378
<223> n = A,T,C or G
```

```
<400> 46
tttttttttt tttttttttc tganacagag tctcattctg ttgcctaggc tggattgcag 60
tggtgccatc tcggctcact gcaacctccg cctcctgggt tccanaaatt ctctgcctc 120
agcctcccgg gtagctggga ctanaggcac acgccaccac gccaggctaa tttttatatt 180
ttaggtanan atggcggttt accatgttga ccanactgat ctggaactcc cgacctcgtg 240
atccaccacac ctccggcctcc caaagtgcgt ggattacagg cgtgaaacca ccaggccccg 300
cctgaaatat ctatttnttt tcagattatt tttaaaattc catttgatga atcttttaaa 360
gtgagctana naaagtgngt gtgtacatgc acacac 396
```

```
<210> 47
<211> 396
<212> DNA
<213> Homo sapiens
```

```
<220>
<221> misc_feature
<222> 290
<223> n = A,T,C or G
```

```

<400> 47
tttttttttt tttttttgct gttgccaact gtttattcag ggccctgaac ggggtggtgcg 60
tgacatgca acacactcgg gccacagca gcgtgaccgg ccgctcccaa gccccgggcg 120
cacaaccaca gccaggagca gccctgcca ccactgggcc accgtccagg gccccacagg 180
accagccgaa ggtgccccgg gccgaggcca gctgggtcag gtgtaccct agcctgggt 240
tgagtgagga gcggcaccac cagtatcctg tgtaccccaa gttgccagn aggccgagg 300
ggccttgggc tccatctgca ctggccaccc cgtgccaaagc atcacagctg cgtgagcagg 360
tttgtgtgtg agcgtgtggc ggggcctggt tgtccc 396

```

```

<210> 48
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 393, 396
<223> n = A,T,C or G

```

```

<400> 48
ctgggcctgt gccgaagggt ctgggcagat cttccaaaga tgtacaaaat gtagaaattg 60
ccctcaagca aatgcaaaga tgctcaacac ccttagtcat caagaaaatg caaatggaat 120
ccacagagag atactgcaca ctgacaaaga tggctgtatt actaaagggtg aataaccagc 180
gcgggggggca cgtggagtca ctggaacatt tgtgcaatgc tgggtgggaat gtcaacccgt 240
gcggccctct ggaataagcc tggcagctcc tccaagagtt acccgtgtga cccagcaatt 300
ccactcctag ctccaccac aggaattgaa agcaaagacg caaacagatg cctgtgcacc 360
aaagttcacg gcagcatcct tcgccatagt gnaaan 396

```

```

<210> 49
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 32, 40, 44, 64, 70, 83, 87, 92, 104, 115, 118, 125, 127,
130, 137, 155, 168, 171, 173, 175, 192, 201, 206, 208, 218,
219, 235, 247, 249, 256, 259, 260, 269, 297, 306, 310, 320,
321, 328, 331, 345, 356, 381, 389, 395
<223> n = A,T,C or G

```

```

<400> 49
accccaaaat gggaaaggaa aagactcata tnaacattgn cgtnattgga cacgtacatt 60
cggncagtn caccactact ggncatntga tntataaatg cggnggcacg gacanaanaa 120
ccatngnaan atttganaag gaggtgctg atatnggaaa gggctccntc nantntgcct 180
gggtcttgga tnaactgaaa nctgancntg aacgtggnt caccattgat atctncttgt 240
ggaaatntna gaccancann tactatgtna ctatcattga tgccccagga cacaganact 300
ttatcnaaan catgattacn nggacatnta nagctgactg tgctngcctg attgtngctg 360
ctggtgttg tgaatttgaa nctggtatnt ccaana 396

```

```

<210> 50
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<400> 50
cgacttcttg ctggtgggtg gggcagtttg gtttagtggt atactttggt ctaagtattt 60

```

```

gagttaaact gcttttttgc taatgagtgg gctggttggt agcaggtttg ttttccctgc 120
tgttgattgt tactagtggc attaactttt agaatttggg ctggtgagat taattttttt 180
taatatccca gctagagata tggcctttaa ctgacctaaa gaggtgtggt gtgatttaat 240
ttttcccggt tcctttttct tcagtaaacc caacaatagt ctaaccttaa aaattgagtt 300
gatgtcctta taggtcacta cccctaaata aacctgaagc aggtgttttc tcttgacat 360
actaaaaaat acctaaaagg aagcttagat gggctg 396

```

```

<210> 51
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 18, 52, 59, 148, 267, 321, 332
<223> n = A,T,C or G

```

```

<400> 51
tttttttttt ttcagcgngg atttatttta ttctattttt tactctcaag anaaagaana 60
gttactattg caggaacaga cattttttta aaaagcgaaa ctctgacac ccttaaaaca 120
gaaaacattg ttattcacat aataatgngg ggctctgtct ctgccgacag gggctgggtt 180
cgggcattag ctgtgccgtc gacaatagcc ccattcaccc cattcataaa tgctgtgct 240
acaggaaggg aacagcggct ctccanaga gggatccacc ctggaacacg agtcacctcc 300
aaagagctgc gactgtttga naatctgcc aagggaana cactcaatgg gacctggata 360
accaggccc gggagtcata gcaggatgtg gtactt 396

```

```

<210> 52
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 81, 189
<223> n = A,T,C or G

```

```

<400> 52
acctcgctaa gtgttcgcta cgcggggcta ccgatcggt cggaaatggc agaggtggag 60
gagacactga agcgactgca nagccagaag ggagtgcagg gaatcatcgt cgtgaacaca 120
gaaggcattc ccatcaagag caccatggac aacccccacca ccaccagta tgccagcctc 180
atgcacagnt tcatcctgaa ggcacggagc accgtgcgtg acatcgacc ccagaacgat 240
ctcaccttcc ttcgaattcg ctccaagaaa aatgaaatta tggttgcacc agataaagac 300
tatttcctga ttgtgattca gaatccaacc gaataagcca ctctcttggc tccctgtgtc 360
attccttaat ttaatgcccc ccaagaatgt taatgt 396

```

```

<210> 53
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 224, 225, 228, 235, 240, 246, 257, 266, 274, 279, 281, 282,
283, 285, 287, 288, 290, 291, 292, 293, 294, 295, 296, 297,
300, 301, 303, 307, 311, 313, 314, 317, 318, 319, 320, 321,
323, 324, 328, 329, 330, 336, 337, 338, 339, 340, 341
<223> n = A,T,C or G

```

<221> misc\_feature

<222> 342, 343, 344, 345, 346, 347, 348, 349, 350, 351, 352, 356,  
357, 358, 359, 362, 363, 364, 365, 366, 367, 373, 380, 381,  
382, 385, 387, 388, 389, 390, 392

<223> n = A,T,C or G

<400> 53

```

tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt 60
tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt 120
tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt tttttttttt 180
tttttttttt tttttttttt tttttttttt tttttttttt ttannttntt tttntttttn 240
ccttnttttt aattcanaaa aagaanaaga aaanataana nnnancnnan nnnnnnnatn 300
ntncttnata ntntttnnnn nannggggnn gcgagnnnnn nnnnnnnnnn nntctnnntt 360
tnnnnnnctt gcncctcttn nnttngnnnn angcaa 396

```

<210> 54

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 367

<223> n = A,T,C or G

<400> 54

```

ctcttggggc tgctgggact cgctcggtt ggcgactccc ggacgtaggt agtttgttgg 60
gccgggttct gaggccttgc ttctctttac ttttcactc taggccacga tgccgcagta 120
ccagacctgg gaggagttca gccgcgtgc cgagaagctt tacctcgctg accctatgaa 180
ggcacgtgtg gttctcaaat ataggcattc tgatgggaac ttgtgtgtta aagtaacaga 240
tgatttagtt tgtttgggtg ataaaacaga ccaagctcaa gatgtaaaga agattgagaa 300
attccacagt caactaatgc gacttatggg agccaaggaa gcccgcaatg ttaccatgga 360
aactgantga atggtttgaa atgaagactt tgcctg 396

```

<210> 55

<211> 396

<212> DNA

<213> Homo sapiens

<400> 55

```

cgacggtttg ccgccagaac acaggtgtcg tgaaaactac ccctaaaagc caaaatggga 60
aaggaaaaga ctcatatcaa cattgtcgtc attggacacg tagattcggg caagtccacc 120
actactggcc atctgatcta taaatgcggg ggcatcgaca aaagaacat tgaaaaattt 180
gagaaggagg ctgctgagat gggaaagggc tccttcaagt atgcctgggt cttggataaa 240
ctgaaagctg agcgtgaacg tggatcacc attgatattc cctgtggaa atttgagacc 300
agcaagtact atgtgactat cattgatgcc ccaggacaca gagactttat caaaaacatg 360
attacaggga catctcaggc tgactgtgct gtcctg 396

```

<210> 56

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 134, 145, 255, 279, 337, 344, 369

<223> n = A,T,C or G

<400> 56  
 tttttttttt tttttttctca ttttaactttt ttaatgggtc tcaaaattct gtgacaaatt 60  
 tttgggtcaag ttgtttccat taaaaagtac tgatttttaa aactaataac ttaaaactgc 120  
 cacacgcaaa aaanaaaacc aaagnggtcc acaaaacatt ctcttttcct tctgaagggt 180  
 ttacgatgca ttgttatcat taaccagtct ttactacta aacttaaatg gccaatgaa 240  
 acaaacagtt ctganaccgt tcttccacca ctgattaana gtgggggtggc aggtattagg 300  
 gataatatcc atttagcctt ctgagcttcc tgggcanact tggngacctt gccagctcca 360  
 gcagccttnt tgtccactgc tttgatgaca cccacc 396

<210> 57  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> 52, 57, 58, 61, 72, 75, 77, 84, 87, 88, 93, 100, 101, 111,  
 117, 119, 121, 131, 132, 133, 134, 142, 143, 154, 156, 159,  
 167, 168, 170, 175, 176, 182, 183, 185, 186, 190, 192, 194,  
 198, 199, 200, 209, 212, 217, 218, 220, 232, 235, 253  
 <223> n = A,T,C or G

<221> misc\_feature  
 <222> 255, 257, 258, 260, 262, 263, 270, 271, 273, 277, 280, 281,  
 284, 285, 289, 296, 297, 298, 303, 305, 307, 309, 310, 317,  
 322, 324, 337, 338, 342, 344, 346, 347, 349, 351, 356, 358,  
 366, 368, 371, 377, 380, 388, 389, 393, 396  
 <223> n = A,T,C or G

<400> 57  
 cctttttttt tttttttttt tttttttttt tttttttttt tttttttttt tnaaaanntt 60  
 ntttttgcaa anccnancaa aaanggnngg aangaaaaan nggaaaaatt ntttttncnt 120  
 ntttgggaac nnnnagccct tnntttgaaa aaangnggnc ttaaaanngn tgaannaaag 180  
 gnnanncccn gntncttnnn tttaaaaana anggggngn ttttttttaa anaanatttt 240  
 ttttttccct aanancnncn anntgaaacn ngncccnacn nctnncttna aagggnnaa 300  
 atnanangnn aaaaaanccc tnanccccc cccttanntt tncnannana naaagncntt 360  
 ttgggncntg naaaaaanaa cttttttntt gcnttn 396

<210> 58  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<400> 58  
 cgacctcaaa tatgccttat tttgcacaaa agactgccaa ggacatgacc agcagctggc 60  
 tacagcctcg atttatattt ctgtttgtgg tgaactgatt ttttttaaac caaagtttag 120  
 aaagaggttt ttgaaatgcc tatggtttct ttgaatggta aacttgagca tcttttccact 180  
 ttccagtagt cagcaaagag cagtttgaat tttctgtcgt cttcctatca aaatattcag 240  
 agactcgagc acagcaccca gacttcatgc gcccggtggaa tgctcaccac atgttggctg 300  
 aagcggccga ccaactgactt tgtgacttag gcggctgtgt tgcctatgta gagaacacgc 360  
 ttcaccccca ctccccgtac agtgcgcaca ggcttt 396

<210> 59  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<220>

<221> misc\_feature

<222> 25, 45, 116, 178, 198, 211, 225, 235, 253, 266, 281, 324, 367, 377, 389

<223> n = A,T,C or G

<400> 59

```
cttttttttt tttttttttt tcagnggaaa ataactttta ttganacccc accaactgca 60
aaatctgttc ctggcattaa gtccttctt ctttgcaat tcggtctttc ttcagnggtc 120
ccatgaatgc tttcttctcc tccatggtct ggaagcggcc atggccaaac ttggaggngg 180
tgtcaatgaa cttaaagnca atcttctcca nagcccgccg cttcntctgc accancaagg 240
acttgccggag gngagcacc cgcttnttgg ttcccaccac ncagcctttc agcatgacaa 300
agtcattggt cacttcacca tagnggacaa agccacccaa agggttgatg ctctttggca 360
aataggnecat agtcacngga ggcatgtgnc ttgatc 396
```

<210> 60

<211> 396

<212> DNA

<213> Homo sapiens

<400> 60

```
acctcagctc tcggcgacag gccagcttc cttcaaaatg tctactgttc acgaaatcct 60
gtgcaagctc agcttgagg gtgatcactc tacaccccca agtgcatatg ggtctgtcaa 120
agcctatact aactttgatg ctgagcggga tgctttgaac attgaaacag ccatcaagac 180
caaagggtgtg gatgagggtca ccattgtcaa cattttgacc aaccgcagca atgcacagag 240
acaggatatt gccttcgcct accagagaag gacccaaaag gaacttgcat cagcactgaa 300
gtcagcctta tctggccacc tggagacggg gatcttgggc ctattgaaga cacctgctca 360
gtatgacgct tctgagctaa aagcttccat gaaggg 396
```

<210> 61

<211> 396

<212> DNA

<213> Homo sapiens

<400> 61

```
tagcttgctc gggacggtaa ccgggacccg gtgtctgtc ctgtcgctt cgcctcctaa 60
tccctagcca ctatgcgtga gtgcatctcc atccacgttg gccaggctgg tgtccagatt 120
ggcaatgcct gctgggagct ctactgcctg gaacacggca tccagcccga tggccagatg 180
ccaagtgaca agaccattgg gggaggagat gactccttca acaccttctt cagtgaagac 240
ggcgtctggca agcacgtgcc ccgggctgtg tttgtagact tggaaacccac agtcattgat 300
gaagttcgca ctggcaccta ccgccagctc ttccaccctg agcagctcat cacaggcaag 360
gaagatgctg ccaataacta tgcccagggg cactac 396
```

<210> 62

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 261, 269, 313, 333, 346, 354, 359, 390, 394, 395, 396

<223> n = A,T,C or G

<400> 62

```
tcgacgtttc ctaaagaaaa ccactctttg atcatggctc tctctgccag aattgtgtgc 60
actctgtaac atctttgtgg tagtcctgtt ttctaataa cttgtttact gtgctgtgaa 120
agattacaga tttgaacatg tagtgtacgt gctgttgagt tgtgaactgg tgggccgtat 180
gtaacagctg accaacgtga agatactggt acttgatagc ctcttaagga aaatttgctt 240
ccaaatttta agctggaaaag ncactggant aactttaaaa aagaattaca atacatggct 300
```

ttttagaatt tcnttacgta tgtaagatt tnggtacaaa ttgaantgtc tgtntcganc 360  
ctcaaccaat aaaatctcag tttatgaaan aaannn 396

<210> 63  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 3, 11, 16, 18, 23, 26, 30, 34, 37, 50, 51, 60, 61, 62, 63,  
64, 75, 82, 83, 84, 85, 87, 89, 93, 94, 97, 98, 99, 118,  
119, 120, 122, 134, 136, 138, 139, 141, 144, 145, 147, 152,  
156, 187, 188, 193, 195, 204, 211, 214, 216, 222, 226  
<223> n = A,T,C or G

<221> misc\_feature  
<222> 228, 235, 242, 258, 264, 265, 269, 275, 294, 298, 301, 307,  
316, 326, 334, 335, 339, 340, 343, 350, 351, 355, 373, 378,  
390  
<223> n = A,T,C or G

<400> 63  
ttnttttttt ntntntnttt ttntcnttgn ttgnacngaa cccggcgctn nttecccaen 60  
nnnnacggcc gccentattc annnntnctt canntannna ccgcaccctc ggactgcnnn 120  
tngggccccg ccgncnannc nccnnncncc anttncnccg ccgcccgccg gccttttttt 180  
attggcnnc atnanaaccg gggncacctc ncangngcgc cnaaantngg ggcangactc 240  
anagggggcc atcaaccncc aagnncaanc tgganctcta caaacggcct acgntttntg 300  
nccatgnggg tagggnttta cccgcnatga tgannatggn aanaactttt ncaanccctt 360  
tattaaccaa tngggtgngg agacggaacn tggtta 396

<210> 64  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 175, 177, 340, 393  
<223> n = A,T,C or G

<400> 64  
tcgacgtcgg gggtttcctgc ttcaacagtg cttggacgga acccggcgct cgttccccac 60  
cccggccggc cgcccatagc cagccctccg tcacctcttc accgcaccct cgactgccc 120  
caaggccccc gccgcgcgtc cagcgccgcg cagccaccgc cgccgcgcgc gcctntnctt 180  
agtgcgcgcc atgacgaccg cgtccacctc gcaggtgcgc cagaactacc accaggactc 240  
agaggccgcc atcaaccgcc agatcaacct ggagctctac gcctcctacg tttacctgtc 300  
catgtcttac tactttgacc gcgatgatgt ggctttgaa aactttgcca aatactttct 360  
tcccaatctc atgaggagaa ggaacatgct ganaaa 396

<210> 65  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 26, 56, 103, 122, 145, 151, 154, 187, 189, 203, 224, 256,

273, 305, 344

&lt;223&gt; n = A, T, C or G

&lt;400&gt; 65

```

tttttttttt tttttttttt ttttttnacca ataatgcttt tattttccac atcaanatta 60
atztatatgt tagtttttagt acaagtacta aaatgtatac ttnttgccct aatagctaag 120
gnatacataa gcttcacccat acatnttgca nccnccgtgc tgtccctatgt cattgttata 180
aatgtanana ttttaggaaa ctnttttatt caacctggga catntatact gtaggagtta 240
gcaactgacct gatgtnttat ttaaaagtaa tgnatattac ctttacatat attccttata 300
tattnaaacg tatttccatg ttatccagct taaaatcaca tggngggttaa aagcatgagt 360
tctgagtcaa atctggactg aaatcctgat gctccc 396

```

&lt;210&gt; 66

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 66

```

tcgacttttt tttttccagg acattgtcat aatittttat tatgtatcaa attgtcttca 60
atataagtta caacttgatt aaagttagata gacatttgta tctattttaa gacaaaaaaa 120
tctcttttatg tacaatatct tgtctagagt ctagcaaata tagtaccttt cattgcagga 180
tttctgctta atataacaag caaaaacaaa caactgaaa aatataaacc aaagcaaacc 240
aaaccccccg ctcaactaca aatgtcaata ttgaatgaag cattaaaaga caaacataaa 300
gtaacttcag cttttatcta gcaatgcaga atgaatacta aaattagtgg caaaaaaaca 360
aacaacaaac aacaaacaaa acaaaaacaaa caaaca 396

```

&lt;210&gt; 67

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 67

```

acgctttttgt ccttcatttt aactgttatg tcatactgtt atgttgacat atttctttat 60
aagagaatag aggcaaaagt atagaactga ggatcatttg tatttttgag ttggaaatta 120
tgaaacttca ccatattatg atcacacata ttttgaagaa cagactgacc aaagctcacc 180
tgttttttgt gttagggtgct ttggctgaac ttgattccag cccccttttc cctttggtgt 240
tgtgtatgtc tcttcatttc ctctcaaata ttcaactctt gcccctatgtc tccttggcag 300
caggatgctg gcatctgtgt agtcctcata ctgtttactg ataaccacaca aattcatttt 360
catggcagac ctaagctcag accctgcctt gtccctg 396

```

&lt;210&gt; 68

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 68

```

acctgagtc tgtcctttct ctctccccgg acagcatgag cttcaccact cgtccacact 60
tctccaccaa ctaccgggtcc ctgggctctg tccaggcgcc cagctacggc gccggccgg 120
tcagcagcgc ggccagcgtc tatgcaggcg ctgggggctc tggttcccg atctccgtgt 180
cccgtccac cagcttcagg ggcggcatgg ggtccggggg cctggccacc gggatagccg 240
ggggtctggc aggaatggga ggcattccaga acgagaagga gaccatgcaa agcctgaacg 300
accgcctggc ctcttacctg gacagagtga ggaacctgga gaccgagaac cggaggctgg 360
agagcaaaat ccgggagcac ttggagaaga agggac 396

```

&lt;210&gt; 69

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

```

<220>
<221> misc_feature
<222> 1, 4, 6, 8, 9, 11, 18, 19, 36, 53, 60, 64, 79, 84, 92, 94,
97, 105, 114, 120, 123, 127, 129, 134, 137, 138, 139, 142,
143, 147, 149, 151, 152, 156, 158, 167, 170, 172, 180, 182,
184, 187, 188, 189, 194, 197, 201, 209, 212, 218, 219
<223> n = A,T,C or G

<221> misc_feature
<222> 220, 222, 223, 225, 228, 229, 230, 232, 233, 236, 242, 244,
247, 250, 251, 253, 256, 257, 259, 261, 270, 271, 274, 277,
278, 279, 282, 284, 288, 289, 296, 298, 300, 310, 315, 316,
320, 321, 324, 328, 330, 331, 334, 336, 340, 347, 350
<223> n = A,T,C or G

<221> misc_feature
<222> 352, 353, 355, 359, 361, 362, 364, 367, 370, 372, 374, 376,
382, 388, 390, 394, 396
<223> n = A,T,C or G

<400> 69
ntcncngnng ntgtggtntt ttttttaatt tttatntttt cttttttttt ctngctagcn 60
cttincttttt ttggaattnc ggtncctttt tntntcnatt ttttngacaa aaanaacctn 120
ttnttttnana ccanagnnng gnnacacnt nnaatntncc ctttttncgn tngggagctn 180
cncntttnnnc gccnacntca ntcgagacng tnccttttnnn tnnancannn tnngtncgtt 240
gncngcnttn ntncannant ntccctatn nacntgnnt cncncatntt tggacnancn 300
cctagccttn ccatnttttn ntntttntn natnancctn gaaaacntcn gntntttcnc 360
nncntttncn cncncncctt cntatgtncn atgncn 396

<210> 70
<211> 396
<212> DNA
<213> Homo sapiens

<220>
<221> misc_feature
<222> 15, 38, 57, 59, 63, 64, 65, 66, 68, 78, 79, 84, 87, 90, 97,
114, 115, 127, 128, 141, 143, 145, 151, 159, 168, 169, 172,
173, 176, 178, 197, 198, 207, 209, 211, 215, 220, 221, 223,
225, 228, 240, 248, 249, 260, 262, 263, 273, 283, 287
<223> n = A,T,C or G

<221> misc_feature
<222> 294, 304, 314, 334, 339, 340, 348, 362, 367, 376, 382, 384,
386, 395
<223> n = A,T,C or G

<400> 70
tttttttttt tttntttttt tttttttttt tttttttntt tttttttttt tttttntnc 60
aannntnaa cttttaanng gccncngcn ccccaanggg gaccctgctt ttgnnggcta 120
aatgccnaa aactttgggg nantnggtat naaacccnc tttgccnnc annttncngg 180
gggggggggg tttttgnngg ggaacangna naacnttttn ncnanggnat caccaaaaan 240
aaagcccnnc cctttttccn annggggggg gnggggggga aantcanccc ccanattgac 300
cttnatttca aaanggggct tataatcctg ggcntggann cttccctnta cccggggggt 360
gnccacnttt tattanaggg gnangnggat ccccnt 396

<210> 71

```

<211> 396  
<212> DNA  
<213> Homo sapiens

<220>

<221> misc\_feature

<222> 15, 21, 30, 33, 35, 36, 42, 43, 44, 45, 46, 51, 56, 58, 59,  
63, 70, 77, 81, 88, 94, 95, 96, 97, 101, 102, 109, 114,  
118, 119, 120, 124, 131, 132, 133, 134, 135, 141, 142, 143,  
144, 145, 146, 148, 149, 154, 158, 162, 164, 166, 172

<223> n = A,T,C or G

<221> misc\_feature

<222> 177, 179, 181, 184, 185, 213, 216, 218, 219, 222, 223, 224,  
230, 231, 240, 241, 242, 245, 247, 251, 252, 255, 258, 259,  
261, 264, 268, 269, 272, 276, 285, 288, 289, 291, 292, 293,  
297, 299, 300, 307, 312, 315, 316, 317, 325, 329, 334

<223> n = A,T,C or G

<221> misc\_feature

<222> 340, 341, 347, 350, 354, 355, 357, 360, 361, 367, 368, 370,  
371, 376, 377, 378, 387, 393, 394

<223> n = A,T,C or G

<400> 71

```
gcacatagag ggccngttaa ntctagaggn ccngnntaaa cnnnnncatc nacctncnnt 60
gcncctgctn gttgccnccc ntctgtgnct tgcnnnnccc nngagcgtnc cttnacccnn 120
gaangtgccct nnnnnnactga nnnnnncnna taanatgngg anantncgtc gncattntnt 180
natnnggggt gatgctattc tgggggggtgg ggngngnna tnnnatactn nggggacgt 240
nnaatnangag nnaatncnng nttntctnnt gntttntggg gggcnatnng nntctntnn 300
ggactcntcg cncannnate aatancttna ttcngtgtan ngcccgncn tagnnncngcn 360
ngtactnnan ngttgnntc attactnttc gtnngg 396
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<210> 72

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 2, 23, 27, 34, 35, 36, 37, 39, 41, 45, 55, 56, 59, 61, 88,  
92, 96, 97, 98, 101, 103, 104, 106, 108, 111, 114, 115,  
121, 128, 129, 131, 159, 170, 191, 202, 227, 233, 235, 240,  
262, 268, 271, 272, 280, 281, 303, 304, 305, 311, 316, 317

<223> n = A,T,C or G

<221> misc\_feature

<222> 321, 324, 336, 344, 345, 353, 360, 362, 363, 364, 365, 366,  
370, 373, 389, 391, 392, 394, 395

<223> n = A,T,C or G

<400> 72

```
tntttttttt tttctaaaac atnactnttt attnnnnang nttntgaac ctctnngcnt 60
natggtgaga gtttgtctga ttaataanaa tngganntt nannanangc ntgnncgcaa 120
ngatggcnnc nctgtatatc ccaccatccc attacactnt gaaccttttn tttgattaat 180
aaaagggaagg natgcgggga anggggaaag agaattgctt aacattacca tgngnccttn 240
gacaaaacttt ccaatggagg cnggaacnaa nnaccaccan ncaactcccc ttttgtaat 300
ttnnnaactt ncaacnncta nctntttatt ttggcntccc tggnggaaac agnctgtatn 360
```

annnnnaagn ccntgagaac atccctggnt nncnna

396

<210> 73

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 1, 7, 9, 14, 23, 35, 38, 44, 48, 50, 61, 74, 76, 79, 80,  
85, 86, 91, 95, 101, 109, 112, 113, 117, 118, 121, 122,  
127, 129, 132, 137, 141, 146, 214, 234, 243, 251, 266, 296,  
305, 306, 336

<223> n = A,T,C or G

<400> 73

```
ntcaacntng actnctgtga ggnatgggtgc tggngncnta tgcngtgngn ttttggatac 60
naccttatgg acantngcnn tcccnnggaa ngatnataat ncttactgna gnnactnnaa 120
nnttcctnt ntcaaaangtt naaaancatt ggatgtgcca caatgatgac agttttattg 180
ctactcttga gtgctataat gatgaagatc ttanccacca ttatcttaac tgangcacc 240
aanatgggtga nttggggaac atatanagta cacctaagtt cacatgaagt tgtttnttcc 300
caggnnctaa agagcaagcc taactcaagc cattgncaca caggtgagac acctctattt 360
tgtacttctc acttttaagg gattagaaaa tagcca 396
```

<210> 74

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 22, 118

<223> n = A,T,C or G

<400> 74

```
cctttttttt tttttttact gngaatatat actttttatt tagtcatttt tgtttacaat 60
tgaaactctg ggaattcaaa attaacatcc ttgccgtga gcttcttata gacaccanaa 120
aaagtttcaa ccttgtgttc cacattgttc tgctgtgctt tgtccaaatg aacctttatg 180
agccggctgc catctagttt gacgcggatt ctcttgccca caatttcgct tgggaagacc 240
aagtcctcaa ggatggcatc gtgcacagct gtcagagtac ggctcctggg acgcttttgc 300
ttattttttg tacggctttt tcgagttggc ttaggcagaa ttctcctctg agcgataaag 360
acgacatgct tcccactgaa ctttttctcc aattcg 396
```

<210> 75

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 14, 38, 41, 43, 47, 53, 73, 75, 78, 83, 96, 112, 113, 117,  
124, 127, 146, 160, 167, 169, 176, 177, 178, 179, 194, 197,  
198, 209, 210, 220, 222, 226, 227, 231, 238, 241, 244, 258,  
259, 260, 270, 271, 274, 288, 301, 302, 305, 307, 316

<223> n = A,T,C or G

<221> misc\_feature

<222> 319, 328, 339, 344, 347, 354, 359, 364, 367, 369, 370, 371,

373, 374, 381, 384, 387, 388

<223> n = A,T,C or G

<400> 75

```

tttttttttt tttttttttt tttttttttt ttttttttnaa ntntaanggg ganggccct 60
ttttttttaa ctngnccntt ttnccttcct ttttttntaaa ggaaaaaaaa anntttnttt 120
ttccttntaaa aacccttttt cccacnaaca aaaaaaacn ttccccntnc cttttnnnna 180
aaaaaaaggg gctnggnntt tccccttann caaaaaacn tntccnnggg naaaaaantt 240
ntnccggggg gggaaacnnn tgggggtgtt nccnaattt gggggccntc ggaagggggg 300
nncncncct aaagangtnt ttcaaaaana aaacccccnt cctnttntaa aaanaaaaaa 360
aaanaangnn ngnttttttt ntctttnncc ccccaa 396

```

<210> 76

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 87, 94, 102, 108, 138, 139, 143, 144, 145, 146, 151, 152,  
158, 168, 170, 171, 187, 204, 206, 224, 261, 262, 267, 268,  
270, 287, 305, 306, 313, 315, 319, 320, 330, 331, 333, 342,  
344, 348, 349, 356, 358, 360, 362, 368, 374, 376, 381

<223> n = A,T,C or G

<221> misc\_feature

<222> 390

<223> n = A,T,C or G

<400> 76

```

acattcttca gaaatacagt gatgaaaatt cattttgaaa ctcaaatatt ttcatttttg 60
atattctcct gtttttatta aaccagngat tacnccctggc cntccctnta aatgttctag 120
gaaggcatgt ctgttgtnnt tttnnnnaaaa nnaaatnttt tttttttngn naaaccccaa 180
atcccanttt atcaggaagt tagncnaatg aaatggaaat tggntaatgg acaaaagcta 240
gcttgtaaaa aggaccaccc nccacnngn ctttaccccc ttggttngtt gggggaaaaa 300
ccatnnttaa ccntntgggn aaaattgggn ncntaaagtt tncntgggna acagtncntn 360
cngtattnaa ttgncnttat nggaaaatcn gggatt 396

```

<210> 77

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 63, 66, 81, 83, 89, 107, 115, 118, 147, 151, 190, 232, 275,  
288, 294, 304, 323, 332, 369, 392

<223> n = A,T,C or G

<400> 77

```

tttttttttt tttttttttt tttttttttt tatcaacatt tatatgcttt attgaaagtt 60
ganaanggca acagttaaatt ncngggacnc cttacaattg tgtaanaaac atgcncanaa 120
acatatgcat ataactacta tacaggngat ntgcaaaaac ccctactggg aaatccattt 180
cattagttaa aactgagcat ttttcaaagt attcaaccag ctcaattgaa anacttcagt 240
gaacaaggat ttacttcagc gtattcagca gctanatttc aaattacnca aagngagtaa 300
ctgngccaaa ttcttaaaat ttntttaggg gnggtttttg gcatgtacca gtttttatgt 360
aaatctatnt ataaaagtcc acacctcttc anacag 396

```

<210> 78  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 8, 14, 16, 20, 26, 28, 36, 38, 39, 40, 51, 52, 55, 57, 58,  
67, 71, 114, 120, 132, 138, 142, 159, 165, 169, 172, 174,  
175, 183, 187, 195, 197, 198, 200, 202, 206, 209, 243, 259,  
260, 267, 283, 292, 305, 311, 315, 317, 319, 323, 324  
<223> n = A,T,C or G

<221> misc\_feature  
<222> 331, 333, 334, 338, 343, 348, 353, 355, 357, 366, 376, 388  
<223> n = A,T,C or G

<400> 78  
agctggcnaa aggnatgn gctgcnangc gattangnnn ggtaacgtca nnggntnncc 60  
agtgcangac nttgtaaaac gacggccaca tgaattgtaa tacgactcac tatngggcgn 120  
attgggccgt gnaggatngt gntcacactc gaatgtatnc tggcngatnc ananngcttt 180  
atngctnttg acggngnntn anccanctng ggctttaggg ggtatcccct cgcccctgct 240  
tcnttgattt gcacgggcnn ctccganttc cttcataata ccngacgctt cnatccccta 300  
gctcngacct ntcantntnt tcnntgggtt ntnnccgntc acngcttncc cgnangntat 360  
aatctnggct cctttnggga tccattantc ttact 396

<210> 79  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 116, 153, 189, 194, 210, 218, 241, 270, 272, 288, 291, 304,  
324, 325, 329, 333, 334, 338, 340, 342, 366, 372, 377, 384,  
396  
<223> n = A,T,C or G

<400> 79  
caccaaccaa aacctggcgc cgttggcatc gtagagtga cacaacccaa aaacgatacg 60  
ccatctgttc tgccctggct gcctcagccc taccagcact ggtcatgtct aaaggncatc 120  
gtattgagga agttcctgaa ctccctttgg tangttgaag ataaagctga aggctacaag 180  
aagaccaang aagntgtttt gtcctttaan aaacttanac gcctggaatg atatcaaaaa 240  
ngctatgcct ctacgcgaat gagactggan angcaaatg agaaaccntc nccgcatcca 300  
gcgnaggggc cgtgcatctc tatnntgang atnntggman cnttcaaggc cttcagaacc 360  
tccctngaaa tncctnctt taangaacca aactgn 396

<210> 80  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 312, 319, 353, 383  
<223> n = A,T,C or G

<400> 80

```

tgtacatagg catcttattc actgcaccct gtcacacca gcaccccccg ccccgcacat 60
tatttgaaag actgggaatt taatggtag ggacagtaaa tctacttctt tttccagga 120
cgactgtccc ctctaaagt aaagtcaata caagaaaact gtctattttt agcctaaagt 180
aaaggctgtg aagaaaattc attttacatt gggtagacag taaaaaaca gtataataac 240
ttgacatgag cacctttaga tccttcctt catggggctt tgggccaga atgacctttg 300
aggcctgtaa anggattgna atttcctata agctgtatag tggagggtt ggnggggtcat 360
ttgagtaagc cctccaagat acnttcaata cctggg 396

```

```

<210> 81
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 240, 286, 361, 364, 374, 375, 379, 380, 381, 387
<223> n = A,T,C or G

```

```

<400> 81
gcagctgaag ttcagcaggt gctgaatcga ttctcctcgg cccctctcat tccacttcca 60
accctccca ttattccagt actacctcag caatttgtgc cccctacaaa tgtagagac 120
tgtatacgcc ttcgaggtct tccctatgca gccacaattg aggacatcct gcatttcctg 180
ggggagttcg ccacagatat tcgtactcat ggggttcaca tggttttgaa tcaccagggn 240
ccgccatcag gagatgcctt tatccagatg aagtctgcgg acagancatt tatggctgca 300
cagaagtggc ataaaaaaaa catgaaggac agatatgttg aagttttcag tgcagctga 360
nganagaaca ttgnngtann ngggggnact ttaaat 396

```

```

<210> 82
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 220, 251, 297, 301, 309, 349, 395
<223> n = A,T,C or G

```

```

<400> 82
gactcagaaa tgtcagttct atgaagttca aaagatcgag aatgtttgct atcttggttg 60
agcagccgca gccaaagcaag taacttgtaa aatgaggaat gccatcaccc ctcgagtgtc 120
catcccatcat aacttggggg tagagcaca gcgttcccag gaactactca ccttaccatc 180
ttggccggtt catttgcttc caccagttct ggaaagagan ggcctagaag ttcaaaaaaa 240
aagtaggaaa ngtgcttttg gagaaaatca cctgctcctc agaactgggc ttacaanctg 300
ngaagtacnc tatgtgccac ctaatcctca tatatgacct caagagacnc caataagcat 360
atttcacca cggaatgacc agtgcttttg gtaana 396

```

```

<210> 83
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 13, 372, 379, 393
<223> n = A,T,C or G

```

```

<400> 83
tttgatttaa ganatttatt atttttttaa aaaaagcaac ttccagggtt gtcattgtac 60

```

```

aggttttgcc cagtctccta tagcatgcta tagtgataac tgatttttta taacaatgac 120
tcagaggcat tgaagatcca taactatctt ctgaattatc acagaaagaa gaaagttaga 180
agagtttaat gttaagtgtg ttaaaaatca tatttctaatt cttttaattt ggttatctga 240
gtatgataat ataggagagc tcagataaca aggaaaaggc attggggtaa gaacactcct 300
tcccacagga tggcattaac agactttttc tgcataatgct ttatatagtt gccaaactaat 360
tcacctttta cncagcttna ttttttttta ctnggg 396

```

```

<210> 84
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 61, 232, 254, 270, 271, 286, 354, 356, 368, 374, 389, 394
<223> n = A,T,C or G

```

```

<400> 84
tttttacagc aatttttttt tattgatgtt taacctgtat acaaccatac ccattttaag 60
ngtacagaca aatgaatttt gacaaaattca ttcactcatc taatcatcac tataaccatg 120
atacagattt ttatcactcc aaaagtccat cctgtgctct tttcaagtcc atcctectca 180
tctgataccc caagccacca ttgttttgct ttctggaact acagttttgg gnttttagaa 240
tttcatatat ggtngaatac taccatttgn natttggggc tgacgncttt cctccaataa 300
tggatttgag aattatctac attttgcatg gatcctgggt tatttatacc aacnangggg 360
tattatgnaa aatnggacca caatttgngn gcanta 396

```

```

<210> 85
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 293, 305, 306, 317, 347, 357, 372, 377, 386, 391
<223> n = A,T,C or G

```

```

<400> 85
cagtgcacgt gtcctacccc agctctgctc cacagcgccc acctgtctcc gccctcggc 60
ccctcgcccg gctttgcta accgccacga tgatgttctc gggcttcaac gcagactacg 120
aggcgtcatc ctcccgtgc agcagcgctc ccccgccgg ggatagcctc tcttactacc 180
actcaccgcg agactccttc tccagcatgg gctcgcctgc aacgcgcagg acttctgcac 240
ggacctggcc gctccagtgc caacttcatt ccacggcaact gcatctcgac canccggact 300
tgcanngggt gggaanccg cccttgtttc tccgtggccc atctaanacc aaaccntca 360
ccttttcgga gncccnccc ctccgntggg nttact 396

```

```

<210> 86
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 5, 6, 28, 50, 58, 90, 108, 110, 118, 145, 154, 194, 244,
285, 292, 300, 312, 315, 342, 344, 346, 359, 374, 378, 380,
396
<223> n = A,T,C or G

```

```

<400> 86

```

```

ttttnnactg aatgtttaat acatttgnag gaacagaaga aatgcagtan ggattaanat 60
tttataatta gacattaatg taacagatgn ttcatttttc aaagaagntn ccccctntc 120
cctatctttt tttaatcttc cttanagcaa taantagtaa ttactatatt tgtggacaag 180
ctgctccact gtgntggaca gtaattatta aatctttatg tttcacatca ttattacctt 240
ccanaattct accttcattt ccctgcacag gttcactgga ctggntcaca ancaaattgn 300
actccactca antanaagag cccaaagaaa ttagagtaac gncnancct atgaattana 360
gacccaaaga ttttaggnngn tgattagaaa cataan 396

```

<210> 87

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 231, 277, 285, 296, 341, 351, 372, 377, 380

<223> n = A,T,C or G

<400> 87

```

atggaggcgc tggggaagct gaagcagttc gatgcctacc ccaagacttt ggaggacttc 60
cggttcaaga cctgcggggg cgccaccgtg accattgtca gtggccttct catgctgcta 120
ctgttcctgt ccgagctgca gtattacctc accacggagg tgcacctga gctctacgtg 180
gacaagtcgc ggggagataa actgaagatc aacatcgatg tactttttcc ncacatgcct 240
tgtgcctatc tgagtattga tgccatggat gtggccngag aacancagct ggatgnggaa 300
cacaacctgt ttaagccacc actagataaa gatgcatccc ngtgagctca nagctgagcg 360
gcatgagctt gngaaantcn aggtgaccgg gtttga 396

```

<210> 88

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 246, 266, 301, 328, 347, 349, 368, 370, 371, 374, 379, 387, 391

<223> n = A,T,C or G

<400> 88

```

tccagagcag agtcagccag catgaccgag cgccgcgtcc ccttctcgct cctgcggggc 60
cccagctggg accccttccg cgactggtac ccgcatagcc gctcttcgac caggccttcg 120
ggctgccccg gctgccggag gagtggctgc agtggttagg cggcagcagc tggccaggct 180
acgtgcgccc cctgcccccc gccgcacga gagccccgca gtggccgcgc ccgctacagc 240
cgcgcnctc agccggcaac tcacancggg gtcggagat ccgggacact gcggaccgct 300
ngcgcgtgcc ctggatgtca ccactttngc ccggacaact gacggtana caaggatggg 360
gggtgganan nccngtaanc caagaanggg naggac 396

```

<210> 89

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 37, 76, 230, 295, 306, 333, 346, 370, 376, 377, 395

<223> n = A,T,C or G

<400> 89

```

gagagaacag taaacatcca gccttagcat ctctcangag tactgcagat cttcattagc 60
tatattcaca tggagnaatg ctattcaacc tatttctctt atcaaaacta attttgtatt 120
ctttgaccaa tgttcctaaa ttcactctgc ttctctatct caatcttttt cccctttctc 180
atctttcctc cttttttcag tttctaactt tcaactgggtc tttggaatgn tttttctttc 240
atctcttttc ttttacattt tgggggtgtcc cctctctttt cttaccctct ttcnccatcc 300
ttctntttct tttgaattgg ctgcccttta tcntctcatc tgctgncatc ttcatttctc 360
ctccctcctn tttccnntca ttctactctc tcccnt 396

```

```

<210> 90
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 82, 110, 115, 120, 121, 125, 126, 129, 131, 140, 141, 144,
145, 146, 148, 149, 150, 153, 154, 157, 158, 160, 161, 163,
164, 166, 170, 172, 173, 174, 175, 179, 182, 184, 189, 193,
194, 195, 200, 206, 213, 215, 217, 218, 219, 220, 227
<223> n = A,T,C or G

```

```

<221> misc_feature
<222> 228, 231, 233, 236, 241, 247, 248, 249, 250, 254, 259, 262,
269, 273, 274, 275, 280, 281, 282, 286, 287, 289, 293, 294,
301, 302, 304, 309, 311, 318, 319, 324, 325, 330, 331, 333,
334, 336, 337, 341, 342, 343, 344, 349, 352, 353, 358
<223> n = A,T,C or G

```

```

<221> misc_feature
<222> 361, 365, 367, 373, 377, 381, 385, 386, 387, 392
<223> n = A,T,C or G

```

```

<400> 90
ggcgcgcggc gcgccccccc acccccgccc cacgtctcgt cgcgcgcgcg tccgctgggg 60
gcggggagcg gtcggggcgg cngcggtcgg ccggcggcag ggtggtgcgn tttcnttttn 120
natnnccnc nttcttcttn nttnnnnnnn ctntanncn nttncttctn cnnntttnc 180
tntntcttna cnnnttttn taatctctt ctntnnnnn tctctnnat ntntnctta 240
nttcctnnnn tttntctnt ctttctcnc ctntntctn nntcnnnc tcnnatattt 300
nntntttnt nccttctnt ctntntctn ntntnnntt nnnnttctnt tntcatntt 360
nccntntta ctntcanctt ntatnnnct ctttt 396

```

```

<210> 91
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 1, 3, 8, 9, 16, 17, 18, 21, 22, 32, 33, 45, 50, 63, 64, 68,
75, 82, 92, 95, 98, 102, 106, 108, 110, 111, 116, 121, 135,
151, 154, 158, 162, 167, 170, 176, 181, 185, 187, 209, 212,
215, 225, 231, 245, 257, 278, 283, 288, 290, 292, 293
<223> n = A,T,C or G

```

```

<221> misc_feature
<222> 312, 324, 326, 330, 331, 333, 334, 344, 345, 349, 351, 352,
357, 358, 382, 384, 390, 392
<223> n = A,T,C or G

```

<400> 91  
 ntntcctnna tttttnnntc ncttttttt tnnaatTTTT ctttnttttn tttataaaaa 60  
 tcnnacanta aaacngcgga anaggggatt tnttnttngg gngtancncn nggccncaaa 120  
 naaccccaaa aatañcccaa aatgcacagg nccngggnaa angaccnacn tgggtntttt 180  
 ntttntnaac aagggggggtt ttaaagggna tnggnatcaa agggnatataa ntttaaacct 240  
 ttganaaatt ttttaanagg cttgcccccc actttgggcc ccncccnncn gnnnggatcc 300  
 aatttttttt cnttgggggt cccngncccn nannttccgg gttnttggnc nntcctnntt 360  
 tttttttttt tgccttcacc cntnccattn cntttt 396

<210> 92  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> 3, 7, 8, 9, 11, 31, 149, 152, 221, 233, 259, 263, 264, 265,  
 266, 274, 278, 279, 283, 286, 294, 302, 307, 309, 310, 311,  
 314, 316, 320, 343, 351, 363, 372, 377, 386, 393  
 <223> n = A,T,C or G

<400> 92  
 ctnttttnnt ntttttttcc ccatcatcca naaatggggt ttattctcag ccgagggaca 60  
 gcaggactgg taaaaactgt caggccacac ggttgccctgc acagcaccct catgcttggg 120  
 aggggggtggg agggatggcg ggggctggnt gnccacaggc cgggcatgac aaggaggtc 180  
 actggaggtg gcacactttg gagtgggatg tcgggggaca ncttcttgg tanttgggcc 240  
 acaagattcc caaggatanc acnnnnactg attnccannc tanagncaag cggntggcca 300  
 tntgtangnn ntntntatn tgactattta tagattttta tanaacaggg naagggcata 360  
 ccncaaaagg gnccaanttt ttaccnccgg gcncct 396

<210> 93  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> 290, 304, 313, 320, 325, 333, 337, 348, 351  
 <223> n = A,T,C or G

<400> 93  
 gctgccacag atctgttctt ttgtccgttt ttgggatcca caggccctat gtatttgaag 60  
 ggaaatgtgt atggctcaga tcctttttga aacatatcat acagggttgca gtcctgaccc 120  
 aagaacagtt ttaatggacc actatgagcc cagttacata aagaaaaagg agtgctaccc 180  
 atgttctcat ccttcagaag aatcctgcga acggagcttc agtaatatat cgtggcttca 240  
 catgtgagga agctacttaa cactagttag tctcacaatg aaggacctgn aatgaaaaat 300  
 ctgnttctaa ccnagtcctn ttanatttt agngcanatc cagaccancg ncggtgctcg 360  
 agtaattctt tcatgggacc tttggaaaac tttcag 396

<210> 94  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> 115, 204, 205, 243, 266, 276, 316, 319, 355, 357, 364

<223> n = A,T,C or G

<400> 94

```
tgccttaacc agtctctcaa gtgatgagac agtgaagtaa aattgagtgc actaaacgaa 60
taagattctg aggaagtctt atcttctgca gtgagtatgg cccaatgctt tctgnngcta 120
aacagatgta atgggaagaa ataaaagcct acgtgttggt aaatccaaca gcaagggaga 180
tttttgaatc ataataactc atanngtgct atctgtcagt gatgccctca gagctcttgc 240
tgntagctgg cagctgacgc ttctangata gttagnntgg aaatggctct cataataact 300
acacaaggaa agtcanccnc cgggcttatg aggaattgga cttaataaat ttagngngct 360
tccnacctaa aatatatctt ttggaagtaa aattta 396
```

<210> 95

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 11, 16, 31, 36, 42, 49, 53, 56, 57, 60, 67, 70, 84, 89, 91,  
92, 99, 105, 106, 112, 120, 121, 125, 127, 128, 133, 137,  
141, 151, 152, 153, 154, 155, 162, 166, 167, 168, 174, 177,  
179, 186, 188, 194, 195, 199, 203, 205, 213, 217, 221

<223> n = A,T,C or G

<221> misc\_feature

<222> 227, 232, 235, 236, 240, 242, 260, 261, 265, 266, 291, 297,  
318, 325, 330, 339, 348, 351, 352, 354, 356, 362, 364, 372,  
380, 392, 395, 396

<223> n = A,T,C or G

<400> 95

```
cctcccaccc ncttanttca tgagattcga naatgncact tntgtgetnt ttncntnntn 60
tattctnaac atttctttct tggngcggna nnaatcccnt ttttnngggc gncctctccn 120
ncttntnntt tcntggngct ntcccttttc nnhnnaaact tntacnnngt ttanaantnt 180
ttctgnangg ggggnntccna aananttttt ccncctnctt nattccnctc tnaannctcn 240
cnaattgttt ccccccccn ntagnttatt ttttctaaaa aattaactcc nacgganaaa 300
attttcccta aaatttcncc tccanatttn gaaaaaacnc gcccgganct nntntnccga 360
tntnaatttt tnaaaaaaan ttattttcat cngggn 396
```

<210> 96

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 161, 193, 253, 259, 281, 288, 299, 309, 318, 319, 335, 340,  
344, 352, 355, 356, 387, 396

<223> n = A,T,C or G

<400> 96

```
cctgggtacc aaatttcttt atttgaagga atggtacaaa tcaaagaact taagtggatg 60
ttttggacaa cttatagaaa aggtaaagga aaccccaaca tgcattgact gccttggcga 120
ccagggaagt caccacacgg ctatggggaa attagccga ngcttaactt tcattatcac 180
tgcttccaag gngtgcttg gcaaaaaaat attccgcca ccaaatcgga cgctccatct 240
tgcccagttg gtnccgggnc cccaattctt ggatgctttc ncctcttntt ccggaatgng 300
ctcatgaant cccccaanng gggcattttg ccagnggccn tttngccatt cnagnnggcc 360
tgatccattt tttccaatgt aatgccnctt cattgn 396
```

<210> 97  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>

<221> misc\_feature

<222> 13, 15, 16, 19, 23, 31, 38, 39, 41, 45, 68, 94, 95, 100,  
119, 131, 133, 141, 144, 164, 171, 182, 186, 190, 191, 195,  
196, 198, 213, 229, 231, 235, 239, 247, 257, 265, 269, 272,  
278, 279, 286, 289, 291, 306, 309, 310, 312, 317, 320

<223> n = A,T,C or G

<221> misc\_feature

<222> 321, 327, 328, 337, 340, 343, 351, 360, 361, 368, 375, 381,  
385, 386, 387, 388

<223> n = A,T,C or G

<400> 97

```
ctcaccctcc tcntnntnt canaatattg ngaacttnt nctgntcgaa tcaactggcat 60
taaagganca ctagctaatt gcactaaatt tacnnactan ggaaactttt ttataatant 120
gcaaaaacat ntnaaaaaga ntgnagtctg cccatttctg cttinggaaga nctcttcact 180
tntaancccn natgnngncc tttgggtcaa aanctccgcg attattacng ngttncncnc 240
tatttgnctt tcctttntcc ccaangccnc anatttcnna actttncnt naaatgcctt 300
tatttnatnn cntttcnacn ncttaannnt ccctttnaan aangatccct ncttcaaant 360
ntttccngt tectngcatt nccnnnnnat ttctct 396
```

<210> 98  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>

<221> misc\_feature

<222> 130, 202, 285, 296, 299, 308, 314, 321, 322, 336, 373

<223> n = A,T,C or G

<400> 98

```
acaggagcaa tgaagccttt gaagtgccag tctatgaaga ggccgtggtg ggactagaat 60
cccagtgcg ccccaagag ttggaccaac cacccttac agcactgttg tgataccccc 120
agcacctgan gaggaacaac ctaccatcca gaggggccag gaaaagccaa actggaacag 180
aggcgaatgg ctgagagggg tncatggcca agaaggaagc cctggaagaa cttcaatcac 240
cttcggtttc gggaccaccg gcttgtgtcc ctgttctgac tgcanaactt ggcgngtnc 300
cccattanaa cctntgactc nncccttget ataagnctgt tttggcccct gatgatgata 360
gggtttttat gangacactt gggcaccccc ttaatg 396
```

<210> 99  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>

<221> misc\_feature

<222> 1, 4, 13, 15, 26, 31, 43, 46, 48, 52, 54, 55, 60, 62, 68,  
72, 93, 112, 118, 119, 122, 131, 132, 133, 134, 145, 147,  
152, 157, 163, 164, 186, 190, 225, 231, 239, 246, 247, 250,  
255, 262, 285, 314, 316, 319, 325, 332, 339, 343, 345

<223> n = A,T,C or G

<221> misc\_feature

<222> 348, 351, 352, 355, 357, 361, 370, 387

<223> n = A,T,C or G

<400> 99

```

ntnttttttc cgnchnaaagg gcaagngttt ncatctttcc tgnccncnca ananngggtg 60
tntgtgcntt tnttttttcc caaaacccgg gtnggggaca ctttttgagg anccactnnt 120
cntccggggc nnnnttttag aaggngncta anaagcntct tgnnggggga aaaacatctt 180
tttgcncnccn acataccccc aagggggggg ggtgtctggg agganactaa ngacttttnt 240
tttttnnccn caaanaactg anggccccca ttgctcccc cccantcttt aaaaacccc 300
ttcaatttcc ttgncnggna aaaanggttg gnaaaaaang agngngcntc nnttncnttt 360
natggaaggn aaaaggtttt tggttgnaaa accccg                               396

```

<210> 100

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 229, 286, 303, 312, 334, 335, 348, 350, 357, 364, 371, 395

<223> n = A,T,C or G

<400> 100

```

ctaacacggt gaaaccctgt ctctactaaa aatacaaaaa aattagccag gcgtggtggc 60
gggcacctgt agtcccagct gctcaggaag ctgaggcagg agaattggcg gaaccagaa 120
ggcgagctt gcagtgcgt gagatcgtgt cagtgcactc cagcctgggc gacagagcga 180
gactcccgct caaaaaaaaa aaaaaaaga gaaaagaaaa agctgcagng agctgggaat 240
gggccctatc cctccttgg ggatcaatga gaccctttt caaaaanaaaa aaaaaataa 300
tgngattttg gnaacatatg gcactggtgc ttcnngaat tctgtttntn ggcatgnccc 360
cctntgactg nggaaaaatc cagcaggagg ccana                               396

```

<210> 101

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 93, 99, 100, 111, 168, 172, 174, 199, 209, 216, 218, 219,  
227, 242, 243, 269, 272, 297, 300, 301, 308, 315, 317, 323,  
331, 341, 344, 348, 357, 359, 363, 364, 366, 376, 379, 386,  
389, 392

<223> n = A,T,C or G

<400> 101

```

agttataact caacagttca tttatatgct gttcatttaa cagttcattt aaacagttca 60
ttataactgt ttaaaaatat atatgcttat agncaaaann tgttgtggcg nagttgttgc 120
cgcttatagc tgagcattat ttcttaaat cttgaatgtt cttttggngg gntnctaaaa 180
ccgtatatga tccatttttna tgggaaacng aattcntnnc attatcncac cttggaaata 240
cnaaacgtgg gggaaaaaaa tcattcccnc cntccaaaac tatacttctt ttatctngan 300
nttcttgntc ctgcnnggt ttngaata nctgggcaaa nggntttnc aaatccntnt 360
acnntncttt gggaantanc ggcaantcnt cncctt                               396

```

<210> 102

<211> 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; 17, 93, 136, 183, 317

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 102

```

actatacata agaacangct cacatgggag gctggagggtg ggtacccagc tgctgtggaa 60
cgggtatgga caggtcataa acctagagtc agngtcctgt tggcctagcc catttcagca 120
ccctgccact tggagnnggac ccctctactc ttcttagcgc ctaccctcat acctatctcc 180
ctnctcccat ctctacgga ctggcgccaa atggctttcc tgccaatttt gggatcttct 240
ctggctctcc agcctgctta ctctctatt tttaaagggc caaacaatc cttctcttt 300
ctcaaacaca gtaatnggc actgacccta ccacacctca tgaagggggc ttgttgcttt 360
tatttggggc cgatctgggg ggggcaaaat attttg 396

```

&lt;210&gt; 103

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; 91, 174, 176, 188, 201, 214, 254, 277, 299, 325, 349, 355, 365, 372, 390

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 103

```

ttgtgttggg actgctgata ggaagatgtc ttcaggaaat gctaaaattg ggcaccctgc 60
cccaacttca aagccacagc tggatgccca natggtcagg ttaaagatat caacctgctg 120
actacaagg aaaatatggg ggggtcttct tttaccctct tgacttccct ttgngngccc 180
cccgaganca ttgctttccg ngatagggca aaanaaatta aaaaacttaa ctggccagtg 240
aatggggctt ctnggatct cttctggca ttacatnggc aatccctaaa aaacaagang 300
actgggaccc ataacattct tttgnatcaa ccgaagcccc cattgttang atatngggct 360
taaangctga tnaagcatct cgtccgggcn ttttat 396

```

&lt;210&gt; 104

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; 32, 53, 86, 141, 154, 156, 181, 182, 197, 204, 219, 224, 226, 229, 232, 245, 253, 260, 262, 271, 273, 276, 292, 301, 303, 305, 321, 325, 332, 343, 352, 382, 392

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 104

```

aaggaggggc gcgccaagac cttcccactc gngcacactg gggcgccga cangacgcaa 60
cccagtccaa cttggatacc cttggnntta gtctcggac acttctttta tctctccgtc 120
gcaacttgtc aagtctctca nactgtctct ctgngntatc tttttcttc gctgctcttc 180
nncccccgac gtattntca aaangtctgc aattgttgna tacntnganc tncaccactg 240
ttacnaggtc atnaatttcn cntcaactct ntccncttg ttccctgata tntcgccgg 300
ngncnccaat tctgtatttt nctcntcaac gntctcactt ttncctctc cnggccactt 360
tctccccttc cttattccgg cnttgtttgc cnccat 396

```

<210> 105  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 57, 306, 356, 388, 391  
<223> n = A,T,C or G

<400> 105  
tcaatagcca gccagtgttc atttttatcc ttgagctttt agtaaaaact tcctggnttt 60  
atttttagtc attgggtcat acagcactaa agtctgctat ttatggaaac taactttttt 120  
gtttttaatc caggccaaca tgtatgtaaa ttaaattttt agataattga ttatctcttt 180  
gtactacttg agatttgatt atgagatgtg catattgctt tgggaagagc tcgaggaagg 240  
aaataattct ctcttttggg ttgaacctca actagataaa ccctaggaat tgtaactgc 300  
acaagnattt tcattccaca aaacctgagg cagctctttt gccagagcgt tcctgnaccc 360  
ccccaccca cttgccttgg gtctttanaa ngagcc 396

<210> 106  
<211> 396  
<212> DNA  
<213> Homo sapiens

<400> 106  
gctgtgtagc acactgagtg acgcaatcaa tgtttactcg aacagaatgc atttcttcac 60  
tccgaagcca aatgacaaat aaagtccaaa ggcattttct cctgtgctga ccaaccaa 120  
aatatgtata gacacacaca catatgcaca cacacacaca cacaccaca gagagagagc 180  
tgcaagagca tggaattcat gtgtttaaag ataatccttt ccatgtgaag tttaaaatta 240  
ctatatattt gctgatggct agattgagag aataaaagac agtaaccttt ctcttcaaag 300  
ataaaatgaa aagcaattgc tcttttcttc ctaaaaaatg caaaagattt acattgctgc 360  
caaatcattt caactgaaaa gaacagtatt gctttg 396

<210> 107  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 12, 210, 257, 261, 271, 302, 311, 314, 318, 368, 374, 385,  
389, 396  
<223> n = A,T,C or G

<400> 107  
ttcacagaac anggtggttt attatttcaa tagcaaagag ctgaaaaatg tcgggtccca 60  
taaaggagca gaacctgacc cagagcctgc agtacatttc caccacacag gggtgcaggc 120  
tgggccaggc agggccaaag gcagcagaaa tgggagtaag agactgtgcc cactgagaag 180  
ctctgctggg tgtgggcagg tgggcatgan atgatgatga tgtagttaa ggaccagta 240  
ggcaaaacct gtcaggnttg ntgaatgtca nagtggatcc aaaaggctga gggggtcgtc 300  
anaaggcgg nggnccncc cttgccgta tgggccttca aaaagtatgc ttgctcatcc 360  
gttgtttncc ccanggagct gccanggana aggctn 396

<210> 108  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 280, 281, 286, 305, 311, 313, 323, 326, 327, 340, 352, 356,  
363, 369, 378, 388, 392  
<223> n = A,T,C or G

<400> 108  
gcctgctttt gatgatgtct acagaaaatg ctggctgagc tgaacacatt tgcccaattc 60  
caggtgtgca cagaaaaccg agaattattca aaattccaaa tttttttctt aggagcaaga 120  
agaaaatgtg gccctaaagg gggtagttg aggggtaggg ggtagtgagg atcttgattt 180  
ggatctcttt ttatttaaatt gtgaatttca acttttgaca atcaaagaaa agacttttgt 240  
tgaatatagct ttactgcttc tcacgtgttt tggagaaaan natcancct gcaatcactt 300  
tttgnaaactg ncnttgattt tcngcnncca agctatatcn aatatcgtct gngtanaaaa 360  
tgnocctggnc ttttgaanga atacatgngt gntgct 396

<210> 109  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 237, 279, 284, 291, 305, 307, 308, 313, 326, 343, 351, 366,  
376, 392, 394, 395  
<223> n = A,T,C or G

<400> 109  
ggccgtaggc agccatggcg cccagcccgg aatggcatgg tcttgaagcc ccacttccac 60  
aaggactggc agcggcgcggt ggccacgtgg ttcaaccagc cggcccggaa gatccgcaga 120  
cgtaaggccc ggcaagccaa ggcgcgccgc atcgctccgc gcccgcgctc ggggtccatc 180  
cggcccatcg tgcgctgccc acggttcggt accacacgaa gggcgcgccg gcgcggnntc 240  
agcctggagg agctcagggt ggccggattt acaagaagng gccngacatc ngatttcttg 300  
ggatncnnga agnggaacaa gtcacngagt ccttgcagcc acntcagcgg ntgatgacac 360  
cgttcnaact catctnttcc caagaaacct cngnnc 396

<210> 110  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 1, 2, 12, 13, 16, 18, 29, 39, 60, 66, 70, 86, 90, 104, 121,  
122, 127, 128, 146, 165, 171, 172, 173, 176, 188, 189, 193,  
195, 205, 210, 211, 224, 226, 227, 231, 233, 240, 243, 244,  
248, 249, 255, 257, 258, 260, 266, 268, 272, 273, 275  
<223> n = A,T,C or G

<221> misc\_feature  
<222> 278, 280, 287, 292, 294, 303, 308, 312, 315, 320, 322, 332,  
333, 334, 335, 345, 347, 351, 363, 364, 369, 371, 372, 379,  
381, 382, 386, 391, 393  
<223> n = A,T,C or G

<400> 110  
nntgggctcc tnncantnat aataaacng actcatacnc cacaaggaga tgaacaggan 60  
tatgtncatn ctgacgcgga aacagnscan ggagctgagg agngccaag atgagacctt 120  
nnggccnngg tgggcgcatt cccgngggag ggggccacta aggantacga nnntcnagcg 180

gctcttgng gcnncctec tcacnctgn ntattcgatt gtcncnnatg ncntcctatn 240  
atnntcanna ttctntnntn atctcntnta cnnctncn ttcatgntta cngntccctc 300  
tcnttctnac cnttntctgn anctccttcc tnnncttcc atctntnttc ngctttcttt 360  
ctnnaatcnt nntttaacnt nntctncttt ntnatt 396

<210> 111  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 4, 7, 11, 16, 19, 25, 26, 30, 33, 39, 54, 60, 69, 75, 81,  
99, 102, 130, 132, 143, 154, 156, 166, 180, 182, 188, 190,  
192, 194, 198, 201, 226, 242, 253, 261, 264, 295, 305, 313,  
315, 320, 323, 325, 330, 334, 337, 340, 344, 348, 349  
<223> n = A,T,C or G

<221> misc\_feature  
<222> 351, 352, 357, 358, 359, 361, 362, 381, 387, 388, 389, 394  
<223> n = A,T,C or G

<400> 111  
taangancat nctggnttnt gectnnccgn ctnattgant gttaaaggca attntgtggn 60  
tgtcccagng aatgncggct nattttcttt ccacattgng cncattcact cctcccactc 120  
ttggcatgtn gngacataag canggtacat aatngnaaaa atctgnattt ctgatgccan 180  
angggatanan cntnttgnat ntcattccat tgatatacag ccactntttt atttttgatc 240  
ancggccttc ggntcactgc ncanggtact tgacctcagt gtcactatta tgggntttgg 300  
tttctctctt ttncnggcn ttntntttcn cacnttncan cttnttntt nnaaaannna 360  
nnactctctt ctgtctctt ngatacnng tctnaa 396

<210> 112  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 172, 186, 378, 380, 382, 388  
<223> n = A,T,C or G

<400> 112  
tcaacgtcac caattactgc catttagccc acgagctgcg tctcagctgc atggagagga 60  
aaaagggtcca gattcgaagc atggatccct ccgccttggc aagcgaccga tttaacctca 120  
tactggcaga taccaacagt gaccggctct tcacagtga cgaatgttaa gntggaggct 180  
ccaagnatgg tatcatcaac ctgcaaagtc tgaagacccc tacgtcaag gtgttcatgc 240  
acgaaaacct ctacttcacc aaccggaagg tgaattcggg gggctgggcc tcgctgaatc 300  
acttggtatc cacattctgc tatgcctcat gggactcgca gaacttcagg ctggccaccc 360  
tgctcccacc atcactgntn gncaatantc acccag 396

<210> 113  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 1, 2, 3, 4, 7, 8, 9, 10, 11, 65, 273, 279, 280, 289, 321,

338, 380

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 113

```

nnnntttnnn nggagcctta atttcagagt tttattgtat tgcactaaag gaacagcagg 60
atggntatac aattttctct cattcagttt tgaaaaatctg tagtacctgc aaattcttaa 120
gaataccttt accaccagat tagaacagta agcataataa ccaatttctt aataagtaat 180
gtcttacaaa taaaaacaca tttaaaatag ctttaaagtc attcttcaca agtaattcag 240
catatatattt atatcatggg tacttatgct tangaattnn agcaggatnt ttattctttt 300
gatggaaata tgggaaaact ntattcatgc atatacangg ataatttca gcgaagggaa 360
aatcccgttt ttattttggn aatgattcat atataa 396

```

&lt;210&gt; 114

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

<222> 40, 82, 114, 116, 146, 164, 166, 174, 185, 212, 215, 219,  
 224, 236, 242, 254, 258, 263, 270, 286, 299, 308, 327, 328,  
 329, 345, 363, 378, 382, 385

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 114

```

aaatgggaca acgtgattct tttgttttaa ataaataactn agaacacgga cttggctcct 60
acaagcattt ggactctaag gnttagaact ggagagtctt acccatgggc ccncncagg 120
gacgccacgg ttccctccca ccccgngatc aagacacgga atcngntggc gatngttgga 180
tcgcnatgtg ccccttatct atagccttcc cnggncatnt acangcagga tgcgngtggg 240
anaactacaa ctgnaatntc tcnaacggtn atggccccca ccgatnaaga ttctacctng 300
tcttttctnc ccttgaggatg tgagtgnnng aggaagaagc ccttncctta catcaccttt 360
tgnacttctg aacaaganca anacnatggc cccccc 396

```

&lt;210&gt; 115

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; 277, 297, 321, 341, 381, 391

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 115

```

ccgcctgggt cggcccgcct gcctccaact ctgcctctac catgtccatc agggtgaccc 60
agaagtccta caaggtgtcc acctctggcc cccgggcctt cagcagccgc tcctacacga 120
gtgggcccgg ttcccgcatc agctcctcga gcttctcccg agtgggcagc agcaactttc 180
gcggtggcct ggcggcggct atgggtgggc cagcggcatg ggaggcatca cccgcagtta 240
cggcaaccag agcctgctga gcccttgcc tggaggngga cccaacatc aagccgngcg 300
caccaggaa aaggagcaga ncaagaccct caacaacaag nttgcttctt catagacaag 360
ggaccggtcc ttgaacagca naacaagatg ntggag 396

```

&lt;210&gt; 116

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

<221> misc\_feature  
<222> 267, 290, 343, 351, 376  
<223> n = A,T,C or G

<400> 116  
atctcagttt actagctaag tgactttggg caagggattt aacctctcgt ccctcagttt 60  
cctcctatgt aaaatgacaa ggataatagt accaacccaa thtagattaa atgagtttac 120  
gaagtgttag aatagtgtt ggcacattag tgctttacaa ctgctatttt gattgttgtt 180  
gtgggctctc tcaaatgcat tgtctctaga tgccagtgc ccaggtcaaa atttaccttt 240  
aaccaagctg catgtttccc agactgntgc acagtctct accctgagan aaagcttcca 300  
cccaaggata cttttacttt ctgctggaaa actgatgagc aanggcaaca ngggacactt 360  
atcgccaact ggaaangaga aattcttcct tttgct 396

<210> 117  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 228, 267, 318, 331, 357, 368, 376  
<223> n = A,T,C or G

<400> 117  
aaacattttt taataaaatt cctatagaaa gtcagtcatt agggcaaata ctacagttctc 60  
tttcccatat caccgaggat tgagagctcc caatattctt tggagaataa gcagtagttt 120  
tgctggatgt tgccaggact cagagagatc acccatttac acattcaaac cagtagttcc 180  
tattgcacat attaacatta cttgccccta gcaccctaaa tataatggnac ctcaacaaat 240  
aacttaaaga tttccgtggg gcgcganacc atttcaattt gaactaatat ccttgaaaaa 300  
aatcacatta ttacaagnnt taataaatac nggaagaaga gctggcattt ttctaanaac 360  
tgaattcnga cttggnttta ttccataaat acggtt 396

<210> 118  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 4, 5, 12, 14, 15, 16, 24, 59, 80, 87, 225, 280, 286, 287,  
295, 297, 298, 337, 349, 362, 375, 387, 394  
<223> n = A,T,C or G

<400> 118  
accnncacct gntnnntttt aacnattaca acttctttat atggcagttt ttactgggng 60  
cctaacactc tctttactgn ctcaagngga agtccaaaca aatttcattt ttgtagtaaa 120  
aaatctttat ttccaaaatg atttgtttagc caaaagaact ataaaccacc taacaagact 180  
ttggaagaaa gagacttgat gcttcttata aattcccat tgcanaaaaa aaataacaat 240  
ccaacaagag catggtaccc attcttacca ttaacctggn tttaannctc caaancnnga 300  
tttaaaaatg accccactgg gcccaatcca acatganacc taggggggnt tgccttgatt 360  
angaatcccc cttanggact ttatctnggc tganaa 396

<210> 119  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>

<221> misc\_feature

<222> 251, 281, 298, 301, 308, 326, 332, 337, 351, 358, 362, 388, 394

<223> n = A,T,C or G

<400> 119

```
atggccagct cactttaaat accacctcaa gactcatcga aatgaccgct cttcatctg 60
tcctgcagaa ggttgtgga aaagcttcta tgtctgcag aggtgaagg tgcacatgag 120
gaccacaat ggagagaagc cctttatgtg ccatgagtct ggctgtgga agcagtttac 180
tacagctgga aacctgaaga accaccggcg catccacaca ggagagaaac ctttcctttg 240
tgaagcccaa ngatgtggcc gtcctttgct gagtattcta ncttcgaaaa catctggngg 300
ntactcanga gagaaagcct cattantgcc antctgnggg aaaaccttct ntacagagngg 360
angcaggaat gtgcatatta aaaagctncc ttgnac 396
```

<210> 120

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 261, 263, 265, 272, 273, 288, 308, 310, 330, 379

<223> n = A,T,C or G

<400> 120

```
catgggtcag tcggtcctga gagttcgaag agggcacatt cccaaagaca ttcccagtca 60
tgaaatgtag aagactggaa aattaagaca ttatgtaaag gtagatatgg cttttagagt 120
tacattatgc ttggcatgaa taagggtgcc ggaaaacagt ttaaaattat acatcagcat 180
acagactgct gttagaaggt atgggatcat attaaagataa tctgcagctc tactacgcat 240
ttattgttaa ttgagttaca nangncattc annactgagt ttatagancc atattgctct 300
atctctgn gn agaacatttg attccattgn gaagaatgca gtttaaaata tctgaatgcc 360
atctagatgt attgtaccna aaggggaaaa ataaca 396
```

<210> 121

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 77, 125, 130, 142, 155, 162, 166, 176, 204, 227, 242, 243, 245, 246, 249, 251, 252, 265, 279, 306, 310, 314, 336, 341, 354, 367, 382, 385, 390, 395

<223> n = A,T,C or G

<400> 121

```
ttttttttt ttttttttaa aatcaagtta tgtttaataa acattaataa atgtttactt 60
aaaagggtta ataaacnttt actacatggc aaattatttt agctagaatg cttttggctt 120
caagncatan aaaccagatt cnaatgccct taaanaattt tnaaanatcc attgangggg 180
ataactgtaa tccccaaggg gaanagggtt gggtatgaca ggtacanggg gccagcccag 240
tnntnncana nncagactct tacntcttt ctgctgtgnc accctcaggc attggctcca 300
ttctcngggg tgcncatggg aagatggctt tggacntaac nacacccttt tgnacacgta 360
aaggccngat gcagggtcaa anagnttccn ccatnt 396
```

<210> 122

<211> 396

<212> DNA

<213> Homo sapiens

```

<400> 122
gtcgacatgg ctgccctctg ggctcccaga acccacaaca tgaaagaaat ggtgctaccc 60
agctcaagcc tgggcctttg aatccggaca caaaaccctc tagcttggaa atgaatatgc 120
tgcacttttac aaccactgca ctacctgact caggaatcgg ctctggaagg tgaagctaga 180
ggaaccagac ctcatcagcc caacatcaaa gacaccatcg gaacagcagc gcccgagca 240
cccccccgcc accggcgact ccattctcat ggccaccccc tgcggtggac ggttgaccac 300
cagccaccac atcatcccag agctgagctc ctccagcggg atgacgccgt cccaccacc 360
tccctcttct tctttttcat ccttctgtct ctttgc 396

```

```

<210> 123
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 74, 94, 142, 149, 194, 219, 233, 279, 316, 335, 368
<223> n = A,T,C or G

```

```

<400> 123
gccctttttt tttttttttt tttcctagtg ccaggtttat tccctcacat ggggtggttca 60
catacacagc acanaggcac gggcaccatg gganagggca gcactcctgc cttctgaggg 120
gatcttggcc tcacggtgta anaagggana ggatggtttc tcttctgccc tcaactaggc 180
ctaggaacc cagnagcaaa tcccaccacg ccttccatnt ctacgccaag ganaagccac 240
cttggtagcg tttagttcca accattatag taagtggana agggattggc ctggtcccaa 300
ccattacagg gtgaanatat aaacagtaaa ggaanataca gtttggatga ggccacagga 360
aggagcanat gacaccatca aaagcatatg cagggga 396

```

```

<210> 124
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<400> 124
gaccattgcc ccagacctgg aagatataac attcagttcc caccatctga ttaaaacaac 60
ttcctccctt acagagcata caacagaggg ggcacccggg gaggagagca catactgtgt 120
tccaatttca cgcttttaat tctcatttgt tctcacacca acagtgtgaa gtgctgtgta 180
taatctccat ttcaaaacca aggaagcagc ctgagagtgg tcgagtgaca cacctcacgc 240
aggctgagtc cagagcttgt gtcctcttgc attcctggtt tgactcagtt ccaggcctga 300
tcttgccctgt ctggctcagg gtcaaagaca gaatgggtga gtgtagcctc cacctgatat 360
tcaggctact cattcagtc caaatatgta ttttcc 396

```

```

<210> 125
<211> 396
<212> DNA
<213> Homo sapiens

```

```

<220>
<221> misc_feature
<222> 43, 88, 91, 94, 139, 141, 150, 163, 193, 202, 212, 215, 222,
238, 253, 256, 286, 297, 331, 343, 350, 360, 376, 385, 396
<223> n = A,T,C or G

```

```

<400> 125
cccttttttt tttttttttt tttttttttt ttttttactt tgnaacaaaa atttattagg 60
attaagtcaa attaaaaaac ttcatgcnc nccncttgct atatttacct gaaatgacaa 120
agttatactt agcttgagng naaaacttgn gccccaaaaa ttntgtttgg aaagcaaaaa 180

```

```

aataattgat gcncatagca gngggcctga tncnccaca gngaagtgtg ttttaaggnt 240
aacaacacagg ggnccancaa gcatacatta cttttaagct ttgggnccaa ggaaaangtc 300
attccctacc tccttcaaaa gcaaacatcat natagcctgg gcncctaggc ctggagcctn 360
ttttttcgag tctaanatga acatntggat ttcaan 396

```

<210> 126

<211> 396

<212> DNA

<213> Homo sapiens

<400> 126

```

cgcgtcgact cgcaagtgga atgtgacgtc cctggagacc ctgaaggctt tgcttgaagt 60
caacaaaggg cagcaaatga gtcctcaggt ggccaccctg atcgaccgct ttgtgaagg 120
aaggggccag ctagacaaag acaccctaga caccctgacc gccttctacc ctgggtacct 180
gtgtccctc agccccgagg agctgagctc cgtgcccccc agcagcatct gggcggtcag 240
gccccacgac ctggacacgc tggggctacg gctacagggc ggcacccca acggctacct 300
ggtcctagac ctcagcatgc aagaggccct ctgggggacg ccctgcctcc taggacctgg 360
acctgttctc accgtcctgg cactgtcctc agcctc 396

```

<210> 127

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 15

<223> n = A,T,C or G

<400> 127

```

tttttttttt ttgnggtaa aatgcaaatg ttttaaaata tgttatattt gtatgtttta 60
caatgaatac ttcagcaaaag aaaataatta taatttcaaa atgcaatccc tggatttgat 120
aaatatcctt tataatcgat tacactaatc aatatctaga aatatacata gacaaagtta 180
gctaataaat aaaataagta aaatgactac ataaactcaa tttcagggat gagggatcat 240
gcatgatcag ttaagtcaact ctgccacttt ttaaaataat acgattcaca tttgcttcaa 300
tcacataaac attcattgca ggagttacac ggctaatacat tgaaaattat gatcctttgtt 360
agcttaaaag aaaattcagt ttaatacaaa gacatt 396

```

<210> 128

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 220, 244, 351, 384

<223> n = A,T,C or G

<400> 128

```

gccctttttt ttttttttta aaggcaaata aaataagttt attgggatgt aaccccatca 60
taaattgagg agcatccata caggcaagct ataaaatctg gaaaatttaa atcaaattaa 120
attctgcttt taaaaggtg ccttaagtta accaagcatt ttgataacac attcaaattt 180
aatatataaa aatagatgta tcctggaaga tataatgaan aacatgccat gtgtataaat 240
tcanaatacg cttttttacac aaagaactac aaaaagttac aaagacagcc ttcaggaacc 300
acacttagga aaagttagcc gagcagcctt cagcgaagc ctccttcaaa naagtctcac 360
aaagactcca gaaccagccg agtntgtgaa aaagga 396

```

<210> 129

<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 104, 164, 177, 204, 217, 234, 273, 312, 350, 353, 370  
<223> n = A,T,C or G

<400> 129  
gccctttttt tttttttttt ttttactcag acaggcaata tttgctcaca tttattctct 60  
tgcatcgtaa atagtagcca actcacaaaa ataaagtata caanaatgta atatttttta 120  
aaataagatt aacagtgtaa gaaggaaaat ctcaaaaaaa gcanatagac aatgtanaaa 180  
attgaaatga aatcccacag taanaaaaaa aaaacanaaa agtgcctatt taanaattat 240  
gctacatgtg gaacttaact agaccathtt aanaaagacc aatttctaata gcaaattttc 300  
tgaggttttc anattttatt tttaaaaatat gttatagcta catgttgctn acncggccgc 360  
tcgagtctan agggcccgtt taaaccgcgt gatcag 396

<210> 130  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 23, 24, 26, 32, 56, 191, 286, 355  
<223> n = A,T,C or G

<400> 130  
cgcccttttt tttttttttt tanngnacgt gncctttatt ctggatgata taaaanaaaa 60  
aacttaaaaa acaccccaaa ccaaacacca atggatcccc aaagcgatgt gactccctct 120  
tcccacccgg ataaatagag acttctgtat gtcagtctac cctcccgcgc ccataacccc 180  
ctctgctata nacatactct gggtatatat tactctactc ggcaatagac atctcccgaa 240  
aatagaattc ctgccctgac acctgactct tccctggcgc catcanacca cccgccactg 300  
tagcacactg gtgtccttgc cccctgtggt caggggccatg ctgtcatccc acaanaagge 360  
cacatttgtc acatggctgc tgtgtccacc gtactt 396

<210> 131  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 49, 68, 69, 83, 88, 93, 136, 140, 154, 158, 166, 167, 168,  
170, 172, 173, 187, 226, 239, 241, 247, 257, 259, 271, 293,  
301, 318, 334, 336, 342, 344, 357, 377, 384  
<223> n = A,T,C or G

<400> 131  
gccctttttt tttttttttt tttttttttt ttcagtttac acaaaaaacnc ttttaattgac 60  
agtatacnnt tttccaaaat atnttttngt aanaaaatgc aataattatt aactatagtt 120  
tttacaaaca agttnttcan taaattccag tgtntctnaa acccnnncn annaaaacat 180  
atatgancce ccagttcctg ggcaaaactgt tgaacattca ctgcanacaa aaagaccanc 240  
nccaaanagt catctgngnc ctccatgctg ngtttgacc aaacctgagg gancagctag 300  
ngaccgtgac aaaagctntg ctacagtttt actntngccc tntntgcctc ccccatnatg 360  
tttccttggt ccctcantcc tgnngagta agttcc 396

<210> 132  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 69  
<223> n = A,T,C or G

<400> 132  
cgcgctcgacc gcggccgtag cagccgggct ggtcctgctg cgagccggcg gcccgagtg 60  
gggcggcgnt atgtaccttc cacattgagt attcagaaaag aagtgatctg aactctgacc 120  
attctttatg gatacattaa gtcaaataata agagtctgac tacttgacac actggctcgg 180  
tgagttctgc tttttctttt taatataaat ttattatgtt ggtaaattta gcttttggct 240  
tttcactttg ctctcatgat ataagaaaat gtaggttttc tctttcagtt tgaattttcc 300  
tattcagtaa aacaacatgc tagaaaacaa acttttggaa aggcatgtga actatttttt 360  
caaatagaac cataataaca agtcttgtct taccct 396

<210> 133  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 1, 17, 18, 20, 21, 25, 26, 30, 31, 40, 44, 45, 46, 51, 52,  
66, 67, 68, 74, 89, 109, 122, 166, 193, 214, 218, 266, 269,  
291, 307, 315, 348, 375, 378, 379, 386, 393  
<223> n = A,T,C or G

<400> 133  
ntattacccc tcctggnnan ntggnnatan nctgcaaggn gatnnncccc nngaacttca 60  
ctgatnnncc aatnaaaaact gcttttaaanc tgactgcaca tatgaattnt aatacttact 120  
tngcgggagg ggtggggcag ggacagcaag ggggaggatt gggaanacaa tagacaggca 180  
tgctggggat gcngcgggct ctatggcttc tgangcgnaa agaaccagct ggggctctag 240  
ggggtatccc cagcgcccct gtagcngcnc attaaacgcg gcgggtgtgg nggttacttc 300  
gcaaagngac cgatncactt gccagcgccc tagctgcccg ctccttngc tttcttccct 360  
tcctttctcg ccacnttnc cgctntccc cgncaa 396

<210> 134  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 133, 144, 221, 229, 302, 358  
<223> n = A,T,C or G

<400> 134  
tttttttttt ttctgctttt tatatgttta aaaatctctc attctattgc tgctttattt 60  
aaagaaagat tactttcttc cctacaagat ctttattaat tgtaaaggga aaatgaataa 120  
ctttacaatg ganacacctg gcanacacca tcttaaccaa agcttgaagt taacataacc 180  
agtaatataga ctgatcaata tcttgtgcct cctgatatgg ngtaactaana aaaacacaac 240  
atcatgccat gatagtcttg ccaaagtgatg ataacctaaa tctaatacata aggaaacatt 300  
anacaaactc aaattgaagg acattctaca aagtgccctg tattaaggaa ttattcanag 360  
taaaggagac ttaaaagaca tggcaacaat gcagta 396

<210> 135  
<211> 396  
<212> DNA  
<213> Homo sapiens

<400> 135  
gcgtcgacgc tggcagagcc acacccaag tgctgtgcc cagagggtt cagtcagctg 60  
ctcactcctc cagggcactt ttaggaaagg gtttttagct agtggttttc ctgcgtttta 120  
atgacctcag ccccgctgc agtggttaga agccagcagg tgcccatgtg ctactgacaa 180  
gtgcctcagc tccccccgg cccgggtcag gccgtgggag ccgctattat ctgcgttctc 240  
tgccaaagac tcgtgggggc catcacacct gccctgtgca gcggagccgg accaggctct 300  
tgtgtcctca ctacaggtttg cttccctgt gccactgct gtatgatctg ggggccacca 360  
ccctgtgccg gtggcctctg ggctgcctcc cgtgggt 396

<210> 136  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 18, 185, 188, 191, 193, 396  
<223> n = A,T,C or G

<400> 136  
ttatgcttcc ggctcgntg ttgtgtggaa ttgtgagcgg ataacaattt cacacaggaa 60  
acagctatga ccatgattac gccaaagctat ttaggtgaca ctatagaata ctcaagctat 120  
gcatcaagct tggtagcgag ctccgatcca ctagtaacgg ccgccagtgt gctggaattc 180  
gcggncgntc nantctagag ggccggttta aaccgctga tcagcctcga ctgtgccttc 240  
tagttgccag ccatctgttg ttgcccctc ccccgctgcct tccttgaccc tggaaagggtgc 300  
cactcccaact gtcctttcct aataaaatga ggaaattgca tcgcattgtc tgagtaggtg 360  
tcattctatt ctgggggggtg gggtggggca ggacan 396

<210> 137  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 156, 216  
<223> n = A,T,C or G

<400> 137  
tttttttttt ttctgctttg tacttgagtt tatttcacaa aaccacggag aaagatactg 60  
aatggagct ctttccagcc tccaagcaag gagggcccag cagccagtct ccagcccctt 120  
gagccctttt tgtaggccc acacccaaaa gagganaacc agtggtgtgc cgaagggtaca 180  
tggcaaggca cttttgaaaa catcccagtt taccngggtg aaattgaact tactctgaaa 240  
cagatgaaaa gggacatgca aaattgctga gcacatggag gtgtttgtta gtaggtgaaa 300  
atcatgtcct ggggtataacc cagcttctcc aggttagggg gagccgccgt ctggatcagt 360  
ggtggcgggc cacacaccag gatgagcgtg gacttc 396

<210> 138  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> 69, 136, 265, 272  
 <223> n = A,T,C or G

<400> 138  
 cccttttttt ttttttttac aaatgagaaa aatgtttatt aagaaaacaa ttttagcagct 60  
 ctcttttana attttacaga ctaaagcaca acccgaaggc aattacagtt tcaatcatta 120  
 acacactact taaggngctt gcttactcta caactggaaa gttgctgaag tttgtgacat 180  
 gccactgtaa atgtaagtat tattaaaaat tacaaattgt ttggtgatta ttttgatgac 240  
 ctcttgagca gcagctcccc ccaanaatgc ancaatggta tgtggctcac cagctccata 300  
 tcggcaaaat tcgtggacat aatcatcttt caccattaca gataaacat attcctgaag 360  
 gaagccagtg agacaagact tcaactttcc tatatc 396

<210> 139  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> 51, 105, 126, 147, 210, 212, 236, 241, 258, 263, 348  
 <223> n = A,T,C or G

<400> 139  
 ccgccctttt tttttttttt ttcacaaaag cactttttat ttgaggcaaa nagaagtctt 60  
 gctgaaagga ttccagttcc aagcagtcaa aactcaaccg ttagnngcac tattttgacc 120  
 tgggtanattt tgcttctctt tggtcanaaa aggggtattca ggttgactt tccccagcag 180  
 ggtaaaaaga agggcaaaagc aaactggaan anacttctac tctactgaca gggctnttga 240  
 natccaacat caagctanac acnccctcgc tggccactct acaggttgct gtccactgc 300  
 tgagtgcacac aggcataact acatttgcaa ggaaaaaaat gaggcaanaa acacaggtat 360  
 aggtcacttg gggacgagca ggcaaccaca gtttca 396

<210> 140  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> 50, 60, 63, 100, 133, 135, 172, 183, 190, 196, 220, 240,  
 262, 266, 273, 278, 293, 327, 332, 341, 348, 355, 380, 391  
 <223> n = A,T,C or G

<400> 140  
 tttttttttt tttttttttt tttttttctc atttaacttt tttaatgggn ctcaaaaatn 60  
 tgngacaaat ttttggtcaa gttgtttcca ttaaaaagtn ctgattttta aaactaataa 120  
 cttaaaactg ccncncccaa aaaaaaaaaac caaaggggtc cacaaaacat tntcctttcc 180  
 ttntgaagggn ttacnatgc attgttatca ttaaccagtn ttttactact aaacttaaan 240  
 ggccaattga aacaaacagt tntganaccg ttnttcncc actgattaaa agnggggggg 300  
 caggtattag ggataatatt catttancct tntgagcttt ntgggcanac ttgngnacct 360  
 tgccagctcc agcagccttn ttgtccactg ntttga 396

<210> 141  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

&lt;400&gt; 141

```

acgccgagcc acatcgctca gacacccatgg ggaagggtgaa ggtcggagtc aacggatttg 60
gtcgtattgg gcgcctggtc accagggtcg cttttaactc tggtaaagtg gatattgttg 120
ccatcaatga ccccttcatt gacctcaact acatgggtta catgttccaa tatgattcca 180
cccatggcaa attccatggc accgtcaagg ctgagaacgg gaagcttgtc atcaatggaa 240
atcccatcac catcttccag gagcgagatc cctccaaaat caagtggggc gatgctggcg 300
ctgagtacgt cgtggagtc actggcgtct tcaccaccat ggagaaggct ggggctcatt 360
tgcagggggg agccaaaagg gtcacatct ctgcc 396

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&lt;210&gt; 142

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 142

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acgcaggaga ggaagcccag cctgttctac cagagaactt gcccagggtca gaggtctgcg 60
tagaagccct tttctgagca tcctctctc tcctcacacc tgccactgtc ctctgcgttg 120
ctgtcgaatt aaatcttgca tcacccatgg gcacttctgt ggccactca cctccaccg 180
ggagccagtgc ccgctgaaga gtatctctgt gacgtgaac atttacgagt ttgtggctgg 240
tgtgtctgca actttgaact acgagaatga ggagaaagt cctttggagg ccttctttgt 300
gttccccatg gatgaagact ctgctgttta cagctttgag gccttggtgg atgggaagaa 360
aattgtagca gaattacaag acaagatgaa ggcccc 396

```

&lt;210&gt; 143

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; 19, 48, 69, 122, 183, 227, 332, 390

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 143

```

tttttttttt ttccatana aaataggatt tattttcaca ttttaaggnga acacaaatcc 60
atgttccana aatgttttat gcataacaca tcatgagtag attgaatttc tttacacac 120
anaaaatca aagcctacca ggaaatgctt ccctccggag cacaggagct tacaggccac 180
ttntgttagc aaacaggaa ttcacattgt ctaggcacag ctcaagngag gtttgtccc 240
aggttcaact gtcctaccc ccatgggccc tcctcaaaaa cgacagcagc aaaccaacag 300
gcttcacagt aaccaggagg aaagatctca gngggggaac cttcacaaaa gccctgagtt 360
gtgtttcaaa agccaagctc tggggtctgn ggccctg 396

```

&lt;210&gt; 144

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; 221, 331

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 144

```

tttttttttt ttctgctctt tggctctgaca agaaaagagt tttagggtgtg tgaagtaggg 60
tgggaaaaaa ggtcagtttc aaattcagta acatatggta aactaagtt aggctgctgc 120
attcttttct ttgggtactt aagccagctg gcacttcac tttgtaacca attatattat 180
gatcaacaac taatcagtta gttcctcagc ttcaactgaa nagttcctga ttacctgatg 240
aaggacatac ttgctctggc ttcaattagc atgctgtcaa gcatccctct ccatgcttaa 300

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catggcaaca caaaacccaa gagtccttct ntttttttca ttagccatga ataaacactc 360  
 acaaagggga agagtagaca ctgcttttag taaacg 396

<210> 145

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 45, 56, 61, 63, 120, 122, 147, 151, 158, 259, 262, 274, 339,  
 345, 353

<223> n = A,T,C or G

<400> 145

ttttttttt tttttttcaa tggatccggt agctttacta ctaanatctt gctganatca 60  
 nanaagggt tctgggcagg ctgagcactg ggggtgtgca acatggtaac tctgaataan 120  
 anaaaccctg agttttactg ggcaanaaaa naacaagngg taggtatgat ttctgaacct 180  
 ggaaatagcg aaaatgaagg aaattccaaa agcgcgtatt tccaaataat gacaggccag 240  
 caagaggaca ccaaacctnt anaaagaggt attntttctt ccagctactg atggctttgg 300  
 catcccacag gcacattcct ttggccttca ggatcttana tgcanatgtg ganagtcaag 360  
 aggtaggctg actctgagtc ttcagctaaa ttcttt 396

<210> 146

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 120, 130, 176, 180, 185, 208, 238, 254, 259, 261, 275, 285,  
 296, 347

<223> n = A,T,C or G

<400> 146

ttttttttt ttttcattag caaggaagga tttattttt cttttgaggg gagggcggaa 60  
 cagccgggat ttttgaaca ctaccttgt ctttcacttt gttgtttgtg tgtaaacacn 120  
 aataaatcan aagcgacttt aaatctccct tcgcaggact gtcttcacgt atcagngcan 180  
 acaanaaaac agtggcttta caaaaaanat gttcaagtag gctgcacttt gcctctgngg 240  
 gtgaggcaca ctgngggana nacaaggtcc cctgnaacca gagnggggaa ggacanagct 300  
 ggctgactcc ctgctctccc gcattctctc ctccatgtgt tttgaanagg gaagcaacat 360  
 gttgaggtct gatcatttct acccagggaa cctgtt 396

<210> 147

<211> 396

<212> DNA

<213> Homo sapiens

<400> 147

acggggaagc caagtgaccg tagtctcatc agacatgagg gaatgggtgg ctccagagaa 60  
 agcagacatc attgtcagtg agcttctggg ctcatattgct gacaatgaat tgcgcctga 120  
 gtgcctggat ggagcccagc acttcctaaa agatgatggg gtgagcatcc ccggggagta 180  
 cacttccttt ctggctccca tctcttcttc caagctgtac aatgaggtcc gagcctgtag 240  
 ggagaaggac cgtgaccctg aggccagtt tgagatgcct tatgtggtac ggctgcacaa 300  
 cttccaccag ctctctgcac cccagccctg tttcaccttc agccatccca acagagatcc 360  
 tatgattgac aacaaccgct attgcacctt ggaatt 396

<210> 148

<211> 396  
 <212> DNA  
 <213> Homo sapiens

<400> 148  
 acgtcccatg attgttccag accatgactc ttcctggttg tgggtttggt acagagcagg 60  
 agaagcagag gttatgacag ttatgcagac tttccccctc ctttttctct tttctcttcc 120  
 ccttgctttt ccaactgtttc ttcctgctgc cacctgggcc ttgaattcct gggctgtgaa 180  
 gacatgtagc agctgcaggg tttaccacac gtgggagggc agcccagtac tgtccctctg 240  
 ccttccccac tttgagaata tggcagcccc tttcattcct ggcttggggg aggggagacc 300  
 attgaagtag aagcctcaaa gcagactttt ccctttactg tgtgtactcc aggacgaaga 360  
 aggaagatca tgcttgatac ttagattggt tttccc 396

<210> 149  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> 214, 295  
 <223> n = A,T,C or G

<400> 149  
 tttttttttt tttaaagagt cacattttat tcaatgccta tttgtacatg ttactagcaa 60  
 taaactcttt tatctttaat tttgagaagt tttacaaata cagcaaagca gaatgactaa 120  
 tagagccggg aaccaggaca cagatttgga aaaatagggtc taattgggtg ttacactgtg 180  
 tttatgtcat acatttcgct tatttttatc aaanaaaaat cagaatttat aaaatgttaa 240  
 ttaaaaggaa aacattctga gtaaatttag tcccgtgttt cttcctccaa atctntttgt 300  
 tctacactaa caggtcagga taagtatgga tggggagggt ggaaaaaggg catccttccc 360  
 catgcgggtcc ccagagccac cctctccaag caggac 396

<210> 150  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<400> 150  
 acgcctctct tcagttggca cccaaacatc tggattggca aatcagtggc aagaagttcc 60  
 agcatctgga cttttcagaa ttgatcttaa gtctactgtc atttccagat gcattatatt 120  
 acaactgtat ccttggaat atatttctag ggagaatatt attgaagaaa atgttaatag 180  
 cctgagtcaa atttcagcag acttaccagc atttgtatca gtggtagcaa atgaagccaa 240  
 actgtatctt gaaaaacctg ttgttccttt aaatatgatg ttgccacaag ctgcattgga 300  
 gactcattgc agtaatatct ccaatgtgcc acctacaaga gagatacttc aagtctttct 360  
 tactgatgta cacatgaagg aagtaattca gcagtt 396

<210> 151  
 <211> 396  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> 146, 299, 332  
 <223> n = A,T,C or G

<400> 151  
 acaaaatgcc cagcctacag agtctgagaa ggaaatttat aatcagggtga atgtagtatt 60

```

aaaagatgca gaaggcatct tggaggactt gcagtcatac agaggagctg gccacgaaat 120
acgagaggca atccagcatc cagcanatga gaagttgcaa gagaaggcat ggggtgcagt 180
tgttccacta gtaggcaaatt taaagaaatt ttacgaattt tctcagaggt tagaagcagc 240
attaagaggt cttctgggag ccttaacaag taccatcat tctcccaccc agcatctana 300
gcgagagcag gctcttgcta aacagtttgc anaaattctt catttcacac tccgggttga 360
tgaactcaag atgacaaatc ctgccatata gaatga 396

```

<210> 152

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 249

<223> n = A,T,C or G

<400> 152

```

acgcagcgct cggcttcctg gtaattcttc acctcttttc tcagctccct gcagcatggg 60
tgctggggccc tccttgctgc tcgcccgcct cctgctgctt ctctccggcg acggcgccgt 120
gcgctgcgac acacctgcc aactgcaccta tcttgacctg ctgggcacct gggcttcca 180
gggtgggctcc agcgggtccc agcgcgatgt caactgctcg gttatgggac cacaagaaaa 240
aaaagtagn ggtgtacctc agaagctgga tacagcatat gatgaccttg gcaattcttg 300
ccatttcacc atcatttaca accaaggctt tgagattgtg ttgaatgact acaagtgggt 360
tgcctttttt aagtataaag aagagggcag caaggt 396

```

<210> 153

<211> 396

<212> DNA

<213> Homo sapiens

<400> 153

```

ccagagacaa cttcgcgggtg tggatgaactc tctgaggaaa aacacgtgctg tggcaacaag 60
tgactgagac ctgaaatcc aagcgttggg ggtcctgagg ccagcctaag tcgcttcaaa 120
atggaacgaa ggcgtttgctg ggggtccatt cagagccgat acatcagcat gagggtgtgg 180
acaagcccac ggagacttgt ggagctggca gggcagagcc tgctgaagga tgaggccctg 240
gccattgccg ccctggagtt gctgcccagg gagctcttcc cgccactctt catggcagcc 300
tttgacggga gacacagcca gacctgaag gcaatgggtgc aggcctggcc cttcacctgc 360
ctccctctgg gagggtctgat gaagggacaa catctt 396

```

<210> 154

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 42, 45, 59, 82

<223> n = A,T,C or G

<400> 154

```

acagcaaac tcctcacagc ccactgggtcc tcaagagggg cnacntcttc acacatcanc 60
acaactacgc attgcctccc tncactcgga aggactatcc tgctgccaag aggggtcaagt 120
tggacagtgt cagagtctctg agacagatca gcaacaaccg aaaatgcacc agccccaggt 180
cctcgacac cgaggagaat gtcaagaggc gaacacacaa cgtcttggag cgccagagga 240
ggaacgagct aaaacggagc ttttttggcc tgcgtgacca gatcccgag ttggaaaaca 300
atgaaaaggc ccccaaggta gttatcctta aaaaagccac agcatacatc ctgtccgtcc 360
aagcagagga gcaaaaagctc atttctgaag aggact 396

```

<210> 155  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 15, 17, 202, 280, 339  
<223> n = A,T,C or G

<400> 155  
tttttttttt tgaananaca ggtctttaat gtacggagtc tcacaaggca caaacaccct 60  
caccaggacc aaataaataa ctccacggtt gcaggaaggc gcggtctggg gaggatgcgg 120  
catctgagct ctcccagggc tgggtgggca gccgggggtc tgcagtctgt gaggggcctc 180  
ctgggtgtgt ccgggcctct anagcgggtc cagtctccag gatggggatc gctcactcac 240  
tctccgagtc ggagtagtcc gccacgaggg aggagccgan actgcagggg tgccgcgtgt 300  
cgggggtgtc agctgcctcc tgggaggagc ctgctggcna caggggcttg tcctgacggc 360  
tcccttctctg cccctcggg ctgctgcact tggggg 396

<210> 156  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 11, 30, 32, 37, 309, 332  
<223> n = A,T,C or G

<400> 156  
gaaggggggc ngggcagggg cggaatgtan anattantgc catgattgaa gatttaagaa 60  
acgtgagatt caggattttc accacatccc catttagtta gcttgctcgt ttggctgggtg 120  
caaattgccag atggattatg aacaatgaca gtaaattaat gcaacataat caggtaatga 180  
tgccaagcgt atctggtgtt ccaggatttg tacctttacc ggaacaaatc agtaaattcca 240  
caatccctgg cacctgtagg gcagctatta acctagtaa tgctcccca tcccatctca 300  
atcagcaang acaatcaaaa acatttgctt tnagtggcag gaacactggt acatttttac 360  
ttgtccaag ggctgtgcc aacgtccctc tctctg 396

<210> 157  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 121, 202, 204, 255, 314, 332, 368  
<223> n = A,T,C or G

<400> 157  
tttttttttt tttttgggga atgtaaatct tttattaaaa cagttgtctt tccacagtag 60  
taaaagctttg gcacatacag tataaaaaat aatcaccacac cataattata ccaaattcct 120  
nttatcaact gcatactaag tgttttcaat acaatttttt ccgtataaaa atactgggaa 180  
aaattgataa ataacaggta ananaaagat atttctaggc aattactagg atcatttgga 240  
aaaagtgtgagt actgnggata tttaaaatat cacagtaaca agatcatgct tgttcctaca 300  
gtattgcggg ccanacactt aagtgaagc anaagtgtt ggggtgacttt cctacttaaa 360  
attttggncat tatcatttca aaacatttgc atcttg 396

<210> 158  
<211> 396  
<212> DNA  
<213> Homo sapiens

<400> 158  
tttccgaaga cgggcagctt cagagaagag gattattcgg gagattgctg gtgtggccca 60  
tagactcttt ggcatagact ctttcgcagg cagccactct gactgtggcc agttctataa 120  
ccatccccaact actagctgga gcctgatgga taggaacggg tagtctgtcc tcttcccat 180  
aaaaatgttc caaaaagtta tctccagaga gagtccctta tgaagacagt tgccaagctg 240  
tattctcatt ctttaaacca ataccaggt cagggttagt tcacactagc actgttaggg 300  
acatgggtgtg gctagaaatg aattgagtgt gacttctccc tacaaccca ggcccaggga 360  
taggaggagg cagaggggtg cctggagttt ctgcac 396

<210> 159  
<211> 396  
<212> DNA  
<213> Homo sapiens

<400> 159  
tccgcgcgtt gggagggtga gcgcggctct gaacgcgctg agggccggtt agtgtcgcag 60  
gcggcgaggg cgcgagttag gagcagaccc aggcacgcg cgcgcagaag gccgggcgtc 120  
cccacactga aggtccgga aggcgacttc cgggggcttt ggcacctggc ggacctccc 180  
ggagcgctcg cactgaacg cgaggcgctc cattgcgcgt gcgcgttagg gggcttccc 240  
cacctgatcg cgagaccca acggtcgttg gcgtcgctc cgcgtctcgg ctgagctggc 300  
catggcgag ctgtgcgggc tgaggcggag ccgggcgttt ctgcacctgc tgggatcgct 360  
gctcctctct ggggtcctgg cggccgaccg agaacg 396

<210> 160  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 96, 102, 122, 124, 129, 146, 148, 184, 189, 196, 205, 208,  
229, 246, 259, 261, 269, 272, 281, 297, 305, 308, 327, 331,  
337, 338, 339, 343, 346, 354, 366, 367, 369, 378, 379, 380,  
381, 391, 395  
<223> n = A,T,C or G

<400> 160  
ggaaaccttc tcaactaaga gaacatcatt tctggcaaac tatttttggt agtcacaaat 60  
atatgtcgta cactctacaa tgtaaatagc actganccac ancttacaga aggtaaaaag 120  
angnataana acttccttta caaaanant cctgttggtt ttaatactcc ccattgctta 180  
tganaattnt ctatangtct ctcangantg ttcgcaccca tttctttnt aacttctact 240  
aaaaanccat ttacattgna nagtgtacna cntatatttg ngagctaaca aaaaatngtt 300  
ttcnganat gatgttcttt tagtttnaga nggttcnnc aanttntac tccngcccgc 360  
cactgnncnc cacatttnnn naattacacc ncacng 396

<210> 161  
<211> 396  
<212> DNA  
<213> Homo sapiens

<220>  
<221> misc\_feature  
<222> 271, 273, 325, 364

<223> n = A,T,C or G

<400> 161

```

tttttgtttg attattttta ttataatgaa attaaactta tgactattac agtatgtctca 60
gcttaaaaca ttatgagta ctgcaaggac taacagaaac aggaaaaatc ctactaaaaa 120
tatttgttga tgggaaatca ttgtgaaagc aaacctocaa atattcattt gtaagccata 180
agaggataag cacaaccata tgggaggaga taaccagtct ctcccttcat atatatctt 240
ttttatttct tggatatacct tcccaaaaaca nanacattca acagtagtta gaatggccat 300
ctccaacat tttaaaaaaa ctgcncccc caatgggtga acaaagtaaa gagtagtaac 360
ctanagttca gctgagtaag ccactgtgga gcctta 396

```

<210> 162

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 33, 38, 51, 62, 71, 72, 88, 97, 98, 100, 106, 142, 155, 160, 161, 163, 168, 170, 174, 183, 190, 194, 203, 214, 216, 231, 232, 241, 242, 252, 258, 260, 264, 265, 267, 276, 278, 282, 287, 289, 292, 295, 297, 301, 311, 319, 322, 325

<223> n = A,T,C or G

<221> misc\_feature

<222> 330, 337, 341, 342, 347, 348, 354, 356, 361, 367, 368, 375, 379, 385, 391, 394, 395

<223> n = A,T,C or G

<400> 162

```

tttttttttt tttttttttt tttttttttt ttnggggncc aaattttttt ntttgaagga 60
angggacaaa nnaaaaaact taaggggntg ttttggnnncn acttanaaaa aagggaaagg 120
aaaccccaac atgcatgccc tnccttgggg accanggaan ncncncncn ggnttgggga 180
aantaaccn aggnnttaact ttattatca ctgncnccca gggggggctt nnaaaaaaaa 240
nnttccccca anccaaantn gggnncccc attttncnca anttggncnc cnggncnccc 300
nattttttga ngggtttcnc cngcncattn agggaanggg nntcaannaa accncncaaa 360
nggggggnnat tttntcang ggccnatttg ngcnnt 396

```

<210> 163

<211> 396

<212> DNA

<213> Homo sapiens

<400> 163

```

cactgtccgg ctctaacaca gctattaagt gctacctgcc tctcaggcac tctcctcgcc 60
cagtttctga ggtcagacga gtgtctgcga tgtcttcccg cactctattc cccagcctc 120
tttctgcttt catgctcagc acatcatctt cctaggcagt ctcttcccca aagtctcacc 180
ttttcttcca atagaaaatt ccgcttgacc tttggtgac tgcccacttc ccagctccac 240
tgccccagt ctgagccgga ggcccttggt ttggggggcg ggggagagtt ggatgtgatt 300
gcccttgaag aacaaggctg acctgagagg ttcctggcgc cctgaggtgg ctcagcacct 360
gccagggtga ggcctggcat gaggggttag gtcagc 396

```

<210> 164

<211> 396

<212> DNA

<213> Homo sapiens

<400> 164

```

gacacgcggc ggtgtcctgt gttggccatg gccgactacc tgattagtgg gggcacgtcc 60
tacgtgccag acgacggact cacagcacag cagctcttca actgcggaga cggcctcacc 120
tacaatgact ttctcattct ccctgggtac atcgacttca ctgcagacca ggtggacctg 180
acttctgctc tgaccaagaa aatcactctt aagacccac tggtttcctc tcccatggac 240
acagtcacag aggcgtgggat ggccatagca atggcgctta caggcggtat tggcttcac 300
caccacaact gtacacctga attccaggcc aatgaagtgc ggaaagtga gaaatatgaa 360
cagggattca tcacagaccc tgtggtcctc agcccc 396

```

<210> 165

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 29, 33, 55, 57, 65, 77, 82, 87, 98, 101, 103, 114, 118, 124,  
169, 171, 173, 183, 186, 188, 216, 219, 227, 230, 242, 243,  
245, 252, 265, 273, 290, 296, 321, 324, 332, 338, 340, 342,  
345, 359, 372, 380

<223> n = A,T,C or G

<400> 165

```

tttttttttt tttttttttt ttttttcang ggnccactgag gctttttatt ttgancncaa 60
aaccnccggg gatctancct gnggccnccc cggaaatnac ncnaggctca catnactnta 120
aacncttggg ggaaagggag gcaaaaaaaa caatgacttg ggccaattnc ncnactgcaa 180
agntanant gccaacaggg ctccaggagag cttgntntnt gtaaaanttn taaggaagcg 240
gnncnaactc cncggggggg gggcnctaac tancaggagc ccctgcaagn gttggnccgg 300
ggcctcaacc tgctgagct nacncaaggg gnggggtntn tntanccaac aggggaccna 360
agggcttgcc tnccacagn ttacttgccc aagggg 396

```

<210> 166

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 151, 255

<223> n = A,T,C or G

<400> 166

```

ttttttcaaa ttcagagcat ttttattaaa agaacaaaat attaaggcac aaaatacatc 60
aatttttcaa atgaaaaccc ttcaaacggg tatgtcctac attcaacgaa acttcttcca 120
aattacggaa taatttaact ttttaaaata naaaaataca agttcttaaa tgcctaaaat 180
ttctcccaaa ataaatgttt tcttagtttt aatgaagtct cttcatgcag tactgagctc 240
caatattata atgtncactt ccttaaaaaat ctagttttgc cacttatata cattcaatat 300
gtttaaccag tatattaacc agtatattaa ccaatatgtt aaacttcttt taagtataag 360
gcttggtatt ttgtattgct tattgcatgc tttgat 396

```

<210> 167

<211> 396

<212> DNA

<213> Homo sapiens

<400> 167

```

tggcggcagc ggcgggtggcg gtggctgagc agaggaccgc gcgggcggcc tcgcgggtca 60
ggacacaatg tttgcacgag gactgaagag gaaatgtgtt ggccacgagg aagacgtgga 120
gggagccctg gccggcttga agacagtgtc ctcatacagc ctgcagcggc agtcgctcct 180

```

```

ggacatgtct ctggtgaagt tgcagctttg ccacatgctt gtggagccca atctgtgccg 240
ctcagtcctc attgccaaca cgggccggca gatccaagag gagatgacgc aggatgggac 300
gtggcgacaca gtggcacccc aggctgcaga gcgggcgccg ctcgaccgct tggctctccac 360
ggagatcctg tgccgtgcag cgtgggggca agaggg 396

```

&lt;210&gt; 168

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 168

```

taggatggta agagtattat aaggattggt acaaggcatg atgagtcctt ttgcttttag 60
gcttttgact tctggtttta gactttcttt agcttctggt gttagacaac attgtgcaag 120
cttggttttt ataagtttgc atggattaata ctgaacttaa tgaaattgtc cctcccccca 180
aattctcagc acaattttta ggcccacaag gagtcaagca cctcaaggag atcttcagtt 240
tgaacttggt gtagacacag ggatactgat gaatcaatat tcaaattagc tgttacctac 300
ttaagaaaga gaggagacct tggggatttc gaggaagggt tcataaggga gattttagct 360
gagaaatacc atttgcacag tcaatcactt ctgacc 396

```

&lt;210&gt; 169

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

```

<222> 16, 58, 76, 84, 99, 111, 114, 124, 136, 140, 161, 167, 184,
189, 204, 206, 210, 228, 230, 232, 243, 275, 277, 289, 301,
303, 312, 319, 321, 323, 325, 333, 345, 349, 355, 359, 364,
365, 372, 375, 377, 379, 383, 387, 389, 394, 396

```

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 169

```

tttttttttt tttcanaatt aaattcttta atacaaaatg cttttttttt tttaaaaanat 60
atctgtatatt ctttgnctgt gttnaaaaaat aaatatgtnc tacggaatat ntcnaaaaaac 120
tgcnctaaaa acaaanacgn gatgttaata tcttttcccc ncaattntta cggataaaca 180
gtancccccna taaataaatg atancnaatn ttaaaaattaa aaaagganan anatttagta 240
tgnaaaattc tctatttttt cttggttttg ttttncntat aaaaaacana atagcaatgt 300
ntntttttatc anaatcccnt ntntnccctaa acnttttttt tttnttttnc cccnnaatnc 360
aagnngccaa anatntntnt agnatgnana tgtntn 396

```

&lt;210&gt; 170

&lt;211&gt; 396

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 170

```

tgagaagtac catgccgctt ctgcagagga acaggcaacc atcgaacgca acccctacac 60
catcttccat caagcactga aaaactgtga gcctatgatt gggctggtac ccatcctcaa 120
gggaggccgt ttctaccagg tccctgtacc cctaccggac cggcgtcgcc gcttcctagc 180
catgaagtgg atgatcactg agtgccggga taaaaagcac cagcggacac tgatgccgga 240
gaagctgtca cacaagctgc tggaggcttt ccataaccag ggccccgtga tcaagaggaa 300
gcatgacttg cacaagatgg cagaggccaa ccgtgccttg gccactacc gctggtggta 360
gagtctccag gaggagccca ggccctctg cgcaag 396

```

&lt;210&gt; 171

&lt;211&gt; 396

&lt;212&gt; DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 133, 224, 260, 264, 268, 279, 283, 317, 322, 338, 360, 370, 371, 378

<223> n = A,T,C or G

<400> 171

```
ggtcctcgtc gtggtgagcg cagccactca ggctggtcct gggggtgggg ctgtagggga 60
aagtgctaaa gccgctgagt gaagtaagaa ctctgctaga gaggaaaatg ggcttgcttt 120
catcatcatc ctntcagct ggtgggggtca agtgggaagt tctgtcactg ggatctggtt 180
cagtgtctca agaccttgcc ccaccacgga aagccttttt cacntacccc aaaggacttg 240
gagagatggt agaagatggn tctnaaanat tctctgcna atntgttttt agctatcaag 300
tggcttcccc ccttaancag gnaaaacatg atcagcangt tgctcggatg gaaaaactan 360
cttggtttgn naaaaaanct ggaggcttga caatgg 396
```

<210> 172

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 239, 242, 244, 246, 249, 257, 260, 314, 329, 355, 372, 378, 385, 387, 388, 395

<223> n = A,T,C or G

<400> 172

```
agccttgggc caccctcttg gagcatctgg ctgtcgaatt cttgtgacct tgttacacac 60
actggagaga atgggcagaa gtcgtggtgt tgcagccctg tgcattgggg gtgggatggg 120
aatagcaatg tgtgttcaga gagaatgaat tgcttaaaact ttgaacaacc tcaatttctt 180
tttaaaactaa taaagtacta gggtgcaata tgtgaaaaaa aaaaaaaaaag ggcggccgnt 240
cnantntana gggcccnttn aaacccttg atcaacctcg actgtgcctt ctagttgcca 300
gccatctggt gttngccctt cccctgtgnc tttcttgacc ttgaaagggg cccnccct 360
gtctttccta anaaaaanga agaantnncc ttcent 396
```

<210> 173

<211> 396

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 209, 210, 232, 244, 270, 275, 284, 341, 343, 349, 359, 364, 368, 376, 380, 382, 388, 389, 390, 392

<223> n = A,T,C or G

<400> 173

```
aagcatgtgg atatgttttag ctacgtttac tcacagccag cgaactgaca ttaaaataac 60
taacaaacag attcttttat gtgatgctgg aactcttgac agctataatt attattcaga 120
aatgactttt tgaaagtaaa agcagcataa agaatttgtc acagggaaggc tgtctcagat 180
aaattatggt aaaattttgc aggggacann ctttttaaga cttgcacaat tnccggatcc 240
tgcnctgact ttggaaaagg catatatgtn ctagnngcat gganaatgcc ccatactcat 300
gcatgcaaat taaacaacca agtttgaate tttttggggg ngngctatnc ttttaaccng 360
tacnggcntt attatntaan gncctgnnn cntgtg 396
```

<210> 174

<211> 924  
 <212> DNA  
 <213> Homo sapiens

<400> 174  
 cctgacgacc cggcgacggc gacgtctctt ttgactaaaa gacagtgtcc agtgcctccag 60  
 cctaggagtc tacggggacc gcctcccgcg ccgccaccat gcccacttc tctggcaact 120  
 ggaaaatcat ccgatcggaa aacttcgagg aattgctcaa agtgcctggg gtgaatgtga 180  
 tgctgaggaa gattgctgtg gctgcagcgt ccaagccagc agtggagatc aaacaggagg 240  
 gagacacttt ctacatcaaa acctccacca ccgtgcgcac cacagagatt aacttcaagg 300  
 ttggggagga gtttgaggag cagactgtgg atgggaggcc ctgtaagagc ctggtgaaat 360  
 gggagagtga gaataaaatg gtctgtgagc agaagctcct gaaggagag ggccccaaga 420  
 cctcgtggac cagagaactg accaacgatg gggaactgat cctgaccatg acggcggatg 480  
 acgttgtgtg caccagggtc tacgtccgag agtgagtggc cacaggtaga accgcggccg 540  
 aagcccacca ctggccatgc tcaccgccct gcttactgc cccctccgtc ccaccccctc 600  
 cttctaggat agcgtccccc ttacccagc cacttctggg ggtcactggg atgcctcttg 660  
 cagggctcttg ctttctttga cctcttctct cctcccctac accaacaag aggaatggct 720  
 gcaagagccc agatcaccca ttccgggttc actcccgc tcccaagtc agcagtccta 780  
 gcccacaacc agccagagc aggtctctc taaaggggac ttgaggcct gagcaggaaa 840  
 gactggccct ctagcttcta cctttgtcc ctgtagccta tacagtttag aatatttatt 900  
 tgtaatttt attaaaatgc tttta 924

<210> 175  
 <211> 3321  
 <212> DNA  
 <213> Homo sapiens

<400> 175  
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&lt;211&gt; 487

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 176

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&lt;210&gt; 180

&lt;211&gt; 2382

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 180

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&lt;210&gt; 181

&lt;211&gt; 2377

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 181

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 Ser Met Cys Lys Ala Glu Thr Ser Gln Ala Glu Lys Cys Met Met Pro  
 595 600 605  
 Glu Cys His Thr Ile Pro Cys Leu Leu Ser Pro Trp Ser Glu Trp Ser  
 610 615 620  
 Asp Cys Ser Val Thr Cys Gly Lys Gly Met Arg Thr Arg Gln Arg Met  
 625 630 635 640  
 Leu Lys Ser Leu Ala Glu Leu Gly Asp Cys Asn Glu Asp Leu Glu Gln  
 645 650 655  
 Val Glu Lys Cys Met Leu Pro Glu Cys Pro Ile Asp Cys Glu Leu Thr  
 660 665 670  
 Glu Trp Ser Gln Trp Ser Glu Cys Asn Lys Ser Cys Gly Lys Gly His  
 675 680 685  
 Val Ile Arg Thr Arg Met Ile Gln Met Glu Pro Gln Phe Gly Gly Ala  
 690 695 700  
 Pro Cys Pro Glu Thr Val Gln Arg Lys Lys Cys Arg Ile Arg Lys Cys  
 705 710 715 720  
 Leu Arg Asn Pro Ser Ile Gln Lys Pro Arg Trp Arg Glu Ala Arg Glu  
 725 730 735  
 Ser Arg Arg Ser Glu Gln Leu Lys Glu Glu Ser Glu Gly Glu Gln Phe  
 740 745 750  
 Pro Gly Cys Arg Met Arg Pro Trp Thr Ala Trp Ser Glu Cys Thr Lys  
 755 760 765  
 Leu Cys Gly Gly Gly Ile Gln Glu Arg Tyr Met Thr Val Lys Lys Arg  
 770 775 780  
 Phe Lys Ser Ser Gln Phe Thr Ser Cys Lys Asp Lys Lys Glu Ile Arg  
 785 790 795 800  
 Ala Cys Asn Val His Pro Cys  
 805

&lt;210&gt; 187

&lt;211&gt; 892

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 187

```
tttattgatg tttcaacagg cacttattca aataagttat atatttgaaa acagccatgg 60
taagcatcct tggcttctca cccattcctc atgtggcatg ctttctagac tttaaaatga 120
ggtaccctga atagcactaa gtgctctgta agctcaagga atctgtgcag tgctacaaag 180
cccacaggca gagaaagaac tcctcaagtg cttgtgggtca gagactaggt tccatatgag 240
gcacacctat gatgaaggtc ttcacctcca gaagggtgaca ctgttcagag atcctcatth 300
cctggagagt gggagaaaat ccctcctttg ggaaatccct tttcccagca gcagagccca 360
cctcattgct tagtgatcat ttggaaggca ctgagagcct tcaggggctg acagcagaga 420
aatgaaaatg agtacagttc agatgggtga agaagcatgg cagtgcacac ttccatgctc 480
tttttctcag tgtctgcaac tccaaagatc aaggccataa cccaggagac catcaacgga 540
agattagttc tttgtcaagt gaatgaaatc caaaagcacg catgagacca atgaaagttt 600
ccgctgtttg taaaatctat tttcccccac ggaaagtcct tgcacagaca ccagtgcagtg 660
agttctaaaa gatacccttg gaattatcag actcagaaac ttttattttt tttttctgta 720
acagtctcac cagacttctc ataatgctct taatatattg cacttttcta atcaaaagtc 780
gagtttatga gggtaaagct ctactttcct actgcagcct tcagattctc atcattttgc 840
atctattttg tagccaataa aactccgcac tagcaaaaaa aaaaaaaaaa aa 892
```

&lt;210&gt; 188

&lt;211&gt; 1448

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;220&gt;

&lt;221&gt; misc\_feature

&lt;222&gt; 1124

&lt;223&gt; n = A,T,C or G

&lt;400&gt; 188

```
tgtgactcac atttctttta ctgtgacaca ataatgtgat cctaaaactg gcttatcctt 60
gagtgtttac aactcaaaca actttttgaa tgcagtagtt tttttttttt aaaaacaaac 120
ttttatgtca aatttttttt cttagaagta gtcttcatta ttataaattt gtacaccaa 180
aggccatggg gaactttgtg caagtacctc atcgctgagc aaatggagct tgctatgttt 240
taatttcaga aaatttcctc atatacgtag tgtgtagaat caagtctttt aataattcat 300
tttttcttca taatattttac tcaaagttaa gcttaaaaat aagttttatc ttaaaatcat 360
atttgaagac agtaagacag taaactatth taggaagtca acccccattg cactctgtgg 420
cagttattct ggtaaaaata ggcaaaagtg acctgaatct acaatgggtg cccaaagtaa 480
ccaagtaaga gagattgtaa atgataaacc gagctttaaa ggataaagtg ttaataaaga 540
aagggaagctg ggcacatgtc aaaaaggag atcgaaatgt taggtaatca tttagaaagg 600
acagaaaata tttaaagtgg ctcataggta atgaatattt ctgacttaga tgtaaatcca 660
tctggaatct ttacatcctt tgccagctga aacaagaaag tgaagggaca atgatatttc 720
atggtcagtt tattttgtaa gagacagaag aaatttatatc tatacattac cttgtagcag 780
cagtacctgg aagccccagc ccgtcacaga agtgtggagg ggggctcctg actagacaat 840
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aaatttctga ctgtcagtag aagtgcagata agatcctcag aggaaacagt aagaagggat 1080
aatcattaag atagtaaaac aggcaaagca gaatcacatg tgcncacaca catacacatg 1140
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tactaactgc cctctcccaa gattcattta gtcaaacag tatccgtaaa ctagggaataa 1260
tgccacatgc attcaatggg atcttttaag tactcttcag tttgttccaa gaaatgtgcc 1320
tactgaaatc aaattaattt gtattcaatg tgtacttcaa gactgcta at tgtttcatct 1380
gaaagcctac aatgaatcat tgttcamcct tgaaaaataa aattttgtaa atcaaaaaaa 1440
aaaaaaaaa 1448
```

&lt;210&gt; 189

<211> 460  
 <212> DNA  
 <213> Homo sapiens

<400> 189  
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 cctcctctgt cttttggaga accagggtc ttctcagggg ctctagggac tgccaggctg 120  
 tttcagccag gaaggccaaa atcaagagt agatgtagaa agttgtaaaa tagaaaaagt 180  
 ggagttgggt aatcggttgt tctttcctca catttggatg attgtcataa ggtttttagc 240  
 atgttcctcc ttttcttcac cctccccttt tttcttctat taatcaagag aaacttcaaa 300  
 gttaatggga tggtcggatc tcacaggctg agaactcgtt cacctccaag catttcatga 360  
 aaaagctgct tcttattaat cataaaaact ctcacatga tgtgaagagt ttcacaaatc 420  
 cttcaaaata aaaagtaatg acttaaaaaa aaaaaaaaaa 460

<210> 190  
 <211> 481  
 <212> DNA  
 <213> Homo sapiens

<400> 190  
 aggtgggtgga agaaactgtg gcacgaggtg actgaggtat ctgtgggagc taatcctgtc 60  
 caggtggaag taggagaatt tgatgatggt gcagaggaaa ccgaagagga ggtgggtggcg 120  
 gaaaatccct gccagaacca ccactgcaaa cacggcaagg tgtgagagct ggatgagAAC 180  
 aacacccccca tgtgcgtgtg ccaggacccc accagctgcc cagcccccat tggcgagttt 240  
 gagaaggtgt gcagcaatga caacaagacc ttcgactctt cctgccactt ctttgccaca 300  
 aagtgcacc tggagggcac caagaagggc cacaagctcc acctggacta catcgggcct 360  
 tgcaaatata tcccccttg cctggactct gagctgaccg aattccccct gcgcagtcgg 420  
 gactggctca agaacgtcct ggtcaccctg tatgagagg atgaggacaa caaccttctg 480  
 a 481

<210> 191  
 <211> 489  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> 312, 455  
 <223> n = A,T,C or G

<400> 191  
 atataaatta gactaagtgt tttcaaataa atctaaatct tcagcatgat gtgttggtga 60  
 taattggagt agatattaat taagtcccct gtataatgtt ttgtaatttt gcaaaacata 120  
 tcttgagttg ttttaacagt caaaatgttt gatattttat accagcttat gagctcaaag 180  
 tactacagca aagcctagcc tgcatatcat tcacccaaaa caaagtaata gcgcctcttt 240  
 tattattttg actgaatgtt ttatggaatt gaaagaaaca tacgttcttt tcaagacttc 300  
 ctcatgaatc tntcaattat aggaaaagtt attgtgataa aataggaaca gctgaaagat 360  
 tgattaatga actattgtta attcttccta ttttaatgaa tgacattgaa ctgaattttt 420  
 tgtctgttaa atgaacttga tagctaataa aaagncaact agccatcaaa aaaaaaaaaa 480  
 aaaaaaaaaa 489

<210> 192  
 <211> 516  
 <212> DNA  
 <213> Homo sapiens

<400> 192  
 acttcaaagc cagctgaagg aaagaggaag tgctagagag agcccccttc agtgtgcttc 60

tgactttttac	ggacttggct	tgtagaagg	ctgaaagatg	atggcaggaa	tgaaaatcca	120
gcttgatgc	atgctactcc	tggttttcag	ctcctggagt	ctgtgctcag	attcagaaga	180
ggaaatgaaa	gcattagaag	cagatttctt	gaccaatatg	catacatcaa	agattagtaa	240
agcacatgtt	ccctcttgga	agatgactct	gctaaatgtt	tgagtccttg	taaataattt	300
gaacagccca	gctgaggaaa	caggagaagt	tcatgaagag	gagcttggtg	caagaaggaa	360
cttcttactg	cttttagatgg	cttttagcttg	gaagcaatgt	tgacaatata	ccagctccac	420
aaaatctgtc	acagcagggc	ttttcaacac	tgggagttaa	tccaggaaga	tattcttgat	480
actggaaatg	acaaaaatgg	aaaggaagaa	gtcata			516

&lt;210&gt; 193

&lt;211&gt; 1409

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 193

tgattctttt	ccaaaacttt	tagccatagg	gtcttttata	gacagggata	gtaaaatgaa	60
aattgagaaa	tataagatga	aaaggaatgg	taaaaatatc	ttttaggggg	cttttaattg	120
gtgatctgaa	atcttgggag	aagctgttct	tttcaggcct	gaggtgctct	tgactgtcgc	180
ctgcgcactg	tgtaccccga	gcaacattct	aagggtgtgc	tttcgccttg	gctaactctt	240
ttgacctcat	tcttcatata	gtagtctagg	aaaaagttgc	aggtaattta	aactgtctag	300
tggtacatag	taactgaatt	tctattccta	tgagaaatga	gaattattta	tttgccatca	360
acacatttta	tactttgcat	ctccaaattt	attgcggcga	gacttgtcca	ttgtgaaagt	420
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tttcatcaga	cccagtgaag	actaaggata	gatgtttttt	aactggaggt	ctcctgataa	540
ggagaacaca	atccaccatt	gtcattttaag	taataagaca	ggaaattgac	cttgacgctt	600
tcttgttaaa	tagattttaac	aggaacatct	gcacatcttt	tttccttggtg	cactatttgt	660
ttaattgcag	tggattaata	cagcaagagt	gccacattat	aactaggcaa	ttatccattc	720
ttcaagactt	agttattgtc	acactaattg	atcgtttaag	gcataagatg	gtctagcatt	780
aggaacatgt	gaagctaata	tgctcaaaaa	gatcaacaaa	ttaatatgtt	tgctgatatt	840
tgcataaattg	gctgcaattta	tttaatgttt	aattgggttg	atcaaatgag	attcagcaat	900
tcacaagtgc	attaatataa	acagaactgg	ggcacttaaa	atgataatga	ttaacttata	960
ttgcatgttc	tcttcctttc	acttttttca	gtgtctacat	ttcagaccga	gtttgtcagc	1020
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aaacctgagc	agttcctaaa	aagatttgct	gttagaaaat	ttctttgtgg	cagtcattta	1140
ttaaggattc	aactcgtgat	acaccaaaaag	aagagttgac	ttcagagatg	tgttccatgc	1200
tctctagcac	aggaatgaat	aaattttataa	cacctgtctt	agcctttgtt	ttcaaaaagca	1260
caaaggaaaa	gtgaaaggga	aagagaaaca	agtgactgag	aagtcttggt	aaggaatcag	1320
gttttttcta	cctggtaaac	attctctatt	cttttctcaa	aagattgttg	taagaaaaaa	1380
tgtaagmcaa	aaaaaaaaaa	aaaaaaaaaa				1409

&lt;210&gt; 194

&lt;211&gt; 441

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 194

cagatttcgg	tagccatctc	cctccaaata	tgtctctttc	tgctttctta	gtgccatta	60
tttccccttc	tcctttcttc	tgtaactgcc	atctccttct	tggtcttccc	attgttcttt	120
aactggccgt	aatgtggaat	tgatatttac	attttgatac	ggtttttttc	ttggcctgtg	180
tacgggattg	cctcatttcc	tgctctgaat	tttaaaatta	gatattaaag	ctgtcatatg	240
gtttcctcac	aaaagtcaac	aaagtccaaa	caaaaatagt	ttgccgtttt	actttcatcc	300
attgaaaaag	gaaattgtgc	ctcttgacgc	ctaggcaaaag	gacatttagt	actatcgatt	360
ctttccaccc	tcacgatgac	ttgcggttct	ctctgtagaa	aagggatggc	ctaagaaata	420
caactaaaaa	aaaaaaaaaa	a				441

&lt;210&gt; 195

&lt;211&gt; 707

&lt;212&gt; DNA

<213> Homo sapiens

<400> 195

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cagaaaaata tttggaaaaa atataccact tcatagctaa gtcttacaga gaagaggatt 60
tgctaataaa acttaagttt tgaaaaattaa gatgcaggta gagcttctga actaatgccc 120
acagctccaa ggaagacatg tcctatttag ttattcaaat acaagttgag ggcattgtga 180
ttaagcaaac aatatatttg ttagaacttt gtttttaaat tactgttcct tgacattact 240
tataaagagt ctctaacttt cgatttctaa aactatgtaa tacaaaagta tagtttcccc 300
atgtgataaa aggccaatga tactgagtag gatatatgag tatcatgcta cttcattcag 360
tgtgtctgtt ttttaacta ataaggcagt ttgacagaaa ttatttcttt gggactaagg 420
tgattatcat ttttttcccc ttcaaaattg tgctttaagt gctgataacc acaggcagat 480
tgcaaaagac tgataaggca acaaaagtag agaatttttag gatcaaaggc atgtaactga 540
aaggtaaaca cagtacataa gcgacaactg gggaaggcag cagtgaaca tgtttggtgg 600
gttaagttag tcattgtaaa taagggaattt gcacatttat tttctgtcga cgcggccgcc 660
actgtgctgg atatctgcag aattccacca cactggacta gtggatc 707
```

<210> 196

<211> 552

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 61, 129, 189, 222, 241, 278, 324, 338, 363, 408, 415, 463, 483

<223> n = A,T,C or G

<400> 196

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tggccagcca gcctgatgtg gatggcttcc ttgggggtgg gcttccctca agcccgaatt 60
ngtggacatc atcaatgcca aacaatgagc cccatccatt ttccctaccc ttcctgcca 120
gccagggant aagcagccca gaagcccagt aactgccctt tccctgcata tgcttttgat 180
ggtgtcatnt gctccttcct gtggcctcat ccaaactgta tnttccttta ctgtttatat 240
nttcaccctg taatggttgg gaccaggcca atccctntc cacttactat aatggttga 300
actaaacgtc accaagggtg cttntccttg gctgaganat ggaaggcgtg gtgggatttg 360
ctnctgggtt ccctaggccc tagtgagggc agaagagaaa ccacccntc ccttnttaca 420
ccgtgaggcc aagatcccct cagaaggcag gagtgtgcc ctncccatg gtgcccgtgc 480
ctntgtgctg tgtatgtgaa ccacccatgt gaggaataa acctggcact agggaaaaaa 540
aaaaaaaaaa aa 552
```

<210> 197

<211> 449

<212> DNA

<213> Homo sapiens

<220>

<221> misc\_feature

<222> 56, 58, 76

<223> n = A,T,C or G

<400> 197

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ctccagagac aacttcgcgg tgtggtgaac tctctgagga aaaacacgtg cgtggnanca 60
agtgactgag acctanaaat ccaagcggtg gaggtcctga ggccagccta agtcgcttca 120
aatggaacg aaggcgtttg cgggggtcca ttcagagccg atacatcagc atgagtgtgt 180
ggacaagccc acggagaatt gtggagctgg cagggcagag cctgctgaag gatgaggccc 240
tggccattgc ccgccctgga gtgctgccc agggagctct tcccgcact cttcatggca 300
gcctttgacg ggagacacag ccagaccctg aaggcaatgg tgcaggcctg gcccttcacc 360
tgccctcctc tgggagtgtg gatgaaggga caacatcttc acctggagac cttcaaagct 420
gtgcttgatg gacttgatgt gtccttgc 449
```

<210> 198  
 <211> 606  
 <212> DNA  
 <213> Homo sapiens

<400> 198  
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 attgtcacac ctgggtgggg aacatgctac tggcatctaa tgcatagagg gcagtaatgc 120  
 tgctaaacat ctttcaacgc acaggacaga gccccacaaa agagaattat ctagcccca 180  
 atgtccataa cactgctgtt gagaaaacct accgcaggat cttactgggc ttcataaggta 240  
 agcttgccctt tgttctggct tctgtagata tataaaataa agacactgcc cagtcctcc 300  
 ctcaacgtcc cgagccaggg ctcaaggcaa ttccaataac agtagaatga acactaaata 360  
 ttgatttcaa aatctcagca actagaagaa tgaccaacca tcctggttgg cctgggactg 420  
 tcctagtittt agcattgaaa gtttcagggt ccaggaaagc cctcaggcct gggctgctgg 480  
 tcaccctagc agctgaggga ctcttcaata cagaattagt ctttgtgcac tggagatgaa 540  
 tatactttta tttgtaacat gtgaaaacat ctataaacat ctactgaagc ctgttcttgt 600  
 ctgcac 606

<210> 199  
 <211> 369  
 <212> DNA  
 <213> Homo sapiens

<220>  
 <221> misc\_feature  
 <222> 29, 345  
 <223> n = A,T,C or G

<400> 199  
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 gtgtgaagaa ttccagctga acaacgactg ctccctcccc gagttcattg tgaattgcac 120  
 ggtgaacgtt caagacatgt gtcagaaaga agtgatggag caaagtgccg ggatcatgta 180  
 ccgcaagtcc tgtgcatcat cagcggcctg tctcatcgcc tctgccgggt accagtcctt 240  
 ctgctoccca gggaaactga actcagtttg catcagctgc tgcaacaccc ctctttgtaa 300  
 cgggcccaagg cccaagaaaa ggggaagttc tgctcgggcc ctcangccat ggctccgcac 360  
 caccatcct 369

<210> 200  
 <211> 55  
 <212> PRT  
 <213> Homo sapiens

<400> 200  
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 His Thr Phe Glu Thr Arg Asp Leu Ser Arg Leu Ser Ser Asp Ser Gln  
 20 25 30  
 Pro Thr Ser Asn Val Ser Gln Ser Ile Ser His Lys Val Leu Ser Phe  
 35 40 45  
 Ser Gly Val Ile Val Thr Pro  
 50 55

<210> 201  
 <211> 67  
 <212> PRT  
 <213> Homo sapiens

&lt;400&gt; 201

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Met Gln Leu Leu Ser Pro Asn Thr Lys Phe Thr Ser Cys Leu Ser Arg
 1           5           10           15
Gln Arg Gly Asn Leu Val Phe Leu Gly Asp Leu Lys Gly Cys Ser Glu
           20           25           30
Leu Lys Asn Phe Gln Glu Leu Ile Asn Gln Ser Ala Leu Val His Pro
           35           40           45
Arg Val Asp Val Trp Trp Tyr Cys Gly Gly Pro Leu Leu Gly Thr Leu
           50           55           60
Pro Asn Asn
65

```

&lt;210&gt; 202

&lt;211&gt; 73

&lt;212&gt; PRT

&lt;213&gt; Homo sapiens

&lt;400&gt; 202

```

Met Thr Pro Glu Lys Leu Arg Thr Leu Cys Glu Ile Asp Trp Leu Thr
 1           5           10           15
Leu Glu Val Gly Trp Leu Ser Glu Glu Ser Leu Glu Arg Ser Leu Val
           20           25           30
Ser Lys Val Trp His Lys Val Thr Cys Lys Pro Lys His Pro Asp Gln
           35           40           45
Phe Leu Tyr Ile Asp Ser Tyr Ser Trp Phe Arg Pro Leu Pro Pro Leu
           50           55           60
Pro Thr Val Val Lys Arg Thr Ala Ala
65           70

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&lt;210&gt; 203

&lt;211&gt; 2008

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 203

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aaatggaacg aagcggtttg cggggttcca ttcagagccg atacatcagc atgagtgtgt 180
ggacaagccc acggagactt gtggagctgg cagggcagag cctgctgaag gatgaggccc 240
tgccattgac ccgcccctgga gttgctgccc agggagctct tcccgcact cttcatggca 300
gcctttgacg ggagacacag ccagaccctg aaggcaatgg tgcaggcctg gcccttcacc 360
tgctccctc tgggagtgtg gatgaaggga caacatcttc acctggagac cttcaaagct 420
gtgcttgatg gacttgatgt gtccttgcc caggaggttc gcccaggag gtggaaactt 480
caagtgtctg atttacggaa gaactctcat caggacttct ggactgtatg gtctggaac 540
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aaagtagatg gtttgagcac agaggcagag cagcccttca ttccagtaga ggtgctcgta 660
gacctgttcc tcaaggaagg tgccgtgat gaattgttct cctacctcat tgagaaagt 720
aagcgaagaa aaaaatgtact acgcctgtgc tgtaagaagc tgaagatttt tgcaatgcc 780
atgcaggata tcaagatgat cctgaaaatg gtgcagctgg actctattga agatttgga 840
gtgacttgta cctggaagct accaccttg gcgaaatttt ctcttacct gggccagatg 900
attaatctgc tagactcct cctctcccac atccatgcat ctctctacat ttccccggag 960
aaggaagagc agtatatcgc ccagttcacc tctcagttcc tcagttctgca gtgcctgca 1020
gctctctatg tggactcttt atttttctt agaggccgcc tggatcagtt gctcaggcac 1080
gtgatgaacc ccttggaac cctctcaata actaactgcc ggctttcgga aggggatgtg 1140
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atgctgaccg atgtaagtcc cgagcccctc caagctctgc tggagagagc ctctgccacc 1260
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&lt;210&gt; 204

&lt;211&gt; 923

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 204

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&lt;210&gt; 205

&lt;211&gt; 1619

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 205

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&lt;210&gt; 206

&lt;211&gt; 2364

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 206

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<210> 207

<211> 787

<212> PRT

<213> Homo sapiens

<400> 207

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			20					25					30		
Thr	Arg	Arg	Glu	Gly	Tyr	Thr	Glu	Phe	Ser	Leu	Arg	Val	Glu	Gly	Asp
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Pro	Asp	Phe	Tyr	Lys	Pro	Gly	Thr	Ser	Tyr	Arg	Val	Thr	Leu	Ser	Ala
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Ala	Pro	Pro	Ser	Tyr	Phe	Arg	Gly	Phe	Thr	Leu	Ile	Ala	Leu	Arg	Glu
65					70					75					80
Asn	Arg	Glu	Gly	Asp	Lys	Glu	Glu	Asp	His	Ala	Gly	Thr	Phe	Gln	Ile
				85					90					95	
Ile	Asp	Glu	Glu	Glu	Thr	Gln	Phe	Met	Ser	Asn	Cys	Pro	Val	Ala	Val
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Thr	Glu	Ser	Thr	Pro	Arg	Arg	Arg	Thr	Arg	Ile	Gln	Val	Phe	Trp	Ile
		115					120					125			
Ala	Pro	Pro	Ala	Gly	Thr	Gly	Cys	Val	Ile	Leu	Lys	Ala	Ser	Ile	Val
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Gln	Lys	Arg	Ile	Ile	Tyr	Phe	Gln	Asp	Glu	Gly	Ser	Leu	Thr	Lys	Lys
145					150					155					160
Leu	Cys	Glu	Gln	Asp	Ser	Thr	Phe	Asp	Gly	Val	Thr	Asp	Lys	Pro	Ile
				165					170					175	
Leu	Asp	Cys	Cys	Ala	Cys	Gly	Thr	Ala	Lys	Tyr	Arg	Leu	Thr	Phe	Tyr
		180						185					190		
Gly	Asn	Trp	Ser	Glu	Lys	Thr	His	Pro	Lys	Asp	Tyr	Pro	Arg	Arg	Ala
		195					200					205			
Asn	His	Trp	Ser	Ala	Ile	Ile	Gly	Gly	Ser	His	Ser	Lys	Asn	Tyr	Val
	210					215					220				
Leu	Trp	Glu	Tyr	Gly	Gly	Tyr	Ala	Ser	Glu	Gly	Val	Lys	Gln	Val	Ala
225					230					235					240
Glu	Leu	Gly	Ser	Pro	Val	Lys	Met	Glu	Glu	Glu	Ile	Arg	Gln	Gln	Ser
				245					250					255	
Asp	Glu	Val	Leu	Thr	Val	Ile	Lys	Ala	Lys	Ala	Gln	Trp	Pro	Ala	Trp
			260					265					270		
Gln	Pro	Leu	Asn	Val	Arg	Ala	Ala	Pro	Ser	Ala	Glu	Phe	Ser	Val	Asp
		275					280					285			
Arg	Thr	Arg	His	Leu	Met	Ser	Phe	Leu	Thr	Met	Met	Gly	Pro	Ser	Pro
	290					295					300				
Asp	Trp	Asn	Val	Gly	Leu	Ser	Ala	Glu	Asp	Leu	Cys	Thr	Lys	Glu	Cys
305					310					315					320
Gly	Trp	Val	Gln	Lys	Val	Val	Gln	Asp	Leu	Ile	Pro	Trp	Asp	Ala	Gly
			325						330					335	
Thr	Asp	Ser	Gly	Val	Thr	Tyr	Glu	Ser	Pro	Asn	Lys	Pro	Thr	Ile	Pro
			340					345					350		
Gln	Glu	Lys	Ile	Arg	Pro	Leu	Thr	Ser	Leu	Asp	His	Pro	Gln	Ser	Pro
		355					360					365			
Phe	Tyr	Asp	Pro	Glu	Gly	Gly	Ser	Ile	Thr	Gln	Val	Ala	Arg	Val	Val
	370					375					380				
Ile	Glu	Arg	Ile	Ala	Arg	Lys	Gly	Glu	Gln	Cys	Asn	Ile	Val	Pro	Asp

385		390		395		400
Asn Val Asp Asp	Ile Val Ala Asp Leu Ala Pro Glu Glu Lys Asp Glu					
	405		410			415
Asp Asp Thr Pro	Glu Thr Cys Ile Tyr Ser Asn Trp Ser Pro Trp Ser					
	420		425			430
Ala Cys Ser Ser	Ser Thr Cys Asp Lys Gly Lys Arg Met Arg Gln Arg					
	435		440			445
Met Leu Lys Ala	Gln Leu Asp Leu Ser Val Pro Cys Pro Asp Thr Gln					
	450		455			460
Asp Phe Gln Pro	Cys Met Gly Pro Gly Cys Ser Asp Glu Asp Gly Ser					
	465		470			475
Thr Cys Thr Met	Ser Glu Trp Ile Thr Trp Ser Pro Cys Ser Ile Ser					
	485		490			495
Cys Gly Met Gly	Met Arg Ser Arg Glu Arg Tyr Val Lys Gln Phe Pro					
	500		505			510
Glu Asp Gly Ser	Val Cys Thr Leu Pro Thr Glu Glu Thr Glu Lys Cys					
	515		520			525
Thr Val Asn Glu	Glu Cys Ser Pro Ser Ser Cys Leu Met Thr Glu Trp					
	530		535			540
Gly Glu Trp Asp	Glu Cys Ser Ala Thr Cys Gly Met Gly Met Lys Lys					
	545		550			555
Arg His Arg Met	Ile Lys Met Asn Pro Ala Asp Gly Ser Met Cys Lys					
	565		570			575
Ala Glu Thr Ser	Gln Ala Glu Lys Cys Met Met Pro Glu Cys His Thr					
	580		585			590
Ile Pro Cys Leu	Leu Ser Pro Trp Ser Glu Trp Ser Asp Cys Ser Val					
	595		600			605
Thr Cys Gly Lys	Gly Met Arg Thr Arg Gln Arg Met Leu Lys Ser Leu					
	610		615			620
Ala Glu Leu Gly	Asp Cys Asn Glu Asp Leu Glu Gln Val Glu Lys Cys					
	625		630			635
Met Leu Pro Glu	Cys Pro Ile Asp Cys Glu Leu Thr Glu Trp Ser Gln					
	645		650			655
Trp Ser Glu Cys	Asn Lys Ser Cys Gly Lys Gly His Val Ile Arg Thr					
	660		665			670
Arg Met Ile Gln	Met Glu Pro Gln Phe Gly Gly Ala Pro Cys Pro Glu					
	675		680			685
Thr Val Gln Arg	Lys Lys Cys Arg Ile Arg Lys Cys Leu Arg Asn Pro					
	690		695			700
Ser Ile Gln Lys	Leu Arg Trp Arg Glu Ala Arg Glu Ser Arg Arg Ser					
	705		710			715
Glu Gln Leu Lys	Glu Glu Ser Glu Gly Glu Gln Phe Pro Gly Cys Arg					
	725		730			735
Met Arg Pro Trp	Thr Ala Trp Ser Glu Cys Thr Lys Leu Cys Gly Gly					
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Gly Ile Gln Glu	Arg Tyr Met Thr Val Lys Lys Arg Phe Lys Ser Ser					
	755		760			765
Gln Phe Thr Ser	Cys Lys Asp Lys Lys Glu Ile Arg Ala Cys Asn Val					
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His Pro Cys						
785						

&lt;210&gt; 208

&lt;211&gt; 1362

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 208

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&lt;210&gt; 209

&lt;211&gt; 453

&lt;212&gt; PRT

&lt;213&gt; Homo sapiens

&lt;400&gt; 209

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Met Ala Ser Pro Ser Leu Pro Gly Ser Asp Cys Ser Gln Ile Ile Asp
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His Ser His Val Pro Glu Phe Glu Val Ala Thr Trp Ile Lys Ile Thr
20          25          30
Leu Ile Leu Val Tyr Leu Ile Ile Phe Val Met Gly Leu Leu Gly Asn
35          40          45
Ser Ala Thr Ile Arg Val Thr Gln Val Leu Gln Lys Lys Gly Tyr Leu
50          55          60
Gln Lys Glu Val Thr Asp His Met Val Ser Leu Ala Cys Ser Asp Ile
65          70          75          80
Leu Val Phe Leu Ile Gly Met Pro Met Glu Phe Tyr Ser Ile Ile Trp
85          90          95
Asn Pro Leu Thr Thr Ser Ser Tyr Thr Leu Ser Cys Lys Leu His Thr
100         105         110
Phe Leu Phe Glu Ala Cys Ser Tyr Ala Thr Leu Leu His Val Leu Thr
115         120         125
Leu Ser Phe Glu Arg Tyr Ile Ala Ile Cys His Pro Phe Arg Tyr Lys
130         135         140
Ala Val Ser Gly Pro Cys Gln Val Lys Leu Leu Ile Gly Phe Val Trp
145         150         155         160
Val Thr Ser Ala Leu Val Ala Leu Pro Leu Leu Phe Ala Met Gly Thr
165         170         175
Glu Tyr Pro Leu Val Asn Val Pro Ser His Arg Gly Leu Thr Cys Asn
180         185         190
Arg Ser Ser Thr Arg His His Glu Gln Pro Glu Thr Ser Asn Met Ser
195         200         205
Ile Cys Thr Asn Leu Ser Ser Arg Trp Thr Val Phe Gln Ser Ser Ile

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210	215	220
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	245	250
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	260	265
Glu Ser Arg Thr Ala Arg Arg Gln Thr Ile Ile Phe Leu Arg Leu Ile		270
	275	280
Val Val Thr Leu Ala Val Cys Trp Met Pro Asn Gln Ile Arg Arg Ile		285
	290	295
Met Ala Ala Ala Lys Pro Lys His Asp Trp Thr Arg Ser Tyr Phe Arg		300
305	310	315
Ala Tyr Met Ile Leu Leu Pro Phe Ser Glu Thr Phe Phe Tyr Leu Ser		320
	325	330
Ser Val Ile Asn Pro Leu Leu Tyr Thr Val Ser Ser Gln Gln Phe Arg		335
	340	345
Arg Val Phe Val Gln Val Leu Cys Arg Leu Ser Leu Gln His Ala		350
	355	360
Asn His Glu Lys Arg Leu Arg Val His Ala His Ser Thr Thr Asp Ser		365
370	375	380
Ala Arg Phe Val Gln Arg Pro Leu Leu Phe Ala Ser Arg Arg Gln Ser		385
	390	395
Ser Ala Arg Arg Thr Glu Lys Ile Phe Leu Ser Thr Phe Gln Ser Glu		400
	405	410
Ala Glu Pro Gln Ser Lys Ser Gln Ser Leu Ser Leu Glu Ser Leu Glu		415
	420	425
Pro Asn Ser Gly Ala Lys Pro Ala Asn Ser Ala Ala Glu Asn Gly Phe		430
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Gln Glu His Glu Val		445
450		

<210> 210  
 <211> 625  
 <212> DNA  
 <213> Homo sapiens

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 <221> misc\_feature  
 <222> 607  
 <223> n = A,T,C or G

<400> 210  
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 acagtgnaaa aaaaaaaaaa aaaaa 625

<210> 211  
 <211> 1619

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 211

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&lt;210&gt; 212

&lt;211&gt; 1010

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 212

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&lt;210&gt; 213

&lt;211&gt; 480

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 213

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cttgcttcca ggctttgcgc tgcaaatacca gtgctaccag tgtgaagaat tccagctgaa 360
caacgactgc tcctcccccg agttcattgt gaattgcacg gtgaacgttc aagacatgtg 420
tgagaaagaa gtgatggagc aaagtgccgg gatcatgtac cgcaagtcct gtgcatgac 480
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&lt;210&gt; 214

&lt;211&gt; 1897

&lt;212&gt; DNA

&lt;213&gt; Homo sapiens

&lt;400&gt; 214

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aatgctttag acagtgtaaa aaaaaaaaaa aaaaaaa 1897
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&lt;210&gt; 215

&lt;211&gt; 141

&lt;212&gt; PRT

&lt;213&gt; Homo sapiens

&lt;400&gt; 215

Met	Trp	Val	Leu	Gly	Ile	Ala	Ala	Thr	Phe	Cys	Gly	Leu	Phe	Leu	Leu
1				5					10					15	
Pro	Gly	Phe	Ala	Leu	Gln	Ile	Gln	Cys	Tyr	Gln	Cys	Glu	Glu	Phe	Gln
			20					25						30	
Leu	Asn	Asn	Asp	Cys	Ser	Ser	Pro	Glu	Phe	Ile	Val	Asn	Cys	Thr	Val
	35						40					45			
Asn	Val	Gln	Asp	Met	Cys	Gln	Lys	Glu	Val	Met	Glu	Gln	Ser	Ala	Gly
	50					55					60				
Ile	Met	Tyr	Arg	Lys	Ser	Cys	Ala	Ser	Ser	Ala	Ala	Cys	Leu	Ile	Ala
65				70						75				80	
Ser	Ala	Gly	Tyr	Gln	Ser	Phe	Cys	Ser	Pro	Gly	Lys	Leu	Asn	Ser	Val
			85						90					95	
Cys	Ile	Ser	Cys	Cys	Asn	Thr	Pro	Leu	Cys	Asn	Gly	Pro	Arg	Pro	Lys
			100					105					110		
Lys	Arg	Gly	Ser	Ser	Ala	Ser	Ala	Leu	Arg	Pro	Gly	Leu	Arg	Thr	Thr
	115					120						125			
Ile	Leu	Phe	Leu	Lys	Leu	Ala	Leu	Phe	Ser	Ala	His	Cys			
	130					135					140				